Design and Synthesis of

Heteroleptic Cyclometalated Iridium(III) Complexes

and

Their Applications to Material and Biological Sciences

(ヘテロレプティックなシクロメタレート型イリジウム**(**Ⅲ**)**錯体

の設計・合成と材料科学および生物科学への応用**)**

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Chapter 4 **Synthesis and Anticancer Properties of Bis- and Mono(cationic peptide) Hybrids of Bis-Heteroleptic Cyclometalated Iridium(Ⅲ) Complexes: Effect of the Number of Peptide Units on Anticancer Activity**

List of abbreviation

Chapter 1

General Introduction

General Introduction

In the past decades, a significant research effort has been focused on the synthesis and photophysical characterization of octahedral 4d 6^6 and 5d 6^6 metal complexes.¹ The typical examples of d^6 complexes are ruthenium(II) (Ru(II)) and osmium(II) (Os(II)) complexes of 2,2'-bipyridine (bpy), which have been widely used in a variety of photonic applications, including photocatalysis and photoelectrochemistry,^{2,3} because of their long excited-state lifetimes and high luminescence efficiencies. The photophysics of related tris-chelate iridium(III) (Ir(III)) complexes have also been investigated since the 1980s.⁴⁻⁶ These d⁶ Ir(III) complexes are prepared from cyclometalated ligand such as 2-phenylpyridine (ppy) and exhibit longer excited-state lifetimes, typically in the order of microseconds, and higher luminescence efficiencies [e.g., φ_{phos} (*fac*-Ir(ppy)₃) = 0.4]⁷ than those of the Ru(II) and Os(II) complexes, due to efficient intersystem crossing between the singlet and triplet excited states induced by the strong spin−orbit coupling of the Ir(III) complex core. Moreover, color diversity arises from the ligand-related excited states in Ir complexes. The energy of the lowest excited state can therefore be controlled by adjusting the energy levels of ligand orbitals through substituent effects or by the conversion of the cyclometalated ligand structures. In addition, most of cyclometalated Ir(III) complexes can be classified into IrL₃ and IrL₂A (where L is cyclometalating ligand and A is an ancillary ligand). The synthesis, structure, and photophysical properties of IrL₃ and IrL₂A of Ir complexes and their applications are yet to be studied.

Synthesis and structure of IrL³ type complexes

In 1984, Watts et al. reported the first IrL₃-type Ir complex (L: ligand), *facial*-Ir(ppy)₃ (*fac*-**1**), which was prepared in 10% yield as a by product in the reaction of ppyH (2-phenylpyridine) with hydrated IrCl₃ \cdot H₂O.^{5b} They subsequently attempted at the reaction of methyl-substituted ppy ligands, but only trace amount of the desired complex was obtained.^{5h} In 1991, they developed a new procedure for the synthesis of *fac*-IrL³ with ppy and its analogs by using Ir(acac)₃ (acac: acetylacetonate) instead of hydrated IrCl₃ (Chart 1-1). Although this method typically produced *fac*-IrL³ in 40–75%, it was not appropriate for the synthesis of other cyclometalated ligands. To solve this problem, Güdel et al. reported the reaction of a *μ*-dichloro bridged dimer complex $[\{IrL_2(\mu\text{-}Cl)\}_2]$ with an excess amount of free cyclometalated ligands (HC^AN) to obtain *fac*-Ir(C^AN)₃ complexes containing different cyclometalated ligands in high chemical yields. 6b

Chart 1-1

Meridional (abbreviated as *mer*) forms of IrL₃ complexes can also be prepared as stereoisomers of the *fac* forms (Chart 1-2). The results show that fac -IrL₃ complexes are obtained at higher temperature (e.g. 200℃), whereas *mer*-IrL³ can be obtained at lower temperature at 150℃, suggesting that *fac* isomers are thermodynamically stable and that *mer* isomers are kinetically favorable. Therefore, the *mer* isomer can be converted to the *fac* isomer by the reaction of *mer* isomers at high temperature.⁸

Chart 1-2

It should be mentioned that *mer* and *fac* isomers are different not only in their steric geometry configuration but also in Ir–N and Ir–C bond lengths. Typical bond lengths of *fac*-Ir(tpy)₃ (*fac*-**2**, tpy is 4-methylphenylpyridine) and *mer*-Ir(tpy)₃ (*mer*-**2**) are compared in Chart 1-3. In the *fac* isomer of **2**, all the Ir–C bonds are *trans* with respect to the Ir-N(pyridine) bonds, and the Ir–N bonds are *trans* with respect to the Ir-C(phenyl) bonds with almost same Ir–C and Ir–N bond lengths. Some bond lengths of *mer*-**2** are different from those of *fac*-**2**, possibly because Ir–C (or Ir–N) bonds in *mer*-**2** possess an different electronic environment from Ir–C (or Ir–N) bonds in *fac*-**2**.

Photophysical properties of IrL³ type complexes

To date, the photophysical properties of different stereoisomers of IrL³ have been examined by several research groups.^{8,9} Two principal transitions include: (1) metal-to-ligand charge transfer (MLCT), in which an electron is promoted from a metal d orbital to a vacant π^* orbital in one of the ligands and (2) ligand-centered (LC) transitions, in which an electron is promoted between π orbitals on one of the coordinated ligands. Phosphorescence in IrL₃-type complexes is induced by strong spin-orbit coupling mainly arising from a mixture of 3 LC and 3 MLCT excited states, whereas the emissive excited state has the lowest energy levels.

The lowest excited state of $IrL₃$ -type complexes can be controlled by the introduction of electron-donating or electron-withdrawing groups on the cyclometalated ligands or by changing the cyclometalated ligand units. Table 1-1 summarizes the photophysical properties of complexes 1, 2, and $Ir(F_2ppy)_3$ 3 (Chart 1-4).⁸ It is known that the energy levels of the highest occupied molecular orbital (HOMO) and the lowest unoccupied molecular orbital (LUMO) levels of these metal complexes are affected by the functionalization of ligand parts and hence photophysical properties of *mer*-IrL3-type complexes are different from those of the corresponding *fac*-forms. Generally, meridional complexes exhibit a broad and red-shifted emission with low quantum yields, which can be attributed to strong *trans* effect from phenyl groups at the opposite side.¹⁰

Compounds	$\lambda_{\rm em}$ (nm)	Quantum yield Ф	Emission liftime τ (µs)
fac 1	510	0.40	1.9
$mer-1$	512	0.036	0.15
fac ₂	510	0.50	2.0
$mer-2$	550	0.051	0.26
fac 3	468	0.43	1.6
$mer-3$	482	0.053	0.21

Table 1-1 Photophysical properties of **1**, **2**, and **3** in 2-methyltetrahydrofuran at 298K

Chart 1-4

Synthesis and structure of IrL2A type complexes

General synthetic method of IrL₂A-type complexes **5** (A: ancillary ligand) is shown in Chart 1-5, in which μ-dichloro bridged dimer complex $[\{IrL_2(\mu-Cl)\}_2]$ 4 is readily prepared from the reaction of the ligand precursor and $IrCl_3 \cdot nH_2O^{11}$ and its chloride ions can be subsequently replaced with an ancillary ligand (A) such as acac, picolinic acid, quinolinol, and so on.^{12,13,14}

The X-ray crystal structure of **6** as one of typical examples of **5** is shown in Chart 1-6, where Ir-C bonds (1.98Å in average) are shorter than those of **2** and **3**, possibly due to weak *trans* effect of the acac ligand.

Photophysical properties of IrL2A type complexes

It should be noted that the lowest triplet energy level of the ancillary ligand A is higher than that of LC and MLCT excited states in most of the IrL2A-type complexes and hence the wavelength of emission maxima of IrL₂A is determined by ³LC and ³MLCT transitions. Similar phosphorescence emission is observed in the *fac*-IrL³ containing the same cyclometalated ligand.12,15 The results of density functional theory (DFT) calculations indicate that HOMOs are located mainly at the metal center and the LUMOs are localized on the heterocyclic rings of the cyclometalated ligand parts, not at the ancillary ligand unit.

Chart 1-7 shows the emission spectra of $Ir(tpy)_{2}(pz_{2}Bpz_{2})$ (7), $Ir(tpy)_{2}(pz_{2}H)$ (8), Ir(tpy)₂(pzH)Cl (9), and Ir(tpy)₂(acac) (6), which have the tpy ligands, in solution at room temperature. It is likely that a blue-shift of the emission maximum of **7**-**9** is caused by the ancillary parts. Namely, the electron-withdrawing ligand lowers the HOMO level, resulting in the blue-shift of the emission. On the other hand, the LUMO of $Ir(tpy)_{2}A$ complexes localized on the pyridyl rings raises in LUMO level.

Chart 1-7

Purpose of this thesis

Homoleptic IrL₃ and bis-heteroleptic IrL₂A types of Ir(III) complexes have received considerable attention as phosphorescent emitters¹⁶ and have been applied in a broad research areas related to organic light emitting diodes $(OLEDs)$,¹⁷ bioimaging probes,¹⁸ oxygen sensors,¹⁹ anticancer agents,²⁰ photoredox catalysts,²¹ pH sensors,²² and so on.²³ However, the selective synthesis and photophysical properties of *tris-heteroleptic* Ir complexes represented by IrLL'A or IrLL'L'' are yet to be studied.

We have discovered the selective degradation reactions of cyclometalated Ir(III) complexes promoted by ZnX_2 (X: halide such as Cl and Br) as presented in Chapter 2 (Chart 1-8).²⁴ Mechanistic studies suggest that the selective degradation reactions is controlled by the hardness and softness of the Brønsted and Lewis acid promoters. This degradation reaction is applied to the synthesis of *tris-heteroleptic tris-cyclometalated* Ir complexes in Chapter 3

(Chart 1-9).²⁵ The selective synthesis of specific isomers among eight possible stereoisomers of Ir complexes having the same combination of three cyclometalating ligands is reported.

In this thesis, hybrid compounds of heteroleptic cyclometalated Ir(III) complexes with specific peptides were designed and synthesized for the induction of programmed cell death (PCD) in cancer cells. It is established that PCD is an essential mechanism for the control of intracellular homeostasis for cell survival, consists of apoptosis, necroptosis, paraptosis, and autophagy (Chart1-10).²⁶ Aoki et al. have reported the design and synthesis of tris-homoleptic Ir(III)-complex with cationic peptides such as GGKK and GGKKK (G: glycine and K: lysine) segments (IPHs: Ir(III)-complex peptide hybrids) that induce the paraptotic PCD in Jurkat cells

(T-lymphocyte leukemia).²⁷ These reports promoted us to study the effect of number of peptide units on anticancer activity. In Chapter 4, design and synthesis of bis- or mono-heteroleptic types of IPHs compared with tris-homoleptic types of IPHs and those of anticancer activity are reported (Chart 1-11).²⁸ The Ir(III) complex-bispeptide conjugate and the monopeptide conjugate exhibit potential anticancer activity against Jurkat cells and dead cells show a strong green emission. These peptide conjugates show more potent activity towards Jurkat cancer cells, indicative of a positive relationship between the number of the KKKGG peptide chains and hence net cationic charges and their anticancer activity (Chart 1-11). Mechanistic studies suggest that tris-homoleptic and bis-heteroleptic types of IPHs induce paraptotic cell death in Jurkat cells with the vacuolization of the cytoplasm, mitochondria, the ER, and lysosomes, as confirmed by TEM images of Jurkat cells.^{27i,k,28}

Chapter 2

Efficient Synthesis of Tris-Heteroleptic Iridium(III) Complexes Based on the Zn2+ -Promoted Degradation of Tris-Cyclometalated Iridium(III) Complexes and Their Photophysical Properties

2-1 Introduction

Chart 2-1 shows the typical synthesis of tris-heteroleptic Ir complexes **11** (IrLL'A) by the reaction of a chloro-bridged dimers 10, which are prepared from IrCl₃, and a mixture of two different cyclometalating ligands (L and L'), followed by the reaction with a corresponding ancillary ligand (A).²⁹ Because this method gives a mixture of IrLL'A with IrL₂A and IrL'₂A, their purification is usually difficult and results in low chemical yields. Baranoff and coworkers reported on the synthesis of the tris-heteroleptic Ir complex, $Ir(ppy)(F_2ppy)(acac)(ppy)$: 2-phenylpyridine, F2ppy: 2-(4',6'-difluorophenyl)pyridine and acac: acetylacetonate), by reacting an Ir(I) complex $[\{Ir(COD)(\mu\text{-}Cl)\}_2]$ complex (COD:1,5-cyclooctadiene) with ppy and F_2 ppy ligands.^{29e} Moreover, they reported that the degradation of $Ir(ppy)(F_2ppy)(acac)$ induced by HCl (2M in Et₂O) provided a heteroleptic μ -complex $[\{Ir(ppy)(F_2ppy)(\mu-Cl)\}_2]$, which is a useful intermediate for the synthesis of other tris-heteroleptic Ir complexes containing ppy and F2ppy ligands. Although this method is also accompanied by tedious purification and low product yields, it is the only reported method to date for preparing tris-heteroleptic Ir complexes.

Chart 2-1

Aoki et al. previously reported on regioselective substitution reactions (*e.g*.; halogenations, nitration, and Vilsmeier-Haack formylation) of *fac*-tris-homoleptic cyclometalated Ir complexes, $fac-Ir(ppy)$ ₃ ($fac-Ir(tpy)$ ₃ ($fac-Ir(tpy)$ ₃ ($fac-2$) (tpy: 2-(4'-tolyl)pyridine), and $fac-$ Ir(mppy)³ (*fac***-12**) (mppy: 2-(4'-methoxyphenyl)pyridine), at the 5'-position (the *p*-position with respect to the C-Ir bond) on 2-phenylpyridine ligands and their subsequent conversions to produce 13 containing a variety of functional groups (Chart 2-2).^{30,31} These findings allowed us to design and synthesize pH-responsive Ir complexes that contain basic groups, $30a,31a,c,d$ bluecolored Ir complexes^{30b} and amphiphilic Ir complexes containing cationic peptides (IPHs) that have cytotoxic activity against cancer cells.^{31b}

During these experiments, it was found that the reaction of *fac***-2** with acetyl chloride in the presence of AlCl³ (Friedel-Crafts acylation conditions) afforded the chloro-bridged Ir dimer **15a** in 68% yield, and not an acetyl derivative **14** (Chart 2-3). 30a This finding prompted us to examine the degradation reactions of tris-cyclometalated Ir complexes with a variety of Lewis acids.

Chart 2-3

In Chapter 2 of this Ph.D thesis, we report that Lewis acids such as zinc halides (ZnX_2) , AlCl₃ and TMSCl promote the degradation of tris-cyclometalated homoleptic Ir complexes including $fac-Ir(ppy)$ 3 ($fac-1$), $mer-Ir(ppy)$ 3 ($mer-1$), $fac-Ir(tpy)$ 3 ($fac-2$), $mer-Ir(tpy)$ 3 ($mer-2$), $fac-1$ Ir(mppy)3 (*fac-***12**), *fac*-Ir(Br-tpy)3 (*fac-***16**) (Br-tpy: 2-(5'-bromo-4'-tolyl)pyridine), *fac*-Ir(mpiq)³ (*fac-***17**) (mpiq: 1-(4'-methylphenyl)isoquinoline) and *mer-*Ir(mpiq)³ (*mer-***17**) to provide the corresponding halogen-bridged Ir dimers (μ-complexes) such as **15a** from *fac***-** and *mer***-2** (Charts 2-4 and 2-5). In contrast, tris-cyclometalated Ir complexes that contain cyclometalated ligands containing an electron-withdrawing group such as $fac-Ir(F_2ppy)$ ₃ (*fac*-**3)**, *fac*-Ir(NO2-tpy)3 (*fac-***18**) (NO2-tpy: 2-(5'-nitro-4'-tolyl)pyridine) and *fac***-**Ir(tfpiq)3 (*fac-***19**) (tfpiq: 1-(4'-trifluoromethylphenyl)isoquinoline) were less reactive. The μ-complex **15a** was reacted with acetylacetone (acacH) to afford $Ir(tpy)_{2}$ (acac) 6 (Chart 2-5). Because these types of Ir complexes that contain two identical ligands are readily available (for example, **6** can be synthesized from tpy via **15a**), we examined the ligand-selective degradation reactions of triscyclometalated heteroleptic Ir complexes, *fac***-20** and *mer-***20**-**24** that contaion two different ligands such as tpy, mppy, mpiq, and F2ppy (Chart 2-4) in the presence of Brønsted and Lewis acids. As a result, it was found that the reaction of $fac-20$ with $ZnBr₂$ gives the heteroleptic μ complex $[\{Ir(tpy)(F_2py)(\mu-Br)\}_2]$ as the major product due to the selective elimination of its

tpy ligand and the product, on reaction with acacH, afforded the tris-heteroleptic Ir complex Ir(tpy)(F2ppy)(acac) **25** (Chart 2-5). Moreover, novel tris-heteroleptic Ir complexes containing an ancillary ligand (*e.g.*; 8-benzenesulfonylamidoquinoline (8BSQ)) were obtained. 24 Mechanistic studies of this degradation reaction and photochemical properties of these newly synthesized tris-heteroleptic Ir complexes are also described.³² The chemical structures and numbers of Ir complexes presented in this chapter are also summarized in Chart 2-6.

Chart 2-4

mer-1: R^1 , R^2 , R^3 = H *mer*-2: R^1 = Me, R^2 = R^3 = H $fac-3$: $R^1 = F$, $R^2 = H$, $R^3 = F$ *mer*-3: $R^1 = F$, $R^2 = H$, $R^3 = F$ *fac*-16: R^1 = Me, R^2 = Br, R^3 = H *fac*-18: R^1 = Me, R^2 = NO₂, R^3 = H

fac-17 (*fac*-Ir(mpiq)₃): R^4 = Me *mer*-17 (*mer*-Ir(mpiq)₃): R^4 = Me *fac*-19 (*fac*-Ir(tfpiq)₃): $R^4 = CF_3$

mer-20: R^5 = Me, n = 2

mer-21: R^5 = Me, n = 1 *mer*-22: R^5 = OMe, n = 2

mer-23: $n = 2$ *mer*-24: $n = 1$

Ir complexes $(1-3$ and $16-24)$

 $(Ir(tpy)₂(acac))$
(from *fac*-2 and *mer*-2) (from $fac-20$ and $mer-20$)

Chart 2-6. A list of structures and numbers of Ir complexes presented in this manuscript.

2-2 Results and Discussions

2-2-1 Synthesis of Cyclometalated Ir Complexes

The substrates for the decomposition reactions, bis- or tris-cyclometalated Ir complexes, were synthesized from IrCl3∙3H2O and cyclometalating ligands. The facial (*fac*) isomers of the cyclometalated homoleptic Ir complexes, Ir(ppy)₃ (*fac***-1**), Ir(tpy)₃ (*fac***-2**), and Ir(F₂ppy)₃ (*fac***-3**) were synthesized at 200 °C and their meridional (*mer*) isomers were obtained at *ca*. 150 °C, as reported by Thompson and co-workers. 8 For example, the synthesis of *fac***-2**, *mer***-2**, *fac***-20**, *mer***-20**, *fac***-17**, and *mer***-23** is shown in Chart 2-7. Although negligible amounts of *fac*-Ir(tpy)2(F2ppy) (*fac***-20**) were obtained by Thompson's method (a mixture of chloro-bridged Ir dimer $[\{\text{Ir(tpy)}_2(\mu\text{-}Cl)\}_2]$ 15a, which was prepared from IrCl_3 ⋅3H₂O and tpy,³³ and F₂ppy in glycerol was refluxed at 200 °C) due to ligand sclambling,³⁴ it was successfully obtained from the same substrates by using AgOTf in 2-ethoxyethanol.³⁵ Meridional isomers of heteroleptic Ir complexes, *mer***-20**, *mer***-22**, and *mer***-23**, were obtained in 74%, 17% and 81% yields, respectively, from the corresponding μ-complexes and ligands and *mer***-21** and *mer***-24** were synthesized according to a method reported in a patent,³⁶ in which the μ -complex [Ir(F₂ppy)₂(μ -Cl)]² was reacted with tpy (for *mer***-21**) or mpiq (for *mer***-24**) in the presence of AgOTf and NEt3. The synthesis of *fac-* and *mer***-20** was carried out as shown in Chart 2-7.

All the Ir complexes were structurally characterized by ¹H NMR, ESI-MS and related techniques, and X-ray structure analyses were carried out for *fac***-20**, *mer***-20**, and *mer***-23** (ORTEP drawings³⁷ in Figure 2-1). As summarized in Table 2-1, selected bond lengths of fac **20** are quite similar to those of fac -Ir(tpy)₃ (fac -2) (Ir-C: 2.02 Å and Ir-N: 2.13 Å).⁸ It should be noted that the Ir-C and Ir-N bond lengths of F2ppy ligand in *mer***-20** (Ir-C: 2.07 Å and Ir-N: 2.11 Å) and *mer***-23** (Ir-C: 2.08 Å and Ir-N: 2.12 Å) are somewhat longer than those between Ir

and tpy ligands (Ir-C: 1.99, 2.04 Å and Ir-N: 2.02, 2.03 Å) and Ir and mpiq ligands (Ir-C: 1.97, 2.05 Å and Ir-N: 2.02, 2.03 Å). For these Ir complexes, the Ir-C and Ir-N bond lengths were estimated by DFT calculations and are listed in Table 2-1, because these calculated values are important in terms of the mechanism of the degradation reactions, as described below (the results of DFT calculations of other tris-cyclometalated homoleptic Ir complexes, *fac***-2**, *fac***-3,** *fac***-12**, *fac***-16**, and *fac***-18**, are shown in Figure 2-2). The experimental Ir-C and Ir-N bond lengths of *fac***-20**, *mer***-20**, and *mer***-23** are in good agreement with the calculated Ir-C and Ir-N bond lengths (DFT calculations in Table 2-1).

Figure 2-1. ORTEP drawings of single crystal structures of (a) *fac***-20**, (b) *mer***-20**, and (c) $mer-23$ with 50% probability ellipsoids. For clarity, the CH_2Cl_2 in (b) and (c) is omitted.

(a)	X-ray structure bond lengths (Å)	Calculated bond lengths (Å)
Ir- $C(1)$ (tpy)	2.01	2.04
Ir-C (2) (tpy)	1.99	2.04
Ir-C (3) (F ₂ ppy)	1.99	2.03
Ir- $N(1)$ (tpy)	2.12	2.17
Ir- $N(2)$ (tpy)	2.12	2.16
Ir-N(3) $(F_2$ ppy)	2.13	2.16
(b)	X-ray structure bond lengths (\AA)	Calculated bond lengths (Å)
Ir-C (1) (tpy)	1.99	2.02
Ir- $C(2)$ (tpy)	2.04	2.09
Ir-C(3) $(F_2$ ppy)	2.07	2.11
Ir- $N(1)$ (tpy)	2.02	2.07
Ir- $N(2)$ (tpy)	2.03	2.08
Ir-N(3) $(F_2$ ppy)	2.11	2.19
(c)	X-ray structure bond lengths (Å)	Calculated bond lengths (Å)
Ir- $C(1)$ (mpiq)	1.97	2.02
Ir- $C(2)$ (mpiq)	2.05	2.08
Ir-C(3) $(F_2$ ppy)	2.08	2.10
Ir- $N(1)$ (mpiq)	2.02	2.09
$Ir-N(2)$ (mpiq)	2.03	2.08
Ir-N(3) $(F_2$ ppy)	2.12	2.20

Table 2-1. Selected bond lengths (Å) of (a) *fac***-20**, (b) *mer***-20**, and (c) *mer***-23** in X-ray crystal structure (left) and those obtained by DFT calculations (right)

	$fac-Ir(tpy)$ ₃ $fac-2$ R^1 = Me $R^2 = H$ $R^3 = H$	$fac-Ir(Br-tpy)$ ₃ $fac-16$ R^1 = Me $R^2 = Br$ $R^3 = H$	$fac-12$ $R^1 = OMe$ $R^2 = H$ $R^3 = H$	$fac-3$ $R^1 = F$ $R^2 = H$ $R^3 = F$	<i>fac</i> -Ir(mppy) ₃ <i>fac</i> -Ir(F ₂ ppy) ₃ <i>fac</i> -Ir(NO ₂ -tpy) ₃ $fac-18$ R^1 = Me $R^2 = NO_2$ $R^3 = H$
Bond type $Ir-C$ $Ir-N$	X -ray structure Cal. 2.04 2.02 2.17 2.13	X -ray $a)$ structure Cal. 2.03 2.02 2.13 2.17	Cal. 2.03 2.15	Cal. 2.03 2.18	Cal. 2.03 2.17

Figure 2-2. Optimized structures of tris-cyclometalated Ir complexes obtained by DFT calculations (B3LYP/LanL2DZ for Ir and Br atoms and 6-31G for H, C, N, O, and F atoms). a) From Ref. 30a in the text.

2-2-2 Degradation Reaction of *fac***-Ir(tpy)³ by Brønsted or Lewis Acids**

The results for the Brønsted or Lewis acid-promoted degradation reactions of *fac*-Ir(tpy)₃ (*fac***-2**) (43 μmol) in 1,2-dichloroethane (2.0 mL) are summarized in Table 2-2. The reaction of *fac*-2 with AlCl₃ gave the corresponding chloro-bridged Ir dimer, **15a**, in 84% yield (Entry 1). It was found that ZnCl² and ZnBr² promoted the degradation of *fac*-**2** at reflux temperature to provide **15a** and the bromo-bridged Ir dimer **15b**, respectively, in reasonably good yields, while that reaction negligibly proceeds at room temperature (Entries 2-4). In Entry 3, the tpy ligand was recovered in *ca*. 30% yield, as determined by ¹H NMR (when 20-30 eq. of ZnCl₂ was used, tpy was recovered in 71-90%). Characterization of **15b** obtained in Entry 4 was carried out by ${}^{1}H$ NMR, ESI-MS, and X-ray structure analysis (See Figure 2-3). The representative parameters for the crystal structure of **15b** in Table 2-3 indicate that two Ir atoms of **15b** adopt a slightly distorted octahedral coordination geometry about the Ir(III) centers, at which two Ir-N bonds connected to the same Ir atom are in a *trans* configuration and the Ir-C

bonds have *cis* configuration. 38 The degradation reaction of *fac***-2** was not promoted by ZnO in the absence or the presence of (*n*Bu)4NBr as a bromide source (Entries 5 and 6). In Entries 7-9, the reaction of $fac-2$ with $Zn(CIO_4)_2$ or $ZnCl_2$ in EtOH and CCl_4 gave 15a in negligible or very low yields. Trimethylsilyl chloride (TMSCl) also promoted the degradation of *fac***-2** to give **15a** in 71% (Entry 10) and FeCl₃ gave the chlorinated *fac* $\text{-}2$ (*fac* $\text{-}Ir$ (Cl $\text{-}typ$) (*fac* $\text{-}27$)) and the chlorinated tpy ligand in 9% and 4% yields and other organic compounds could not be isolated (Entry 11), suggesting a possibility that *fac***-2** was decomposed after chlorination, while CuCl² and PdCl² afforded complex mixtures, the characterization of which was difficult (Entries 12 and 13).³⁹ A negligible amount of **15a** was obtained after the reaction of CdCl₂⋅2.5H₂O and Zn²⁺-1,4,7,10-tetraazacyclodecane (Zn²⁺-cyclen) complex (28a, b)^{40,41} and afforded byproducts, characterization of which was difficult (Entries 14-16). The reaction with TFA and (nBu) ₄NCl afforded **15a** in less than 42% yield, as determined by ¹H NMR (Entry 17). On the other hand, the treatment with TFA and ZnCl₂ resulted in negligible amounts of 15a (Entry 18).

Table 2-2

^{*a*} EtOH was used as the solvent. ^{*b*} CCl₄ was used as the solvent. ^{*c*} **fac-27** was obtained in 9%. ^{*d*} See Ref. 37. e A mixture contaning 15a was obtained f See Ref. 39.

Figure 2-3. ORTEP drawing of single crystal structure of **15b** with 50% probability ellipsoids.

Table 2-3 Selected bond lengths (Å) and selected bond angles (º) of **15b**.

(a)	Bond Length (A)		
	Ir-C (tpy)	2.01, 2.01, 2.02, 2.02	
	Ir-N (tpy)	2.04, 2.04, 2.05, 2.06	
	$Ir-Br$	2.62, 2.62, 2.65, 2.65	
(b)		Bond Angles (°)	
	N (tpy)-Ir-Br	91.2, 91.3, 93.4, 93.7	
	Br-Ir-Br	84.1, 84.3	
	Ir-Br-Ir	95.7, 95.8	

Baranoff and co-workers reported that HCl (2M in Et₂O) and BF₃⋅Et₂O promote the degradation of heteroleptic Ir complexes that contain an ancillary ligand, such as $Ir(F_2ppy)_2(pic)$ (pic: picolinate) **29a** or Ir(F₂ppy)₂(acac) **29b** to give the corresponding Ir dimer $[\text{Ir}(F_2ppy)_2(\mu-$ Cl)]₂ **30a** (Chart 2-8).^{29c} For comparison, the degradation of $fac-2$ by hydrochloric acid (4M HCl in 1,4-dioxane) gave **15a** in 68% yield (Entry 19 of Table 2-2), while the treatment of *fac***-**2 with 2M HCl *aq.* in THF/H₂O (2/1) was negligible, suggesting that H₃O⁺ (pK_a = -1.74) generated by levelling effect is not a sufficiently strong acid to decompose *fac***-2**. Only trace amount of **15a** was obtained in the case of BF₃⋅Et₂O and (*n*Bu)₄NCl (added as a Cl⁻ source)

(Entry 20).⁴² In this work, it was found that **29b** undergoes decomposition in the presence of ZnCl² (60 equiv.) in 1,2-dichroloethane at room temperature to give **30a** in 93% yield (Chart 2-8). These results indicate that $AICI_3$, $ZnCl_2$, $ZnBr_2$, TMSCl and $HCI/1,4$ -dioxane promote the degradation of *fac***-2** to give **15a** or **15b** in moderate good yields and that the reactivity of tris-cyclometalated Ir complexes such as *fac***-2** is different from bis-cyclometalated Ir complexes such as **29**.

Chart 2-8

2-2-3 Degradation Reaction of Tris-Cyclometalated Homoleptic Ir Complexes by ZnBr²

The reactivity of other tris-cyclometalated Ir complexes with ZnBr_2 or HCl (4M in 1,4dioxane) was examined and the results are summarized in Tables 2-4 and 2-5. The decomposition of *fac***-1**, *mer***-1**, *fac***-2**, *mer***-2** and *fac***-12** by ZnBr₂ provided the corresponding bromo-bridged Ir dimers **31b**, **15b** and **32b** as major products (Entries 1-6 in Table 2-4). The -complex **33** was obtained from *fac***-16** and ZnBr² in 25% yield after 5 h and in 83% yield after 25 h (Entries 7 and 8 in Table 2-4). It was found that *fac***-3** and *fac***-18** required longer reaction times than those of *fac***-1** and *fac***-2** (Entries 5-8 vs. Entries 1 and 3 in Table 2-4). A comparison of the reactivity between *fac*- and *mer*-forms were inconclusive (Entry 1 vs. Entry 2, Entry 3 vs. Entry 4 in Table 2-4 and Entry 1 vs. Entry 2 in Table 2-5). Interestingly, the degradation of *fac*-3 and *fac*-18 containing electron-withdrawing groups (two F and NO₂) on the phenyl rings of the ppy ligands negligibly proceeded (Entries 9 and 12 in Table 2-4), while the *mer*-form of **3** underwent decomposition to give **30b** in 55% (Entry 11). Similarly, *fac*-**19**

having CF³ groups on the phenyl rings had a lower reactivity than that of *fac*-**17** and *mer*-**17** (Entry 3 vs. Entries 1 and 2 in Table 2-5). As shown in Table 2-2, HCl (4M in 1,4-dioxane) is a stronger promoter of the degradation reaction than ZnBr2, because the reaction of *fac***-2** in the presence of HCl proceeds at room temperature, while ZnBr₂ requires a refluxing temperature (Entry 4 vs. Entry 19 in Table 2-2). It should be noted that *fac***-3** and *fac***-18** have a low reactivity in the presence of HCl/1,4-dioxane under refluxing conditions (Entries 10 and 13 in Table 2-4) and *mer***-3** is more reactive than *fac***-3** (Entry 11 vs. Entry 9 in Table 2-4).

These results indicate that electron-withdrawing groups on the phenyl ring of cyclometalating ligands stabilize the corresponding Ir complexes. From the results shown in Tables 2-4 and 2-5, the order of susceptibility to degradation for the ligands against $ZnBr₂$ is assumed to be: mpig, tpy and ppy > mppy > Br-tpy > tfpig > F_2 ppy > NO₂-tpy.
Table 2-4

15b: R^1 = Me, R^2 = R^3 = H, X = Br **30b**: $R^1 = R^3 = F$, $R^2 = H$, $X = Br$ 31b: $R^1 = R^2 = R^3 = H$, $X = Br$ **32b**: R^1 = OMe, R^2 = R^3 = H, X = Br **33b**: R^1 = Me, R^2 = Br, R^3 = H, X = Br

 a 4M HCl in 1,4-dioxane was used.

Table 2-5

 a A mixture contaning 26c was obtained.

2-2-4 Degradation of Tris-Cyclometalated Heteroleptic Ir Complexes in the Presence of Brønsted or Lewis Acids

The aforementioned findings prompted us to examine the ligand-selective degradation of heteroleptic Ir complexes such as *fac*- and *mer*-Ir(tpy)₂(F₂ppy) (*fac*- and *mer*-20) by ZnBr₂, ZnCl₂, TMSCl, AlCl₃, and HCl/1,4-dioxane) (Chart 2-9 and Table 2-6). Because it was assumed that the degradation of *fac***-20** and *mer***-20** with ZnBr² would likely provide a mixture of **15b** and **34b**, the non-selective degradation products were reacted with acacH to convert to **6** (from **15b**) and **25** (from **34b**), respectively, in order to characterize the products more accurately (Chart 2-9). As shown in Entries 1 and 2 of Table 2-6, $Ir(tpy)_{2}$ (acac) 6 and Ir(tpy)(F2ppy)(acac) **25** were obtained in 20% and 40% yields, respectively, from *fac***-20**, and the chemical yields of **6** and **25** from *mer***-20** were 13% and 63%, respectively. ¹H NMR spectra in Figures 2-4a and 2-4b support the conclusion that the main products of the reaction of *mer*-**20** with ZnCl₂ or ZnBr₂ are the corresponding heteroleptic μ -complexes $\frac{1}{\pi}$ ($\frac{1}{\pi}$ (tpy)($\frac{1}{\pi}$)(μ - X \geq 34a (X = Cl) or 34b (X = Br), respectively.⁴³ In Entry 3 of Table 2-6, the crude products containing **15a** and **34a** were reacted with acacH to afford **6** in 6% and **25** in 63% yields. Importantly, **6**, but not **25**, was obtained as a major product of the degradation of *mer***-20** with TMSCl, AlCl₃ or HCl (4M in 1,4-dioxane) (Entries 4-6 in Table 2-6). These results indicate that only ZnX_2 (X = Cl or Br) promotes the selective elimination of a tpy ligand on *mer*-20, while TMSCl, AlCl₃, and HCl selectively remove the F₂ppy ligand from the same complex.⁴⁴

Table 2-6 $\mathcal{L}^{\text{univ}}$

 a 4M HCl in 1,4-dioxane was used.

Figure 2-4. Aromatic regions of the ¹H NMR spectra (300 MHz, CDCl₃) of μ -complexes obtained by the reaction of $mer-20$ with $ZnCl₂$ and $ZnBr₂$ (a) $34a(X = Cl)$ and (b) $34b(X = Cl)$ Br).

Complex	HOMO (eV) a	HOMO (eV) b	LUMO (eV) a	LUMO (eV) b	Total Energies (a.u.) a,c	Total Energies (a.u.) b,c
$Ir(ppy)_{3}(1)$						
$fac-1$	-4.80	-4.85	-1.22	-1.22	-1540.62282	-1541.04708
$mer-1$	-4.67	-4.73	-1.28	-1.26	-1540.68201	-1541.03600
$Ir(tpy)$ ₃ (2)						
$fac-2$	-4.71	-4.77	-1.13	-1.12	-1658.61848	-1569.00703
mer ₂	-4.60	-4.66	-1.19	-1.18	-1658.60796	-1568.99514
$Ir(F_2$ ppy) ₃ (3)						
$fac-3$	-5.60	-5.42	-1.75	-1.51	-2135.98582	-2136.44555
$mer-3$	-5.52	-5.32	-1.80	-1.56	-2135.97666	-2136.43469
Ir(mpiq) ₃ (17)						
$fac-17$	\blacksquare				-2119.41655	
$mer-17$	-4.60		-1.69		-2119.40682	
$Ir(tpy)_{2}(F_{2}ppy)$ (20)						
$fac-20$	-4.98		-1.36		-1817.74168 \rightarrow	
$mer-20$	-4.84		-1.42		-1817.73163	0.01 a.u. \approx 26 kJ/mol

Table 2-7. Orbital and total energies for *fac/mer*-Ir(C^N)₃ using Density Functional Theory (DFT)

^a Calculated in this work. ^b Values were reported by Thompson and co-workers (Ref. 8). ^c 1 a.u. = 627.5 kcal/mol

The ligand-selective degradation reactions by ZnBr₂ were further examined using various *mer*-tris-cyclometalated heteroleptic Ir complexes, *mer*-Ir(F₂ppy)₂(tpy) (*mer*-21), *mer*-Ir(mppy)₂(F₂ppy) (*mer***-22**), *mer*-Ir(mpiq)₂(F₂ppy) (*mer***-23**), and *mer*-Ir(F₂ppy)₂(mpiq) (*mer***-24**) (Chart 2-10). The reaction of **mer-21**, which is composed of two F_2 ppy ligands and one tpy ligand, with ZnBr² afforded Ir(F2ppy)2(acac) **29b** in 43% yield and a trace amount of **25**, implying that the tpy ligand is selectively released from *mer***-21** under these reaction conditions. The reaction of $mer-22$ consisting of two mppy and one F_2 ppy ligands provided Ir(mppy)(F₂ppy)(acac) **35** in 54% yield and Ir(mppy)₂(acac) **36** in 13% yield, as predicted by the higher reactivity of tpy and mppy compared to that of F_2 ppy. Similar results were obtained for the reactions of **mer-23** and **mer-24** with ZnBr₂. Namely, Ir(mpiq)(F_2 ppy)(acac) 37 and Ir(mpiq)2(acac) **38** were obtained in 67% yield and trace amounts from *mer***-23**, due to the elimination of the mpiq ligand and **30b** was the major product produced from *mer***-24**. Aromatic regions of the ¹H NMR spectra of **35** and **37** (obtained from *mer***-22** and *mer***-23** as a mixture of enantiomers (Δ and Λ forms)) are shown in Figures 2-5b and 2-5c in comparison with that of **25** (Figure 2-5a).

Chart 2-10

Figure 2-5. Aromatic regions of the ¹H NMR spectra (300 MHz, CDCl3) of (a) **25**, (b) **35**, and (c) **37**.

We recently reported on the preparation of dual-emissive cyclometalated Ir complexes such as **39** and **40** containing 8BSQ ligand **41** as an ancillary ligand (Chart 2-11). 13,45 These Ir complexes exhibit two different emission peaks (dual emission) at *ca.* 475-500 nm (blue to green-color emission) and *ca*. 600 nm (red-color emission). It was concluded that the F_2 ppy unit and the deprotonated 8BSQ group in **39** and **40** contribute to the blue-color emission and red-color emission, respectively, to produce a near-to-white emission.

Chart 2-11

With the aforementioned method in hand, we synthesized the tris-heteroleptic Ir complexes **42a** and **42b** containing F₂ppy as a blue emitting ligand, tpy as a green emitting ligand, and a 8BSQ ligand 41 as a red-emitting ligand. After reacting *mer*-20 with ZnBr_2 or ZnCl_2 , the resulting μ -complexes were treated with 41 to afford 42 (a mixture of diastereomers 42a and **42b**) in 36% or 24% yields, respectively, as summarized in Table 2-8, with a trace amount of **43**. In contrast, 43 was obtained as the major product when TMSCl, AlCl₃, FeCl₃, or HCl was used (Entries 3-6). In Entry 5, treatment with FeCl₃ afforded trace amounts of 42 or 43.

As shown in Figure 2-6a, the ¹H NMR spectrum of **42** shows two sets of signals, indicating that it is composed of a mixture of two diastereomers (**42a** and **42b**) in a 1:0.4 ratio, among 8 possible diastereomers (the enantiomers of all the diastereomers are not considered). Careful separation of these diastereomers by silica gel column chromatography (hexanes/CHCl₃/THF) afforded **42a** as the major product (Figure 2-6b). Its structure was determined by X-ray single crystal analysis, as shown in Figure 2-7 and Table 2-9, in which two N atoms in tpy and F_2 ppy units connected to Ir atom exhibit *trans* configuration, N(1) of quinolone (N(3) in Figure 2-7) and $C(2)$ of tpy $(C(1)$ in Figure 2-7) are *trans*, and NSO_2Ph $(N(4)$ in Figure 2-3) and $C(2)$ of F2ppy are also *trans*. 46 To the best of our knowledge, **42a** is the first example of tris-heteroleptic Ir complexes containing a non-symmetric ancillary ligand isolated as a single stereoisomer (a mixture of Δ and Λ enantiomers).¹⁴

^{*a*} Ratios were determined by ¹H NMR. ^{*b*} 4M HCl in1,4-dioxane was used. ^{*c*} The products were identical to 43 that was prepared according to Ref. 13.

Figure 2-6. Aromatic regions of ¹H NMR spectra (300 MHz in CDCl3) of (a) a mixture of **42a** and 42b (b) 42a alone after purification.

Figure 2-7. ORTEP drawing of single crystal structure of **42a** with 50% probability ellipsoids.

	Bond lengths (A)
Ir- $C(1)$	1.99
Ir- $C(2)$	1.99
Ir- $N(1)$	2.05
Ir- $N(2)$	2.04
Ir- $N(3)$	2.13
Ir- $N(4)$	2.16

Table 2-9. Selected bond lengths (Å) of **42a**.

2-2-5 Mechanistic Study of the Degradation of Ir Complexes Promoted by Brønsted and Lewis Acids

As described above, Baranoff and co-workers reported on the degradation of heteroleptic Ir complexes containing ancillary ligands such as Ir(F2ppy)2(pic) **29a** and Ir(F2ppy)2(acac) **20b** with HCl (2M in Et₂O) and BF₃⋅Et₂O to give the corresponding Ir dimer [Ir(F₂ppy)₂(μ-Cl)]₂ **30a** (Chart 2-8). $29c$

In contrast, our findings reported herein are that: 1) AlCl₃, ZnCl₂, ZnBr₂, TMSCl and HCl/1,4-dioxane promote the degradation of *tris-cyclometalated Ir complexes* such as *fac***-2** to afford the corresponding halogen-bridged Ir dimers (Table 2-2), 2) *fac***-3**, *fac***-18** and *fac***-19** having electron-withdrawing groups confer a lower reactivity than that of *fac***-1** and *fac***-2** (Tables 2-4 and 2-5), 3) ZnX_2 (X = Cl or Br) promotes the selective elimination of tpy, mppy or mpiq ligands from *fac***-** or *mer***-20**, *mer***-22**, and *mer***-23**, respectively, and the successive reaction with acacH affords heteroleptic Ir complexes that contain three different ligands, such as **25** (Chart 2-9), **35**, and **37** (Chart 2-10), while AlCl3, TMSCl and HCl/1,4-dioxane cause the F2ppy ligand to be removed from *mer***-20** (Table 2-6), 4) the order of reactivity of the *fac*- and *mer*-forms is not conclusive, 5) the order of reactivity of acid promoters is assumed to be: HCl/1,4-dioxane, TMSCl, $AICI_3 \gg ZnX_2$ (Tables 2-2 and 2-4), and 6) the optimized structures of *mer***-21** *mer***-22**, and *mer***-24** were obtained by DFT calculations (Figure 2-8), because sufficient crystals of these Ir complexes were not obtained for X-ray crystal structure analysis. The Ir-C and Ir-N bond lengths of *fac***-20**, *mer***-20**, and *mer***-23** observed by these crystal structures and those obtained by DFT calculation are nearly identical (Table 2-1), suggesting that the calculated structures of *mer***-21**, *mer***-22**, and *mer***-24** in Figure 2-8 are sufficiently close to those for the intristic structures (crystal structures, if obtained).

Figure 2-8. Optimized structures of (a) *mer*-Ir(F_2 ppy)₂(tpy) (*mer*-21), (b) *mer*-Ir(mppy)₂(F₂ppy) (*mer***-22**), and (c) *mer*-Ir(F₂ppy)₂ (F₂ppy) (*mer***-24**) obtained by DFT calculation (B3LYP/LanL2DZ for Ir atom and 6-31G for H, C, N, O, and F atoms).

Based on the aforementioned data, we assume two mechanisms for the degradation of triscyclometalated Ir complexes (typically, *mer***-20**) according to the used acids, as shown in Chart 2-12. In hard and soft acids and bases (HSAB) theory, H^+ , Al^{3+} , and Si^{4+} are categorized as hard Lewis acids and Zn^{2+} is categorized as borderline acids.⁴⁷ When H^+ , AlCl₃ or TMSCl are used for the degradation, the nitrogen atom on the F_2 ppy ligand in the *mer*-20 reacts with H^+ , Al^{3+} (of AlCl₃), or Si⁴⁺ (of TMSCl), which are hard acids (44), resulting in the formation of the intermediate **46** via **45** (Chart 2-12a). This can be attributed to the fact that the Ir-N bond of F2ppy is longer (namely weaker) than the other Ir-N bonds between Ir and the tpy ligand (Table 2-1b). Cleavage of the Ir-C bond of **45** affords **46**, which may easily dimerize to give **15a**.

Chart 2-12

In the case of Zn^{2+} -promoted degradation reactions, on the other hand, we hypothesize an interaction between the molecular orbital of the phenyl ring of **mer-20** and that of Zn^{2+} (47 in Chart 2-12b), because Zn^{2+} is one of the borderline acids. This reaction is considered to initiate the metal exchange reaction from Ir-C to Zn-C to give the intermediate **49** via **48**, resulting in the production of **34** and tpy (30-70%) (Entries 3 and 4 in Table 2-2 and Entries 2 and 3 in Table 2-6).

The decomposition reaction of $fac-2$ with ZnX_2 ($X = C1$ or Br) was carried out in the presence of aldehyde to trap the Zn^{2+} -coordinated species such as 49. It was hypothesized that Reformatsky reagent-like species in **49** (Chart 2-12) may undergo C-C bond formation reaction with benzaldehyde, considering an analogy to the nucleophilic $C(sp^2)$ -H addition of manganase reagents to aldehyde, as reported by Wang and co-workers.⁴⁸ In a typical experiment, a solution of benzaldehyde in THF was added to a degradation reaction mixture of *fac*-2 and ZnBr₂ after cooling to room temperature. It should be noted that THF was used as a co-solvent to dissociate **49** and activate organozinc species in it. In this reaction, however, **15b** was obtained in 53% and benzaldehyde was recovered in 73%.

Next, we carried out the quenching experiments of the Zn^{2+} -promoted degradation reaction with D₂O/DCl or MeOD to check the formation of the deuterated tpy ligand. The reaction mixture of $fac - 2$ and $ZnCl_2$ was refluxed for 2 hours, to which D_2O/DCl was added at room temperature for 30 min. However, **15a** and not deuterated tpy were obtained in 82% yield and 13% yield. After the decomposition of *fac*-2 and ZnCl₂, D₂O/DCl or MeOD was added and refluxed again for 30 min. In these experiments, tpy was recovered in 30-70% without substitution with deuterium (D), though deuterated **15a** (*d12*-**15a**) was obtained only quenched by D2O/DCl. The quenching of MeOD gave **15a** and tpy in 77% and 43% yield. It should be noted that a mixture of **15a** and D₂O/DCl was refluxed in 1,2-dichloroethane for 16 hours to afford *d12*-**15a**. Subsequently, *d12*-**15a** was reacted with acac or ppy to obtain the corresponding deuterated **6** (d_6 **-6**) or *mer*-Ir(d_3 -tpy)₂(ppy). For a comparison, the ¹H NMR spectra of **15a** and d_{12} -15a are given in Figure 2-9. The treatment of 31a and 32a with D₂O/DCl in 1,2dichloroethane at reflux for *ca*. 2-6 hours also gave the corresponding deuterated **30a** and **31a** (*d4*-**31a** and *d12*-**32a**) in good yields, respectively. This deuteration mechanism is now in progress. Futhermore, the degradation of $fac-2$ with $ZnCl₂$ in 1,2-dichloroethane- d_4 $(ClCD₂CD₂Cl)$ gave tpy in 28% without deuteriumation, denying that 1,2-dichloroethane functions as a proton donor to **49**. These experimental results suggest that a very small amount

of H2O contained in ZnX² may function as a proton donor to the organozinc species in **49**. Further study is now in progress.

Figure 2-9. Aromatic regions of ¹H NMR spectra (300 MHz in CDCl3) of (a) **15a** and (b) *d12***- 15a**.

The energy levels and shapes of HOMO of *fac***-20**, *mer***-20**, and *mer***-21** are presented in Figure 2-10. The same parameters of *mer***-22**, and *mer***-23**, and *mer***-24** are shown in Figure 2-11. The successive cleavage of Ir-N may afford **49**, which could exist as a hetero μ-complex between them (**48**) and are then decomposed to tpy and **34b** after quenching.

Figure 2-10. HOMOs of tris-cyclometalated Ir complexes *fac***-20**, *mer***-20**, and *mer***-21** calculated by the Gaussian09 program using the B3LYP hybrid functional together with the LanL2DZ basis for the Ir atom and the 6-31G basis sets for the H, C, N, and F atoms.

Figure 2-11. HOMO of (a) *mer***-22**, (b) *mer***-23**, and (c) *mer***-24** calculated by the Gaussian09 program using the B3LYP hybrid functional together with the LanL2DZ basis for the Ir atom and the 6-31G basis sets for the H, C, N, O, and F atoms.

In support of the reactivity of their phenyl groups, the treatment of *mer***-20** with NBS afforded *mer***-50**, whose 5'-position is brominated (Chart 2-13), $30,31$ as confirmed by X-ray structure (Figure 2-12). Note that *mer***-50** is, to the best of our knowledge, the first example of trisheteroleptic tris-cyclometalated Ir complex that is synthesized without the formation of other diastereomers. As shown in Figure 2-3, two Ir atoms in **15a** have *N,N*-*trans* configuration and the same product **15a** is obtained from *fac***-20** and *mer***-20**, in which Ir has *N,N*-*cis* configuration and *N,N*-*trans* configuration, although this mechanism is yet to be studied.

Chart 2-13

Figure 2-12. ORTEP drawing of single crystal structure of *mer*-**50** with 50% probability ellipsoids. For clarity, the H2O was omitted. Selected bond lengths (Å) of *mer*-**50** are shown in the right part.

The data in Table 2-4 indicate, that *fac***-3** and *fac***-18**, which include the electron-withdrawing groups, have lower reactivity in the presence of ZnX_2 and HCl. For reference, it has been reported that the bonds between M and the oxygen atom of the $M(8-hydroxyquinolyl)$ ₃ complex $(M = A)(III)$ or In (III)) are stabilized by the introduction of Br and NO₂ groups on the 8hydroxyquinolyl ligand.⁴⁹ We then asked ourselves why Ir complexes having electronwithdrawing groups such as *fac***-3**, *fac***-18**, and *fac***-19** are less reactive than *fac***-1** and *fac***-2**.

It is well described that Ir complexes are one of most kinetically inert class of metal complexes⁵⁰ and the significant stability of cyclometalated Ir complexes in comparison with organometallic reagents such as organolithium and organomagnesium reagents can be explained by the difference of electronegativity of the carbon (χ_c) and the corresponding metals (χ_{Ir} , χ_{Li} , and χ_{Mg}).⁵¹ The electronegativity values for carbon and iridium are 2.55 (χ_c) and 2.20 (χ_{Ir}), respectively, from which the $\Delta \chi$ value between C and Ir is calculated to be 0.35, as displayed in Figure 2-13. This value is smaller than the corresponding values for reactive organometalic reagents like organolithium, organomagnesium, and organozinc reagents, which explains the higher stability of C-Ir bonds in Ir complexes in comparison with C-Li, C-Mg, and C-Zn bonds. It is possible that electron-withdrawing groups (R in Figure 2-13) on the phenyl rings of *fac***-3**, *mer***-3**, *fac***-18**, and *fac***-19** (Tables 2-4 and 2-5) would decrease the electronegativity of carbon atoms in Ir-C bonds indicated in Figure 2-13, resulting in a decrease in the Δχ value and the higher stability of these complexes compared to *fac***-1** and *fac***-2**.

Figure 2-13. Electronegativity values for carbon and several metals.

The HOMO and LUMO energy levels of the Ir complexes were also considered. Figure 2- 14 provides information on the HOMO and LUMO levels of the Ir complexes used in the decomposition reaction tested. In the left half of Figure 2-14, the HOMO and LUMO levels of the more reactive Ir complexes are shown and the less reactive complexes are listed in the right half.⁵² We next focused on the relationship between HOMO levels of Ir complexes and LUMO levels of ZnBr2. Among the reactive Ir complexes, *mer***-21** (*mer*-Ir(F2ppy)2(tpy)) has the most negative HOMO level (-5.27 eV) and the less reactive Ir complexes (*e.g.*; **3** and **18**) possess more negative HOMO levels (< -5.4 eV).

The LUMO level of the optimized structure of ZnBr₂ is estimated to be *ca*. -1 eV by DFT calculation (B3LYP/LanL2DZ). Thus, we assume that orbital interactions between the LUMO of ZnBr² (*ca.* -1 eV) and the HOMOs (more positive than *ca.* -5.2 eV) of the Ir complexes are important for these degradation reactions. LUMO energy level of $ZnBr₂$ (-0.99 eV) is lower than that of $ZnCl_2$ (-0.85 eV) and we assume that this is the reason why higher yield was obtained in the decomposition of $fac-2$ with $ZnBr_2$ than that with $ZnCl_2$. At the same time, ZnX_2 functions as a halide source to afford μ -complex.

Regarding the mechanism of the selective-degradation reaction of *mer*-tris-cyclometalated heteroleptic Ir complexes such as *fac*-20, *mer*-20, and *mer*-21 promoted by ZnBr₂ (Charts 2-9) and 2-10), the HOMOs of these complexes are located on the major ligand in each complex (Figure 2-10). Namely, **mer-20** and $fac-20$ contain two tpy and one F_2 ppy ligands and its HOMO is located on tpy as well as the Ir metal core. *Mer*-21 includes two F_2 ppy and one tpy groups and its HOMO is located on the F2ppy unit and the Ir core. Therefore, we assume that the HOMO on tpy ligands in 21 and that on the F_2 ppy of 21 interact with the LUMO of $ZnBr_2$, as speculated in **46** of Chart 2-12a, thus initiating the selective decomposition of the corresponding ligand (similarly, the HOMOs of *mer***-22**, *mer***-23**, and *mer***-24** are shown in Figure 2-11, in which similar behavior is observed). It should be noted that *mer***-21** and *mer***-24** are exceptions, from which the minor ligand (tpy or mpiq) is ejected faster than the major ligand (F_2ppy) .

Figure 2-14. Energy diagram for the HOMO and LUMO of tris-cyclomatalated Ir complexes $frac-2$, mer-2, fac-3, mer-3, fac-12, fac-16, fac-17, fac-18, fac-19, fac-20, and mer-20-24) ZnCl₂, and, ZnBr₂ calculated by the Gaussioan09 program using the B3LYP hybrid functional together with the LanL2DZ basis for the Ir, Zn, Cl, and Br atoms and the 6-31G basis sets for the H, C, N, O, and F atoms.

2-2-6 Photophysical Properties of the Ir Complexes

UV/vis absorption spectra of *mer***-17**, *mer***-20-24**, *fac***-20**, **6**, **25**, **29b**, and **35-38** (10 μM) in DMSO at 298 K are shown in Figure 2-15 and their photochemical properties are summarized in Table 2-10. These complexes exhibit a weak absorption in the region of *ca*. 350-500 nm, which are due to spin-allowed and spin-forbidden metal-ligand change transfer (MLCT) transitions and spin-forbidden $\pi-\pi^*$ transitions.

Figure 2-15. UV/vis spectra of Ir complexes in DMSO at 298 K ([Ir complex] = 10 μ M). (a) **6** (plain curve), **25** (bold curve), and **29b** (dashed curve). (b) **35** (bold curve) and **36** (plain curve). (c) **37** (bold curve) and **38** (plain curve). (d) *mer***-20** (bold curve), *fac***-20** (plain curve), *mer***-21** (dashed bold curve), and *mer***-22** (dashed curve). (e) *mer***-17** (bold curve), *mer***-23** (dashed curve) and *mer***-24** (plain curve).

Emission spectra of the substrates of degradation, *mer***-17**, *mer***-20-24**, *fac***-20**, and products obtained in this work, **6**, **25**, **29b**, and **35**-**38** (10 μM in degassed DMSO), were measured at 298 K (excitation at 366 nm), as shown in Figure 2-16, respectively, and their quantum yields were determined based on the Φ value of quinine sulfate in 0.1 M H₂SO₄ (Φ = 0.55) or *fac*-Ir(mpiq)₃ in toluene (Φ = 0.26) used as a standard reference (see Table 2-10).⁵³ The emission maxima of **6**, **25** and **29b** are 518, 508, and 489 nm, respectively, indicating that the emission maxima of tris-heteroleptic Ir complex **25** are located between those of **6** and **29b**.As expected, it was found that **37** exhibits a dual emission at *ca*. 493 nm and *ca*. 602 nm, respectively, because the F2ppy and mpiq ligands emit a blue and a red color, respectively. It should be noted that **37** was recrystallized twice in order to minimize contamination by **29b** and **38** and the exactly the same spectra were observed after recrystallization.

Figure 2-16. Normalized emission spectra of (a) *fac***-20** (plain curve), *mer***-20** (dashed curve), *mer***-21** (dashed bold curve) and *mer***-22** (bold curve). (b) *mer***-17** (bold curve), *mer***-23** (dashed curve) and *mer***-24** (plain curve). (c) **6** (dashed curve), **25** (bold curve), and **29b** (plain curve) in degassed DMSO at 298 K ([Ir complex] = 10 μ M and excitation at 366 nm). Inset: Photograph showing DMSO solution of 25 (10 μ M) excited by UV light at 365 nm. (d) 29b (dashed curve), **35** (bold curve) and **36** (plain curve). Inset: Photograph showing DMSO solution of $35(10 \mu M)$ excited by UV light at 365 nm . (e) $29b$ (dashed curve), 37 (bold curve) and 38 (plain curve). Inset: Photograph showing DMSO solution of 37 (10 μ M) excited by UV light at 365 nm. in degassed DMSO at 298 K ([Ir complex] = 10 μ M and excitation at 366 nm)

Table 2-10. Photophysical properties of *mer***-17**, *mer***-20-24**, *fac***-20**, **6**, **25**, **29b**, and **35-38** in degassed DMSO solutions at 298 K (excitation at 366 nm) ([Ir complex] = 10 μ M).

Compounds	λ_{abs} (nm)	$\lambda_{\rm em}$ (nm)	Quantum yield Ф	Emission liftime $\tau(\mu s)$
$mer-17$ (mer-Ir(mpiq) ₃)	303, 350, 486	614	2.0×10^{-2} ^a	0.17^{c}
<i>fac</i> -20 (<i>fac</i> -Ir(tpy) ₂ (F ₂ ppy))	281, 362	516	0.57^{b}	1.9 ^d
<i>mer</i> -20 (<i>mer</i> -Ir(tpy) ₂ (F ₂ ppy))	336, 366, 400	528	4.7×10^{-2}	0.30 ^e
<i>mer</i> -21 (<i>mer</i> -Ir(F_2 ppy) ₂ (tpy))	278, 389	504	0.31^{b}	1.0 ^d
<i>mer</i> -22 (<i>mer</i> -Ir(mppy) ₂ (F_2 ppy))	278, 409	512	8.3 x 10^{-2} ^b	0.25^{d}
<i>mer-23</i> (<i>mer-Ir</i> (mpiq) ₂ (F_2 ppy))	281, 343, 470	603	0.22^a	1.1 ^c
<i>mer</i> -24 (<i>mer</i> -Ir(F_2 ppy) ₂ (mpiq))	272, 354, 395	581	0.31^{a}	4.7 ^f
6 (Ir(tpy) ₂ (acac))	408, 462	518	0.42^{b}	1.4 ^d
25 (Ir(tpy) $(F_2$ ppy)(acac))	400, 450	508	0.51^{b}	1.3 ^d
29b $(\text{Ir(F2ppy)2(acac))}$	330, 389, 437	489	0.53^{b}	1.0 ^d
35 (Ir(mppy) $(F_2$ ppy)(acac))	283, 395, 472	507	0.52^{b}	1.3 ^d
36 (Ir(mppy) ₂ (acac))	286, 401	505	0.46^{b}	1.2^d
37 (Ir(mpiq)(F_2 ppy)(acac))	280, 340, 455	493, 602	0.38^{a}	$1.9,^{d} 2.0^{c}$
38 (Ir(mpiq) ₂ (acac))	289, 345, 474	616	0.25^{a}	1.3 ^c

^{*a*} Quantum yields were determined using *fac*-Ir(mpiq)₃ as a standard reference (Φ = 0.26 in toluene). *^b* Quantum yields were determined using quinine sulfate as a standard reference (*Φ* $= 0.55$ in 0.1 M H₂SO₄ *aq*.). ^{*c*} A 590 nm long wave pass filter was used. ^{*d*} A 475 nm long wave pass filter was used. ^e A 515 nm long wave pass filter was used. ^f A 550 nm long wave pass filter was used.

The UV/Vis absorption spectra and emission spectra of **40**, **42a**, and **43** (10 μM in degassed DMSO solutions at 298 K) are shown in Figure 2-17 and their photochemical properties are summarized in Table 2-11. These complexes also exhibited a weak absorption in the region of *ca*. 350-530 nm, which are due to spin-allowed and spin-forbidden metal-ligand charge transfer (MLCT) transitions and spin-forbidden $\pi-\pi^*$ transitions.

Since these Ir complexes contain 8BSQ ligand as an ancillary ligand, they exhibit dual emissions at *ca*. 475-525 nm (HE (high energy) emission band) and *ca*. 610 nm (LE (low energy) emission band) ($\Phi = 2.5 \times 10^{-2}$ for $42a$, 9.8×10^{-2} for 40 , and 1.5×10^{-2} for 43) (Figure 2-17b and Table 2-11). 13 The maximam of HEB of **42a** was found to be observed between those of **43** and **40**. As shown in the inset of Figure 2-17b, **42a** exhibits a pale-pink emission

color. The CIE of representative Ir complexes synthesized in this work is presented in Figure 2-18.

Figure 2-17. (a) UV/vis spectra of Ir complexes **40** (plain curve), **42a** (bold curve), and **43** (dashed bold curve) in DMSO at 298 K. (b) Normalized emission spectra of **40** (plain curve), **42a** (bold curve), **43** (dashed bold curve) in degassed DMSO at 298 K ([Ir complex] = 10 μ M and excitation at 366 nm). Inset: Photograph showing DMSO solution of $42a(10 \mu)$ excited by UV light at 365 nm.

Table 2-11. Photophysical properties of **40**, **42a**, and **43** in degassed DMSO solutions at 298 K (excitation at 366 nm) ([Ir complex] = $10 \mu M$).

Compounds	$\lambda_{\rm abs}$ (nm)	$\lambda_{\rm em}$ (nm)		Quantum yield	Emission lifetime
		HER ^a	LEB^a	$\left(\varPhi\right)$	τ (µs)
40	331, 366, 417	474, 493	613	9.8×10^{-2}	1.4, 9.3 ^b
42a	299, 381, 424	485, 515	613	2.5×10^{-2}	1.9, 13^c
43	304, 335, 386, 446	497, 525	617	1.5×10^{-2}	1.6, 10^{c}

^a HEB = Higher energy emission band, and LEB = Lower energy emission band. *b* A 435 nm long wave pass filter was used. ^{*c*} A 475 nm long wave pass filter was used.

Figure 2-18. Chromaticity diagram showing the xy color coordinates of **25**, **35**, **37**, **40**, and **42a** (CIE 1931). The xy color coordinates of these complexes were calculated from their emission spectra in degassed DMSO.

2-2-7 Mechanistic Studies of Dual Emission of Ir complexes (37 and 42a)

TD-DFT calculations were performed for 37 and 42a using the Gaussian09 program.⁵⁴ For **37**, the calculated triplet energies T_1 and T_3 are close to the experimentally determined values $(T_1: 2.06 \text{ eV} (602 \text{ nm})$ and $T_3: 2.52 \text{ eV} (493 \text{ nm})$ (Table 2-12). The HOMO and HOMO-2 of **37** are mainly localized on the Ir center and mpiq ligands and HOMO-1 is mainly localized on the Ir centers and acac ligands, as shown in Figure 2-19a. The LUMO and LUMO+1 are localized on the mpiq and F₂ppy ligands, respectively. TD-DFT calculations show that ³ML_{mpiq}CT (d(π)(Ir)→ π^* (mpiq)), ³L_{mpiq}C (π (mpiq)→ π^* (mpiq)), and ³L_{acac}L_{mpiq}CT transitions $(\pi(\text{acac}) \rightarrow \pi^*(\text{mpiq}))$ (Table 2-12) are included in the lowest-energy triplet excited state T₁. The third lowest-energy state T_3 of 37 is identical to its experimentally determined HEB, indicating that T₃ is a mixture of ³ML_{F2ppy}CT (d(π)(Ir) $\rightarrow \pi^*(F_2ppy)$), ³L_{mpiq}L_{F2ppy}CT $(\pi(mpiq) \rightarrow \pi^*(F_2ppy))$, and ${}^3L_{acac}L_{F2ppy}CT$ transitions $(\pi (acac) \rightarrow \pi^*(F_2ppy))$.

As shown in Table 2-12, The calculated triplet energies T_1 and T_2 of 42a were 1.94 eV and 2.59 eV, respectively, which are in fairly good agreement with the experimentally determined values (T₁: 2.02 eV (613 nm) and T₂: 2.56 eV (485 nm). As shown in Figure 2-19b, the HOMO of **42a** is mainly localized on the Ir center, the quinoline ring (quin), and the nitrogen atom (n) of the sulfonamide group of quinoline, and the HOMO-1 and HOMO-2 are mainly localized on the Ir centers and tpy ligands. On the other hand, its LUMO is localized on the quinoline ring, and LUMO+1 and LUMO+2 are mainly localized on the F_2 ppy and tpy ligands, respectively. The results of TD-DFT calculations suggest that the lowest-energy triplet excited state T_1 is a mixture of ³ML_{quin}CT (d(π)(Ir) $\rightarrow \pi^*$ (quin)), ³L_{quin}C (π (quin) $\rightarrow \pi^*$ (quin)), ³IL_{quin}CT (n(sulfonamide) $\rightarrow \pi^*(\text{quin})$), and ³L_{tpy}L_{quin}CT transitions ($\pi(\text{typ})\rightarrow \pi^*(\text{quin})$) (Table 2-12). Furthermore, the second lowest-energy state T_2 of **42a** is a mixture of ³ML_{tpy}CT $(d(\pi)(Ir) \rightarrow \pi^*(typ)),$ ³ML_{F2ppy}CT $(d(\pi)(Ir) \rightarrow \pi^*(F_2ppy)),$ ³L_{tpy}C $(\pi(tpy) \rightarrow \pi^*(tpy)),$ ³L_{tpy}L_{F2ppy}CT $(\pi(tpy) \rightarrow \pi^*(F_2ppy))$, ³L_{quin}L_{tpy}CT $(\pi(quin) \rightarrow \pi^*(typ))$, and ³L_{quin}L_{F2ppy}CT transitions $(\pi(\text{quin}) \rightarrow \pi^*(F^2 \text{ppy}))$. These results indicate the contribution of each of the three different ligands (F2ppy, tpy, and 8BSQ) to the triplet emission states of **42a**.

Figure 2-19. Selected molecular orbitals of (a) **37** and (b) **42a** obtained from DFT calculations at B3LYP (the LANL2DZ/6-31G level).

Table 2-12. Calculated triplet transition states and characteristics of the transitions of **37** and **42a** using TD-DFT calculation at B3LYP (the LANL2DZ/6-31G level).

^a The metal-to-ligand charge transfer, interligand charge transfer, intraligand charge transfer, and ligand centered are represented by ML_xCT , L_xL_yCT , IL_xCT , and LC_x , respectively. quin: quinoline ligand

2-3 Conclusions

In this manuscript, we report on the degradation of tris-cyclometalated Ir complexes (IrL₃, L: cyclometalating ligand) promoted by Brønsted acids such as HCl/1,4-dioxane and Lewis acids such as ZnX_2 (X = Br or Cl), TMSCl, and AlCl₃. Among the various Lewis acids, $ZnBr_2$ exhibited a good reactivity for $Ir(ppy)$ ₃ **1**, $Ir(tpy)$ ₃ **2**, and $Ir(mpy)$ ₃ **12** to provide the corresponding halogen-bridged Ir dimers (μ -complexes). It was also found that the reactivity of tris-cyclometalated Ir complexes containing electron-withdrawing groups such as fluorine atoms, nitro or CF³ groups on the ligands is quite low. This reaction was applied to the selective degradation of *mer*-tris-cyclometalated Ir complexes such as *mer*-Ir(tpy)₂(F₂ppy) (*mer*-20). The reaction of *mer***-20** with ZnBr² gave a mixture of a halogen-bridged Ir dimer **15b** and **34b**, and the tris-heteroleptic Ir complex **25** (IrLL'A, A: ancillary ligand) was then obtained by treatment with acetylacetone. Furthermore, we successfully isolated a novel tris-heteroleptic Ir complex **42a** having a non-symmetric ancillary ligand after careful separation of its diastereomers. Mechanistic studies suggest that the formation of different products from some Ir complexes by H^+ , TMSCl, and AlCl₃ and ZnX_2 is due to the hardness and softness of these Brønsted and Lewis acids used. The reactivity and selectivity in the Z_1X_2 -promoted degradation can be explained by the interaction of the HOMO of Ir complexes with the LUMO of ZnX² and by the difference of electronegativity of iridium and carbon in the ligand part. Further mechanistic study is now underway. Such selective degradation reactions of triscyclometalated Ir complexes represents a potentially useful synthetic method for preparing a variety of metal complexes.

Chapter 3

Stereospecific Synthesis of

Tris-Heteroleptic Tris-Cyclometalated **Iridium(III) Complexes**

via Different Heteroleptic Halogen-Bridged Iridium(III) Dimers

3-1 Introduction

To date, numerous examples of Ir complexes have been prepared and most of them can be categorized as IrL₃, IrL₂L', IrLL'L'', IrL₂A, IL₂A', IrLL'A, and IrLL'A', as shown in Chart 3-1, where L, L', and L'' are different cyclometalating ligands that chelate to the Ir ion via carbon and hetero atoms, A depicts a symmetric ancillary ligand that binds to Ir via two hetero atoms and A' is a nonsymmetric ancillary ligand (Chart 3-1).

Chart 3-1

Tris-heteroleptic Ir complexes comprising an Ir ion with three different cyclometalating ligands, IrLL'L'', or a combination of two different cyclometalating ligands and one ancillary ligand, IrLL'A or IrLL'A', represent new classes of highly-functionalized Ir complexes, which may open new avenues for the fine-tuning of their photochemical and electrochemical properties and fill a function gap between a series of the homoleptic and bis-heteroleptic Ir complexes.

Representative synthetic methods of IrLL'A and IrLL'A' are shown in Chart 3-2, in which A is a symmetric ancillary ligand such as acacH, bipyridine (bpy), and 1,10-phenanthroline (phen) ligands, and A' presents nonsymmetric ancillary ligands such as picolinate (pic). The synthesis depicted in the left part of Chart 3-2 (Method 1) involves a reaction of a mixture of two different cyclometalating ligands (1:1) with Ir sources such as $[\text{Ir(COD)}(\mu\text{-}Cl)]_2$ or IrCl3·*n*H2O, resulting in the production of the inseparable chloro-bridged Ir dimers **51**, which are converted to a mixture of the corresponding tris-heteroleptic Ir complexes **52** having an ancillary ligand for its purification.^{29,55} As an example of this method, Baranof and co-workers synthesized Ir(ppy)(F2ppy)(acac) **60** (corresponds to **55** in Chart 3-2) from a mixture of 2-(4',6' difluorophenyl)pyridine **58** (F2ppyH) and 2-phenylpyridine **59** (ppyH) (Chart 3-3).29c Furthermore, they carried out the conversion of **60** to Ir(ppy)(F2ppy)(pic) **61a** and **61b** (corresponds to the conversion of 52 to 56 via 53 in Chart 3-2),⁵⁶ while $61a$ and $61b$ exhibit insufficient stability for device applications.⁵⁷

Chart 3-2

The reports on the synthesis of tris-heteroleptic *tris-cyclometalated* Ir complexes represented by IrLL'L'' (54 in Chart 3-2) are still so limited.⁵⁸ Park and co-workers reacted Ir(acac)₃ with a mixture of **58**, **59**, and 1-(4'-fluorophenyl)isoquinoline **62** (F-piqH) in glycerol to afford a diastereomer mixture of Ir(F_2 ppy)(ppy)(F -piq) 63 as shown in Method A of Chart 3-4.^{58a} Lepeltier and Dumur *et al.* reported on the synthesis of $Ir(tpy)(F_2ppy)(ppy)$ 64 (Method B in Chart 3-4).^{58b} In this case, $Ir(tpy)(F_2ppy)(acac)$ 25 was prepared by a reaction of a mixture of **58** and 2-(4'-tolyl) pyridine **63** (tpyH) with IrCl₃ and the successive reaction with acacH and then 25 was converted to heteroleptic chloro-bridged Ir dimer $[\text{Ir}(\text{tpy})(F_2\text{ppy})(\mu-C)]_2$ 34a (corresponds to **53** in Chart 3-2) by the partial decomposition (the release of acac part) by HCl in Et2O. The reaction of **34a** and **59** afforded a diastereomer mixture of **64** (corresponds to **54** in Chart 3-2). To the best of our knowledge, there is no report on the selective synthesis of one or several stereoisomers of tris-heteroleptic tris-cyclometalated Ir complexes, to date. Therefore, the comparison of chemical and photophysical properties of diastereomers of the Ir complexes that have the same combination of three ligands has been yet to be studied.

Quite recently, we have reported on the decomposition reactions of Ir complexes, as shown in Chart 3-5.²⁴ Typically, the reaction of tris-homoleptic Ir complex Ir(tpy)₃ 2 in the presence of Brønsted and Lewis acids such as HCl (in 1,4-dioxane) and ZnX_2 (X = Cl or Br) afforded the corresponding halogen-bridged Ir dimers (μ -complexes) **15a,b**.²⁴ It was also found that triscyclometalated Ir complexes containing electron-withdrawing groups such as fluorine, nitro, and CF³ moieties on the ligand parts are less reactive than **2**. This different reactivity of Ir complexes was applied to the selective degradation of bis-heteroleptic tris-cyclometalated Ir complexes such as Ir(tpy)2(F2ppy) **20** to afford the corresponding heteroleptic μ-complexes **34a,b** (corresponds to the conversion of **57** to **53** in Method 2 of Chart 3-2). The treatment of **34a,b** with acacH and 8-benzenesulfonylamidoquinoline **41** (8BSQ) gave **25** (corresponds to **55** in Chart 3-2) and Ir(tpy)(F2ppy)(8BSQ-) **42a** (corresponds to **56** in Chart 3-2), respectively. The selective elimination of 1-(4'-methylphenyl)isoquinoline unit **65** (mpiqH) from Ir(mpiq)₂(F₂ppy) **23** gave **67a,b**, which were reacted with acacH to give Ir(mpiq)(F₂ppy)(acac) **37**. Interestingly, it was discovered that **25** exhibits single broad emission, while **37** and **42a** have dual emission.

Herein, we report on the stereospecific synthesis and isolation of a *tris-heteroleptic triscyclometalated* Ir complex via different heteroleptic halogen-bridged Ir dimers based on the aforementioned Zn^{2+} -promoted selective degradation reactions of Ir complexes (in a dashed area of Chart 3-5).²⁵ Namely, 20 and 23 were degraded by ZnX₂ to obtain the corresponding halogen-bridged Ir dimers **34** and **67**, respectively. It was expected that the reaction of **34** with **65** and the reaction of **67** with **63** would afford the same product, $\text{Ir}(\text{tpy})(\text{F}_2\text{ppy})(\text{mpiq})$ **66**. Interesting and important findings of this manuscript are that: i) different synthetic routes afford different diastereomer pairs of **66** (**66a-d** were obtained, as described below). ii) different diastereomers of **66** have different stability. iii) different diastereomers of **66** exhibits different photophysical properties. In addition, finding of strong emission from µ-complex **67a** is also reported.

3-2 Results and Discussions

3-2-1 Synthesis of Tris-Heteroleptic Tris-Cyclometalated Ir Complexes.

As shown in Chart 3-5, halogen-bridged Ir dimers **34b** and **67a,b** were prepared as substrates for the synthesis of 66 (by Method 2 in Chart 3-2). A mixture of 20 or 23 with ZnX_2 in 1,2dichloroethane was refluxed to furnish heteroleptic µ-complexes **34b** or **67a,b**. It should be noted that **67a,b** was obtained only by Method 2 in Chart 3-2, though **34** can be obtained by both methods in Chart 3-2 (Chart 3-6 and Chart 3-7).^{58b} The structure of 34b was confirmed by ¹H NMR, ESI-MS, and X-ray structure analysis. The ORTEP drawing³⁷ of 34b is presented in Figure 3-1a and the representative parameters for its crystal structure are summarized in Table 3-1. The stereochemistry of **34b** is important to discuss the stereochemistry of the reaction of **34b** with the third ligand (mpiq in that case). The comparison of the structure of **34b** with that of **15b** ($[Ir(tpy)_2(\mu-Br)]_2)^{24}$ (Figure 3-1b and Table 3-1) indicates that two Ir atoms of both µ-complexes have pseudo-octahedral 6-coordinated structure and two Ir-N bonds connected to the same Ir atom are in a *trans* configuration and the two Ir-C bonds have *cis* configuration.²⁴ The two Ir centers have the same absolute stereochemistry, Δ and Δ or Λ and Λ , as previously described by Watts and Garces.⁵ⁱ

Figure 3-1. (a) ORTEP drawing with 50% probability ellipsoids and the stereochemical presentation of 34b. For clarity, CH₂Cl₂ and H₂O were omitted. (b) stereochemical presentation of **15b**.

Table 3-1 (a) Selected bond lengths (Å) and (b) selected bond angles (º) of **34b** and **15b**.

As shown in left part of Chart 3-8, a reaction of **34b** with **65** in 1,2-dichloroethane in the presence of AgOTf and Et₃N at 80 $^{\circ}$ C afforded a 1:0.4 mixture of two diastereomers of Ir(tpy)(F2ppy)(mpiq) **66** (Figure 3-2a) among its eight possible diastereomers, **66a-h** listed in Chart 3-8 (the enantiomers of all the diastereomers are not considered). Careful separation of **66a,b** by silica gel column chromatography (hexanes/CHCl3) afforded **66a** as a major product (Figure 3-2b). It was not possible to isolate **66b**, possibly due to its instability under the condition silica gel column chromatography. Other four diastereomers shown in the bottom of Chart 3-8 (in a dashed area) have not been obtained yet.

Figure 3-2 Aromatic regions of ¹H NMR spectra (300 MHz in CDCl₃) of (a) a mixture of **66a** and **66b**, (b) **66a** after purification, (c) a mixture of **66c** and **66d**, and (d) **66c** after isomerization.

On the contrary, the reaction of **67b** with **63** provided a mixture of two diastereomers of **66** (**66c** and **66d**), which are different from **66a** and **66b**, in a 1:0.7 ratio (right part of Chart 3-8). The comparison of aromatic regions of the ${}^{1}H$ NMR shown in Figure 3-2 suggests that these two diastereomer pair of **66a,b** and **66c,d** are completely different (Figure 3-2a and 3-2c). The diastereomer mixture of **66c** and **66d** could not be separated by purification using silica gel column chromatography, but the ratio of **66d** against **66c** was somewhat decreased after purification. Because we suspected these changes might be caused by silica gel, the isomerization of a mixture of **66a,b** and a mixture of **66c,d** in 1,2-dichloroethane was carried out in the presence of silica gel at 80 °C (Chart 3-9). The treatment of a mixture of **66a** and

66b with silica gel resulted in the decomposition and the product could not be isolated. On the other hand, the single isomer **66c** was obtained in 50% yield from a mixture of **66c** and **66d**. Note that negligible change was observed when two mixtures of **66a,b** and **66c,d** were warmed at reflux temperature in 1,2-dichloroethane without silica gel. Aromatic regions of the ¹H NMR spectrum of **66c** is shown in Figure 3-2d and its structure was determined by X-ray diffraction (Figure 3-3). These results strongly suggest that different stereoisomers of **66** (at least, **66a-h**) have different stability and that **66c** is the most stable isomer among **66a-d**, as calculated by DFT calculation (Chart 3-10).

Figure 3-3. ORTEP drawing of single crystal structure of **66c** with 50% probability ellipsoids.

For structural comparison with Ir(tpy)(F2ppy)(mpiq) **66c**, X-ray crystal structure of Ir(F_2 ppy)₂(mpiq) *mer***-24**, prepared in our previous work, was determined in this paper, as shown in Figure 3-4. The Ir-C and Ir-N bond lengths for **66c** and *mer***-24** are presented in Table 3-2. The difference of **66c** and *mer***-24** is that contains one tpy ligand instead of one of two F₂ppy ligands in *mer*-24. The bond lengths of Ir-C(n) and Ir-N(n) (n = $1 \sim 3$) in 66c are quite similar to the corresponding pair of *mer***-24**. It is interesting that the Ir-C and Ir-N bond lengths of mpiq ligand in **66c** (Ir-C(1) (mpiq): 1.99 Å, Ir-N(1) (mpiq): 2.04 Å) are *ca*. 0.1 Å shorter than those of *mer***-24** (Ir-C(3) (mpiq): 2.08 Å, Ir-N(3) (mpiq): 2.14 Å), which might be resulted from weaker *trans* influences of N(3) atom in tpy and N(2) atom in F₂ppy than those of *mer*-24.⁸ We assume that this point is important for the explanation of the different photophysical properties of **66c** and *mer***-24**, as described below.

Figure 3-4. Stereochemical presentation and ORTEP drawing of single crystal structure of *mer***-24** with 50% probability ellipsoids.

Table 3-2. Selected bond lengths (Å) in X-ray crystal structure of (a) **66c** and (b) *mer***-24**.

(a)	X-ray structure bond lengths (Å)		X-ray structure bond lengths (\AA)
Ir-C (1) (mpiq)	1.99		Ir- $C(3)$ (mpiq) 2.08
Ir-C(2) (F_2ppy)	2.05		Ir-C(1) $(F_2$ ppy) 1.99
Ir- $C(3)$ (tpy)	2.07		Ir-C (2) (F ₂ ppy) 2.06
Ir- $N(1)$ (mpiq)	2.04		Ir- $N(3)$ (mpiq) 2.14
Ir-N(2) (F_2 ppy)	2.04		Ir-N(1) $(F_2$ ppy) 2.03
Ir- $N(3)$ (tpy)	2.15		Ir-N(2) $(F_2$ ppy) 2.04

3-2-2 Photophysical Properties of Tris-Heteroleptic Tris-Cyclometalated Ir Complexes

UV/Vis absorption spectra of **66a**, **66c**, and *mer***-24** (10 μM in DMSO solutions at 298 K) are shown in Figure 3-5a and their photochemical properties are summarized in Table 3-3. These complexes also exhibited a weak absorption in the region of *ca*. 330-500 nm, which are due to spin-allowed and spin-forbidden metal-ligand charge transfer (MLCT) transitions and spinforbidden $\pi-\pi^*$ transitions.

Emission spectra of **66a**, **66c**, and *mer***-24** (10 μM in degassed DMSO) at 298 K (excitation at 366 nm) are shown in Figure 3-5b and their quantum yields were determined based on the Φ value of *fac*-Ir(mpiq)₃ in toluene (Φ = 0.26) used as a standard reference (See Table 3-3).⁵⁹ Most interestingly, **66a** emits a broad single emission (586 nm) and the spectrum of **66c** presents dual emission at *ca*. 509 nm and *ca*. 600 nm despite **66a** and **66c** have the same three ligands. It should be noted that **66a** and **66c** was recrystallized twice in order to minimize contamination and the exactly the same spectra were observed after recrystallization.

Table 3-3. Photophysical properties of **66a**, **66c**, and *mer***-24** (in DMSO solutions) at 298 K (excitation at 366 nm) ($[66a, 66c, and *mer*-24] = 10 \mu M$).

Compounds	$\lambda_{\rm abs}$ (nm)	$\lambda_{\rm em}$ (nm)	Quantum yield ϕ^\flat	Emission liftime τ (µs)
66a (Ir(tpy)(F_2 ppy)(mpiq))	313, 356, 410	586	0.25	2.4 ^c
66c $(\text{Ir(tpy})(F_2 \text{ppy})(\text{mpiq}))$	276, 342, 411	509,600	0.30	1.5 ^d
$(mer-Ir(F2ppy)2(mpiq))^a$ $mer-24$	272, 354, 395	581	0.31	4.7 ^c

^{*a*} Data have been already reported in Ref 57 (Chapter 2). ^{*b*} Quantum yields were determined using *fac*-Ir(mpiq)₃ as a standard reference (Φ = 0.26 in toluene). ^{*c*} A 590 nm long wave pass filter was used. ΔA 475 nm long wave pass filter was used.

Figure 3-5. (a) UV/vis spectra of Ir complexes **66a** (bold curve), **66c** (dashed bold curve), and *mer***-24** (plain curve) in DMSO at 298 K. (b) Normalized emission spectra of **66a** (bold curve), **66c** (dashed bold curve), and *mer***-24** (plain curve) in degassed DMSO at 298 K ([Ir complex] $= 10 \mu M$ and excitation at 366 nm). Inset: Photograph showing DMSO solution of 66a and 66c (10 μ M) excited by UV light at 365 nm.

3-2-3 Mechanistic Study for Stereoselective Reaction from Heteroleptic Halogen-Bridged Ir Complexes to Tris-Heteroleptic Tris-Cyclometalated Ir Complexes

We asked ourselves why only two stereoisomers of tris-heteroleptic tris-cyclometalated Ir complexes are formed from two heteroleptic µ-complexes **34** and **67**, respectively? To answer this question, we tried the crystallization of **34a,b** and **67a,b** to determine their stereochemistry.

Based on the stereochemistry of **34b**, the reaction of **34** with **65** and **67** with **63** are shown in Chart 3-11, especially focused on the attack of **65** and **63** to **34** and **67**. Namely, we assume that the *cis*-*C*,*C*- or *trans*-*N*,*N*-configurations of **34** and **67** found in its X-ray single crystal structure (Figure 3-6a) are retained in **66a,b** and **66c,d**, and that **65** and **63** attack to Ir atom of **34** and **67** in two different directions (**68** and **69** in Chart 3-11). In support of our mechanism, it should be noted that the synthesis of meridional form of **2,** which was synthesized from **15a** 60 and its stereochemistry have been already discussed by Thompson and co-workers.⁸ As shown in Chart 3-12, they referred that the *trans*-*N,N*-configurations of **15a** are retained when the cyclometallation by **63** was carried out even at 140-145 °C (glycerol was used as the solvent), which is *ca*. 60 °C higher than our synthetic method (direction of the attack of **63** is indicated in Chart 3-12).

Different combinations of diastereomer of 66 are formed.

Chart 3-12

3-2-4 DFT Calculation Study for Emission of Tris-Heteroleptic Tris-Cyclometalated Ir Complexes

TD-DFT calculations of **66a** and **66c** were performed by the Gaussian09 program.⁵⁴ The selected orbitals of **66a** and **66c** are shown in Table 3-4 and Figure 3-6. For **66a**, the calculated triplet energies T_1 is in good agreement with the experimentally determined values (T₁: 2.22 eV (559 nm)) (Table 3-4). The HOMO-1 and HOMO-2 of **66a** are mainly localized on the Ir center and mpiq ligands as shown in Figure 3-6a, respectively. The LUMO is localized on the mpiq. TD-DFT calculations show that ³ML_{mpiq}CT (d(π)(Ir) $\rightarrow \pi^*$ (mpiq)) and ³L_{mpiq}C $(\pi(mpiq) \rightarrow \pi^*(mpiq))$ (Table 3-4) are included in the lowest-energy triplet excited state T₁. Because the emission of cyclometalated Ir complexes generally come from the gap of HOMO-LUMO,⁶¹ this result shows the strong contribution of mpiq ligand to the emission of 66a. Although HOMO-LUMO gap is presented in T_2 state, the experimental result is negligibly matched to the T_2 state.

The calculated triplet energies T_1 and T_2 of 66c was 2.14 eV and 2.44, respectively. As shown in Figure 3-6b, the HOMO of **66c** are mainly localized on the Ir center and mpiq and its LUMO is mainly included in mpiq, respectively. The results of TD-DFT calculations suggest that the lowest-energy triplet excited state T₁ is a mixture of ³ML_{mpiq}CT (d(π)(Ir) $\rightarrow \pi^*$ (mpiq)) and ³L_{mpiq}C (π (mpiq) $\rightarrow \pi$ ^{*}(mpiq)) (Table 3-4). In contrast with **66a**, the T₂ of **66c** is in good accordance with experimental state. Based on the TD-DFT calculations, T₂ of 66c was mainly composed of ³ML_{mpiq}CT (d(π)(Ir) $\rightarrow \pi^*(mpiq)$) and ³L_{tpy}L_{mpiq}CT transitions (π (tpy) $\rightarrow \pi^*(mpiq)$), respectively. These results indicate the tpy in **66c** is strongly contributed to the experimental determined high energy band.

Figure 3-6. Selected molecular orbitals of (a) **66a** and (b) **66c** obtained from DFT calculations at B3LYP (the LANL2DZ/6-31G level). Color code: Ir = green, $C =$ gray, $H =$ white, $N =$ blue, $F = pink$.

Table 3-4. Calculated triplet transition states and characteristics of the transitions of **66a** and **66c** using TD-DFT calculation at B3LYP (the LANL2DZ/6-31G level).

Compound	λ_{em} (nm) Exp.	E(eV) Exp.	E(eV) TD-DFT	State	Assignment	Main transition character ^{a}
66a	586	2.12	2.22	T_{1}	$HOMO-1 \rightarrow LUMO (46%)$ HOMO-2 \rightarrow LUMO (22%)	${}^{3}\text{ML}_{\text{mpiq}}\text{CT}+{}^{3}\text{L}_{\text{mpiq}}\text{C}+{}^{3}\text{IL}_{\text{mpiq}}\text{CT}$
	$\qquad \qquad$	\sim	2.40	T_{2}	$HOMO \rightarrow LUMO (82%)$	${}^{3}\text{ML}_{\text{mpiq}}\text{CT}+{}^{3}\text{L}_{\text{typ}}\text{L}_{\text{mpiq}}\text{CT}$
66c	600	2.06	2.14	$\rm T_1$	$HOMO \rightarrow LUMO (63%)$	${}^{3}\text{ML}_{\text{mpiq}}\text{CT}+{}^{3}\text{L}_{\text{mpiq}}\text{C}+{}^{3}\text{IL}_{\text{mpiq}}\text{CT}$
	509	2.44	2.44	T ₂	HOMO-1 \rightarrow LUMO (56%) $HOMO \rightarrow LUMO (26%)$	${}^{3}\text{ML}_{\text{mpiq}}\text{CT}+{}^{3}\text{L}_{\text{typy}}\text{L}_{\text{mpiq}}\text{CT}$

^a The metal-to-ligand charge transfer, interligand charge transfer, intraligand charge transfer, and ligand centered are represented by ML_xCT , L_xL_yCT , IL_xCT , and LC_x , respectively.

Localization of HOMO at the mpiq unit of **66c** was checked by the electrophilic substitution reaction. Previously, we found the ligand-selective and regioselective electrophilic reactions of heteroleptic Ir complexes.14,24 For example, electrophilic reaction of **20** proceeds at the *p*position of the phenyl group of a cyclometalating ligand that is *trans* to in Ir-N bond. Namely, the bromination of 20 was carried out with NBS to give $Ir(tpy)(Br-tpy)(F_2ppy)$ *mer*-50 in 64% yield (Chart 3-13). It was assumed that **66c** was also brominated to afford Ir(tpy)(F₂ppy)(Brmpiq) in 87% yield, confirming the high reactivity of the HOMO speculated by DFT calculations.

3-2-5 Photophysical Properties of Heteroleptic µ-Complexes

Finally, the unexpected finding on the strong emission from heteroleptic µ-complex is **67a** is described. The UV/Vis absorption spectra for $67a$, $[Ir(mpiq)_2(\mu-C1)]_2$ 26a, and $[Ir(F_2ppy)_2(\mu-C1)]_2$ Cl)]₂ **30a** (5 μ M in CH₂Cl₂ solutions at 298 K) were recorded in aerated DCM at 298 K and are shown in Figure 3-7a. The results of their photophysical properties are summarized in Table 3-5. All µ-complexes possess intense absorption bands below *ca*. 350 nm corresponding to spin-allowed singlet ligand-centered (¹LC) $\pi-\pi^*$ transitions.^{35a} These complexes also exhibited a weak absorption in the region of *ca*. 330-500 nm, which are due to spin-allowed and spin-forbidden metal-ligand charge transfer (MLCT) transitions and spin-forbidden $\pi-\pi^*$ transitions.

Emission spectra of $67a$ and $26a$ (5 μ M in degassed CH₂Cl₂ solutions) at 298 K are shown in Figure 3-7b. We found that strong emission from **67a**, as shown in Figure 3-7b, whose color is almost the same as that of *mer***-21**. The emission spectra of **67a** is broad including a low energy emission shoulder, possibly due to a mix of ${}^{3}\text{MLCT}/{}^{3}\text{LC}$ transition.^{35a}

Figure 3-7. (a) UV/vis spectra of Ir complexes **67a** (bold curve), **26a** (plain curve), and **30a** (dashed curve) in CH2Cl² at 298 K. (b) Normalized emission spectra of **67a** (bold curve) and **26a** (plain curve) in degassed CH₂Cl₂ at 298 K ([Ir complex] = 5 μ M and excitation at 366 nm). Inset: Photograph showing CH_2Cl_2 solution of $67a$ (5 μ M) excited by UV light at 365 nm.

Table 3-5. Photophysical properties of $67a$, $26a$, and $30a$ (in CH₂Cl₂ solutions) at 298 K (excitation at 366 nm) ($[67a, 26a,$ and $30a] = 5 \mu M$).

Compounds	$\lambda_{\rm abs}$ (nm)	$\lambda_{\rm em}$ (nm)	Quantum yield ϕ^{\flat}	Emission liftime τ (µs)
67a [Ir(mpiq)(F ₂ ppy)(μ -Cl)] ₂	284, 356, 437	582	0.17	3.3 ^c
$[Ir(mpiq)_{2}(\mu-Cl)]_{2}$ 26a	296, 359, 453	594	0.04	0.9 ^c
30a $[\text{Ir(F2ppy)2(\mu-Cl)]2a$	303, 339, 386	too week	\sim	$\overline{}$

a Emission too weak to determine *Φ* or *τ*. *b*Quantum yields were determined using *fac*-Ir(mpiq)₃ as a standard reference (*Φ* = 0.26 in toluene). *^c*A 590 nm long wave pass filter was used.

3-3 Conclusions

Herein, we report on the stereospecific synthesis of two single isomers of tris-heteroleptic tris-cyclometalated iridium(III) (Ir(III)) complexes consisting of three different nonsymmetric cyclometalating ligands via heteroleptic halogen-bridged Ir dimers [Ir(tpy)(F2ppy)(μ-Br)]² **34b** and $[\text{Ir}(mpiq)(F_2ppy)(\mu-Br)]_2$ **67b** (tpy: $(2-(4'-tolyl)pyridine)$ and F_2ppy : $(2-(4',6'-t))^2$ difluorophenyl)pyridine), and mpiq: $(1-(4'-methylphenyl)$ isoquinoline)) prepared by Zn^{2+} promoted degradation reactions of $Ir(tpy)_2(F_2ppy)$ **20** and $Ir(mpiq)_2(F_2ppy)$ **23**, as reported by us. Subsequently, **34b** and **67b** was converted to the tris-heteroleptic tris-cyclometalated Ir complexes $Ir(tpy)(F_2ppy)(mpiq)$ 66 consisting of tpy, F_2ppy , and mpiq, as confirmed by spectroscopic and X-ray crystal structure analysis. The important findings of this work are that: i) the specific isomers of tris-heteroleptic tris-cyclometalated such as **66a,b** and **66c,d** can be prepared from the different halogen-bridged Ir dimers **34b** and **67b**, while all the stereoisomers are composed of the same ligands. The mechanism of the formation of only two diastereomers from the µ-complex is discussed based on the X-ray crystal structures of **34b** and the product **66c**. The mechanism of the formation of only two diastereomers from the µ-complex is discussed based on the X-ray crystal structures of **34b** and the product **66c**. ii) each **66** isomers have different stability (in the presence of silica gel, in this work). iii) different stereoisomers of **66** exhibit different emission spectra. Namely, one stereoisomer **66a** exhibits single broad emission from *ca*. 550 nm to *ca*. 650 nm (orange emission), while another stereoisomer **66c** emits dual emission at *ca*. 509 nm and *ca*. 600 nm (pale pink emission). To the best of our knowledge, this is the first report on the selective and efficient synthesis, different stability and different photophysical properties of tris-heteroleptic tis-cyclometalated Ir(III) complexes. These information of tris-heteroleptic tris-cyclometalated Ir complexes represents a potentially useful synthetic method for preparing not only Ir complexes but also other metal complexes.

Chapter 4

Synthesis and Anticancer Properties of Bis- and Mono(cationic peptide) Hybrids of Bis-Heteroleptic Cyclometalated Iridium(III) Complexes: Effect of the Number of Peptide Units on Anticancer Activity

4-1 Introduction

 Aoki and co-workers previously reported Ir complex–peptide hybrids (IPHs) **70a–f** (with net charges from $+9$ to $+12$) containing a KKGG (K: lysine, G: glycine) cationic peptide a connected *via* a C6 or C8 alkyl chain linker between the peptide and the $Ir(tpy)$ ₃ core via regioselective substitution reactions of $fac-Ir(tpy)$ ₃ (2) and $fac-Ir(ppy)$ ₃ (1) at the 5'-position of the cyclometalated ligands (Chart 4-1).^{27,30,31} These IPHs showed considerable levels of anticancer activity against Jurkat cancer cells with EC_{50} values of 7–16 μ M.^{27a,e,f,g} We also reported on the synthesis of the Ir(III) complex-peptide hybrid **71** containing three KKKGG peptide units at the 5'-position of the tpy ligand through C8 alkyl chain linkers, which showed a more potent cytotoxicity against Jurkat cancer cells with an EC_{50} value of 1.5 μ M and which induced considerable morphological changes in Jurkat cells with a strong green emission.^{27f} Mechanistic investigations suggested that **70** induces an intracellular Ca^{2+} overload, resulting in the induction of a paraptosis-like cell death associated with autophagy.

Chart 4-1. Structures of cationic amphiphilic Ir complexes derived from Ir(tpy)₃ 2 and their EC⁵⁰ values against Jurkat cancer cells (net cationic charges in aqueous solution at natural pH are indicated in their structures).

fac-1 $(fac-Ir(ppy)_3)$: $R^1 = H$ *fac*-2 $(fac-Ir(tpy)_3)$: R¹ = Me

70a (n = 2): EC_{50} >50 µM against Jurkat cells

70b (n = 4): EC_{50} >50 µM

70c (n = 6): EC_{50} = 16 µM

70d (n = 8): EC_{50} = 7.3 µM

70e (n = 12): EC_{50} = 32 µM

70f (n = 16): EC_{50} >50 µM

71: $EC_{50} = 1.5 \mu M$ against Jurkat cells

 We report herein on the design and synthesis of Ir complexes, **72** and **73** containing two and one KKKGG sequences, respectively, for an evaluation of the effect of the number of the KKKGG peptide parts on their anticancer activities. The cytotoxicity of **72** (bispeptide) and **73** (monopeptide) against Jurkat, HeLa S3, and A549 cancer cells, and IMR90 normal cells was evaluated in comparison with the trispeptide complex **71**. The results of MTT assay indicated that complexes **71** and **72** exhibited more potent anticancer activity against Jurkat cells than **73**, indicating the existence of a positive correlation between the number of the KKKGG peptide units and their cytotoxicity, and a strong green emission was observed in dead cells. It was suggested that **71** and **72** induce the vacuolization of the cytoplasm in Jurkat cells, which are characteristic phenomena of paraptotic cell death, a relatively newly recognized type of programmed cell death, as confirmed by transmission electron microscopic (TEM) observations. It was also found that Ca^{2+} ions were transferred to mitochondria possibly from the endoplasmic reticulum (ER) and that this played a critical role in the induction of cell death. In addition, the localization of these complexes and intracellular morphology in the Jurkat cells were examined in co-staining experiments using specific probes for intracellular organelles and other microscopic methods.

4-2 Results and Discussions

4-2-1 Synthesis of Ir(III) Complexes Equipped with Bis or Mono Cationic Peptide Units

 The synthesis of the bis-heteroleptic cyclometalated Ir(III) complex-bispeptide conjugate **72** and the monopeptide conjugate **73** is shown in Chart 4-2 and 4-3, respectively. The biscarboxylated Ir(III) complex (**75**) was prepared from *fac*-**20**²⁴ (in the form of a racemic mixture of Δ and Λ forms) via Vilsmeier-Haack formylation, followed by a Pinick oxidation, as described in our previous papers.27a,g,30a The structure (*facial* form) of the formylated intermediate **74** was determined by a X-ray crystal structure analysis, as shown in Figure 4-1 (typical parameters of the crystal structure of **74** are listed in Table 4-1). The condensation of **75** with a mono-Boc-protected diamine (C8 alkyl chain linker) gave **76** and its two Boc groups were removed by treatment with a mixture of TMSCl and NaI to give **77**. The coupling reaction of **77** with the protected KKKGG peptide, which was prepared by a Fmoc solid-phase peptide synthesis method, was conducted, followed by the deprotection with TMSCl and NaI in $MeCN^{27g}$ and purification by reverse phase high performance liquid chromatography (RP-HPLC) afforded a TFA salt of **72**.

Chart 4-2. Synthesis of bis-heteroleptic cyclometalated Ir(III) complex-peptide hybrid **72** containing two cationic peptide units.

Figure 4-1. ORTEP drawing of single crystal structure of **74** with 50% probability ellipsoids.

Similarly, the cyclometalated Ir complex-monopeptide conjugate **73** was prepared from *fac***-21** via **78-82** (Chart 4-3).²⁴ The structure of the intermediate **78** was previously reported by us⁶² and that of **79** was also determined by a single crystal X-ray diffraction analysis, as shown in Figure 4-2 and Table 4-1. It should be noted that the diastereomers of **72** and **73** resulting from the racemic Ir centers (Δ and Λ forms) were not distinguished in the ¹H NMR spectra, and hence, were not separated.

Chart 4-3. Synthesis of bis-heteroleptic cyclometalated Ir(III) complex-peptide hybrid **73** that possess one cationic peptide unit.

Figure 4-2. ORTEP drawing of molecular structure of **79** with 50% probability ellipsoids. The complex crystallizes with solvent which we could not identify unequivocally due to its partial occupancy and extended disorder. Eventually the solvent was SQUEEZE'd using PLATON.

Table 4-1. Crystal data and structure refinement for **74** and **79**.

4-2-2 Photophysical Properties of IPHs 72 and 73

 The UV/Vis absorption and fluorescence emission spectra of **72** and **73** (10 µM) in degassed 100 mM HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid) solution (pH 7.4) at 298 K are displayed in Figure 4-3 and relevant photophysical data are listed in Table 4-2. The UV-Vis spectra of the Ir complexes exhibited an absorption maxima at ca. 280 nm and ca. 355 nm, which correspond to a $1\pi-\pi^*$ transition of the cyclometalated ligands and spin-allowed singletto-singlet metal-to-ligand charge transfer (¹MLCT) transitions, spin-forbidden singlet-to-triplet (³MLCT) transitions, and $3\pi-\pi^*$ transitions, as previously reported.^{27a,g} Luminescence spectra of **72** and **73** exhibited an emission maxima at ca. 502 nm (Figure 4-3b) and the emission quantum yields (Φ) were determined to be 0.44 and 0.12, respectively, and their lifetimes were found to be 1.6 and 1.8 µs, respectively, as summarized in Table 4-2. The spectral results are consistent with those for previously reported cyclometalated Ir complexes. $27a,e,g,30a$

Figure 4-3. (a) UV/Vis absorption and (b) emission spectra of **72** (plain curve) and **73** (dashed curve) in degassed 100 mM HEPES (pH 7.4) at 25 °C. [Ir complex] = 10 µM, excitation at λ = 366 nm. a.u. = arbitrary units. The photograph shows the emission image of **72** in 100 mM HEPES (pH 7.4) at 25 °C (excitation at λ = 366 nm).

Table 4-2. Photophysical properties of 2, 71, 72, and 73 at [Ir complex] = 10 μ M in degassed 100 mM HEPES ($pH = 7.4$) or CH₂Cl₂ at 25 °C.

Compound	$\lambda_{\rm max}$ (absorption)	ε (dm ³ mol ⁻¹ cm ⁻¹)	λ_{\max} (emission)	\varPhi	τ
$(in CH2Cl2)[a]$	287 nm, 373 nm	4.5×10^4 , $1.2 \times$ $10^{4[b]}$	$512 \text{ nm}^{[c]}$	0.50	$2.0 \,\mu s^{[b]}$
$71^{[d]}$	280 nm, 360 nm	7.7×10^4 , 1.4×10^4	507 nm \lbrack c]	$0.41^{[e]}$	$1.4 \,\mu s^{[f]}$
72	277 nm, 358 nm	8.2×10^4 , 2.2×10^4	502 nm ^[c]	$0.44^{[e]}$	$1.6 \,\mu s^{[f]}$
73	278 nm, 352 nm	1.2×10^5 , 2.0×10^4	498 nm ^[c]	$0.12^{[e]}$	$1.8 \,\mu s^{[f]}$

[a] Ref. 30a. [b] Ref. 8. [c] Excitation at 366 nm. [d] Ref. 27f. [e] Quinine sulfate in 0.1 M $H₂SO₄$ (Φ = 0.55) was used as a reference compound. [f] A 495 nm long wave pass filter was used.

4-2-3 Death of Cancer Cells Induced by 71–73.

 The morphological features and fluorescence microscopic images of T-lymphocyte leukemia Jurkat cells treated with trispeptide- (**71**), bispeptide- (**72**), and monopeptide-hybrids (**73**) were observed by fluorescence microscopy. The Jurkat cancer cells were incubated with or without (as a control) **71–73** (25 µM) in Roswell Park Memorial Institute (RPMI) medium containing 10% fetal bovine serum (FBS) for 30 min or 1 h at 37 ºC under 5% CO2, then collected by centrifugation, washed with PBS (phosphate buffer saline), and observed by fluorescence microscopy. The captured images suggest that **71** and **72** induce the death of Jurkat cancer cells and this is accompanied by considerable morphological changes and a strong green emission was detected in dead cells (Figure 4-4d-i for **71**, Figure 4-4j-o for **72**), while **73** weakly induced cell death (Figure 4-4p-u for **73**).

Figure 4-4. Representative luminescence microscopic images (Fluoview, FV-1000, Olympus) of Jurkat cells treated with **71**, **72**, and **73** (25 μ M) complexes at 37 °C for 30 min or 1 h, respectively. (a) Bright image of the control (in the absence of Ir complex), (b) emission image of the control, (c) overlay image of the control, (d) bright field image with **71** for 30 min, (e) emission image with **71** for 30 min, (f) overlay image with **71** for 30 min, (g) bright field image with **71** for 1 h, (h) emission image with **71** for 1 h, (i) overlay image with **71** for 1 h, (j) bright field image with **72** for 30 min, (k) emission image with **72** for 30 min, (l) overlay image with **72** for 30 min, (m) bright field image with **72** for 1 h, (n) emission image with **72** for 1 h, (o) overlay image with **72** for 1 h, (p) bright field image with **73** for 30 min, (q) emission image with **73** for 30 min, (r) overlay image with **73** for 30 min, (s) bright field image with **73** for 1 h, (t) emission image with **73** for 1 h, (u) overlay image with **73** for 1 h. Scale bar (black) denotes $10 \mu m$.

4-2-4 Effect of the Number of Peptide Units in IPHs on cytotoxicity against Cancer (Jurkat, HeLa S3, and A549) and Normal (IMR90) Cell Lines

The cytotoxicities of Ir complexes **71–73** against Jurkat, HeLa S3 (human cervical), A549 (human lung) cancer, and IMR90 (human fetal lung) normal cell lines were evaluated by means of a 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. Jurkat $(2.0 \times 10^4 \text{ cells/mL})$, HeLa S3 ($1.5 \times 10^4 \text{ cells/mL}$), A549 ($1.5 \times 10^4 \text{ cells/mL}$), and IMR90 (1.5 \times 10⁴ cells/mL) cells were incubated at 37 °C for 24 h in 10% FBS/RPMI 1640 (Jurkat and IMR90), FBS/Eagle's Minimum Essential Medium (MEM) (HeLa S3), or FBS/Dulbecco's Modified Eagle Medium (DMEM) (A549) containing 71 , 72 , or 73 (0–25 μ M or 0–50 μ M). The plots of % of cell viability *versus* the concentration of the Ir complex are shown in Figure 4-5a–c, from which the half maximal effective concentration (EC_{50}) values of **71–73** against Jurkat cells were calculated to be 1.4 μ M, 4.1 μ M, and 16.3 μ M, respectively. These results clearly indicate that a positive correlation exists between the number of H2N−KKKGG peptide groups and the degree of cytotoxicity against Jurkat cells. In addition, the cytotoxicity of **71– 73** is not so potent against HeLa S3 cells, A549 cells, and IMR90 cells (model of normal cells) (Figure 4-5b–d).

 It was previously reported that celastrol, a naturally occurring triterpenoid, induces paraptosis with the swelling of the ER and/or mitochondria and an overload in the concentration of intracellular Ca^{2+} , $27e, f, 63$ The EC₅₀ values of celastrol (Chart 4-4) against Jurkat, HeLa S3, and A549 cells were determined to be 0.5 μ M (Jurkat), 26.9 μ M (HeLa S3), and 40.5 μ M (A459), respectively, as shown in Figure 4-5a–c and Table 4-3.

Figure 4-5. MTT assay results for (a) Jurkat cells, (b) HeLa S3 cells, (c) A459 cells, and (d) IMR90 normal cells treated with **71**, **72**, **73**, and celastrol in 10% FBS/RPMI (Jurkat and IMR-90 cells), 10% FBS/MEM (HeLa S3), or 10% FBS/DMEM (A459 cells) medium (incubation at 37 ºC for 24 h). The net charge of each Ir complex is assumed to be +12 (**71**), +8 (**72**), and +4 (**73**), respectively.

Chart 4-4. Chemical structure of celastrol.

Table 4-3. The EC₅₀ values of the Ir complexes 71, 72, 73, and celastrol against Jurkat, HeLa S3, A549 cancer, and IMR90 normal cells.

 $NT = not tested$

4-2-5 IPHs Induce Paraptotic Cell Death of Jurkat Cells

 The programmed cell death (PCD) can be classified into several types, i.e., apoptosis (DNA fragmentation, chromatin condensation, blebbing, cell shrinkage, *etc*.,), autophagy (degradation of lysosomes), necroptosis (plasma membrane rupture or breakdown, swelling of organelles, *etc*.,), and paraptosis (cytoplasmic vacuolation involving the swelling of the ER and mitochondria, enhancement of intracellular Ca^{2+} concentration).⁶⁴⁻⁶⁷ We previously reported that different types of IPHs induce different types of PCD in Jurkat cells.²⁷ More recently, we have reported that **71** and its analogs induce paraptosis-like cell death through cytoplasmic vacuolization by the damage of intracellular organelles (lysosomes, mitochondria, and the ER) and morphological changes in Jurkat cells, as confirmed by methylene blue staining and transmission electron microscopic (TEM) analyses,^{27f} which are characteristic phenomena of paraptotic cell death.⁶⁸

 This background prompted us to determine whether vacuolization of cytoplasm and morphological changes of Jurkat cells treated with **71**, **72**, and celastrol had occurred. Jurkat cells were treated with **71**, **72** (25 μ M, 1 h), or celastrol (1 μ M, 24 h) and fixed with glutaraldehyde and osmium tetroxide $(OsO₄)$, and then embedded in Poly 812 resin, which was sliced and stained for TEM analysis. The vacuolization of the cytoplasm in Jurkat cells shown in Figure 4-6 strongly suggest that these compounds induce similar PCD, which can be classified as paraptosis.⁶⁹

Figure 4-6. TEM images of Jurkat cells treated with (b) celastrol $(1 \mu M, 24 h)$, (c) 71 (25 μ M, 1 h) and (d) 72 (25 μ M, 24 h) at 37 °C. Green arrows in (b), (c), and (d) show cytoplasmic vacuolization induced by **71** and **72**, respectively. Scale bar (black) = $2 \mu m$.

4-2-6 Mitochondrial Ca2+ Overload Induced by 71 and 72

It is known that there is a relationship between intracellular Ca^{2+} signaling and paraptosis and other types of cell death.^{63,70-74} The time-dependent enhancement in intracellular Ca^{2+} concentration was observed in fluorescence microscopic images when using Rhod-2/AM, which is a mitochondrial Ca²⁺ indicator, and Rhod-4/AM, a cytosolic Ca²⁺ indicator.^{27a,f,g} The fluorescent intensity of Rhod-2 was enhanced in mitochondria compared to that of Rhod-4 in the cytosol (Figure 4-7a and 4-7b) within few minutes after the addition of **71** and **72 (**25 μM), and morphological changes were then observed in 30-60 minutes (see also Figure 4-8). In contrast, only a very weak emission was observed in the mitochondria (Figure 4-7a) and in the cytosol (Figure 4-7b) of the **73**–treated Jurkat cells.

Figure 4-7. Representative luminescence confocal microscopic images of Jurkat cells treated with (a) Rhod-2 (red emission) and (b) Rhod-4 (red emission), followed by the treatment with **71–73** (25 μM for 5 min). Excitation at 540 nm for Rhod-2 and Rhod-4.

Figure 4-8. (a) Representative luminescence confocal microscopic images of Jurkat cells treated with Rhod-2 (red emission), followed by **71–73** (25 μM). (b) Representative luminescence confocal microscopic images of Jurkat cells treated with Rhod-4 (red emission), followed by **71–73** (25 μM). Excitation at 540 nm for Rhod-2 and Rhod-4.

 $Ca²⁺$ concentrations in the cytosol and mitochondria were evaluated in more detail by means of flow cytometric analysis (Figure 4-9). Jurkat cells were pretreated with Rhod-4/AM or Rhod-2/AM for 30 minutes and then with $71-73$ (25 μ M) for specified amounts of time, after which the cells were immediately examined by flow cytometry. Figure 4-9a, 4-9c, and 4-9e present the emission intensity of Rhod-4 after the treatment with **71**–**73** for 0-60 min, and Figure 4-9b, 4-9d, and 4-9f display the emission changes in Rhod-2 after treatment with **71**–**73** for 0- 60 min. Considerable enhancement in the red emission of Rhod-2 was induced by **71** and **72** (Figure 4-9b and 4-9d), respectively, while negligible enhancement was detected in the presence of **73** (Figure 4-9f). These results are nearly parallel to the anticancer activity of **71**– **73**, strongly suggesting the IPH–induced cell death is induced by an enhancement in Ca^{2+} concentration mainly in mitochondria. A weak correlation between the anticancer activity of **71–73** and the increase in Ca^{2+} concentrations in cytosol suggests the possible direct transfer of Ca^{2+} into mitochondria, possibly from the ER, which is a main Ca^{2+} reservoir in living cells. In contrast, celastrol induced a Ca^{2+} overload in the cytosol rather than in mitochondria, as shown in Figure 4-9g and4-9h. These data indicate that the paraptosis induced by **71–73** and celastrol proceeds *via* different mechanisms.

Figure 4-9. Flow cytometric analysis of Jurkat cells after the treatment with (a) Rhod-4 (5 μ M) and **71** (25 μ M), (b) Rhod-2 (5 μ M) and **71** (25 μ M), (c) Rhod-4 (5 μ M) and **72** (25 μ M), (d) Rhod-2 (5 μ M) and **72** (25 μ M), (e) Rhod-4 (5 μ M) and **73** (25 μ M), (f) Rhod-2 (5 μ M) and **73** (25 μ M), (g) Rhod-4 (5 μ M) and celastrol (30 μ M), and (h) Rhod-2 (5 μ M) and celastrol (25 M). Different colors correspond to the incubation time with **71–73** and celastrol: control (black), 5 min (light blue), 10 min (orange), 20 min (green), 30 min (dark green), 40 min (brown), 50 min (purple), 60 min (blue) in (a-f), respectively.

4-2-7 Co-staining of Jukat Cells with Ir(III) Complexes 71–73 and Fluorescence Probes for Intracellular Organelles

We next carried out co-staining experiments of Jurkat cancer cells with a mitochondria probe (MitoTracker Red), a lysosomal probe (LysoTracker Red), and an ER probe (ER-Tracker Red) in the presence or absence of **71–73** to examine their localization during the cell death process. In these experiments, Jurkat cells were treated with **71–73** (25 µM) for 30 min or 1 h, and then stained with MitoTracker Red (100 nM), LysoTracker Red (500 nM), or ER-Tracker Red (500 nM) at 37 ºC for 1 h.

Figure 4-10 shows confocal microscopic images (Fluoview, FV-1000, Olympus) of Jurkat cells treated with **71** and MitoTracker Red (Figure 4-10d-k), cells treated with **72** and MitoTracker Red **(**Figure 4-10l-s), and cells treated with **73** and MitoTracker Red (Figure 4- 10t-aa), respectively (a green color emission from **71**–**73** indicates dead cells). Figure 4-11 shows confocal microscopic images of cells that had been treated with **71** and LysoTracker Red (Figure 4-11d-k), with **72** and LysoTracker Red **(**Figure 4-11l-s), and with **73** and LysoTracker Red (Figure 4-11t-aa), respectively. Figure 4-12 shows confocal microscopic images of cells treated with **71** and ER-Tracker Red (Figure 4-12d-k), with **72** and ER-Tracker Red **(**Figure 4- 12l-s), and with **73** and ER-Tracker Red (Figure 4-12t-aa), respectively. Among these images, only Figures 4-10k and 4-10s show a yellow emission region resulting from overlay of the green emission of IPHs (**71** and **72)** and the red emission of MitoTracker, suggesting that **71** and **72** are localized mainly in mitochondria. In addition, a weak red emission of LysoTracker Red and ER-Tracker Red was observed in the presence of **71** and **72** (Figure 4-11j, 4-11k, 4-11r, and 4-11s for LysoTracker Red and Figure 4-12j, 4-12k, 4-12r, and 4-12s for ER-Tracker Red), suggesting the degradation (structural breakdown or disappearance) of lysosomes and the ER during the cell death process induced by these IPHs.

Figure 4-10. Representative luminescent confocal microscopic images (Fluoview, FV-1000, Olympus) of Jurkat cells incubated with **71–73** ($25 \mu M$) for 30 min or 1 h, followed by treatment with MitoTracker Red (100 nM, 1 h). (a), (d), (h), (l), (p), (t), and (x) Bright field images of Jurkat cells, (e) and (i) emission images with **71**, (m) and (q) emission images with **72**, and (u) and (y) emission images with **73**. (b), (f) , (f) , (n) , (r) , (v) , and (z) Emission images with MitoTracker Red, (c) overlay image of (a) and (b), (g) overlay image of (d–f), (k) overlay image of $(h-j)$, (o) overlay image of $(l-n)$, (s) overlay image of $(p-r)$, (w) overlay image of $(t-v)$, and (aa) overlay image of $(x-z)$. Excitation at 405 nm for (e), (i), (m), (q), (u), and (y) and 559 nm for (b), (f), (i), (n), (r), (v), and (z). Exposure time was $12.5 \mu s$ /pixel. Scale bar (black) = 10 μ m.

Figure 4-11. Representative luminescent confocal microscopic images (Fluoview, FV-1000, Olympus) of Jurkat cells incubated with **71**–**73** (25 µM) for 30 min or 1 h, followed by treatment with LysoTracker Red (500 nM, 1 h). (a), (d), (h), (l), (p), (t), and (x) Bright field images of Jurkat cells, (e) and (i) emission images with **71**, (m) and (q) emission images with **72** and (u) and (y) emission images with 73 . (b), (f), (j), (n), (r), (v), and (z) Emission images with LysoTracker Red, (c) overlay image of (a) and (b), (g) overlay image of (d–f), (k) overlay image of $(h-j)$, (o) overlay image of $(l-n)$, (s) overlay image of $(p-r)$, (w) overlay image of $(t-v)$, and (aa) overlay image of $(x-z)$. Excitation at 405 nm for (e), (i), (m), (q), (u), and (y) and 559 nm for (b), (f), (j), (n), (r), (v), and (z). Exposure time was $12.5 \mu s / pixel$. Scale bar (black) = 10 um.

Figure 4-12. Representative luminescent confocal microscopic images (Fluoview, FV-1000, Olympus) of Jurkat cells incubated with **71–73** (25 µM) for 30 min or 1 h, followed by treatment with ER-Tracker Red $(500 \text{ nM}, 1 \text{ h})$. (a), (d), (h), (l), (p), (t), and (x) Bright field images of Jurkat cells, (e) and (i) emission images with **71**, (m) and (q) emission images with **72**, and (u) and (y) emission images with **73**. (b), (f) , (i) , (n) , (r) , (v) , and (z) Emission images with ER-Tracker Red, (c) overlay image of (a) and (b), (g) overlay image of (d–f), (k) overlay image of $(h-j)$, (o) overlay image of $(l-n)$, (s) overlay image of $(p-r)$, (w) overlay image of $(t-v)$, and (aa) overlay image of $(x-z)$. Excitation at 405 nm for (e), (i), (m) , (q) , (u) , and (y) and 559 nm for (b), (f), (j), (n), (r), (v), and (z). Exposure time was $12.5 \mu s / pixel$. Scale bar (black) = 10 μ m.

4-2-8 Plausible Mechanism of Cell death Induced by IPHs

A plausible mechanism of the cell death of Jurkat cells induced by IPHs **71** and **72** is summarized in Figure 4-13 based on the aforementioned experimental results, as summarized below.

- (1) IPHs containing three and two KKKGG peptide segments **71** and **72** induce the cell death of Jurkat cells stronger than the monopeptide conjugate **73**, as shown in Figures 4-5 and 4- 6, indicating the existence of a positive relationship between the number of KKKGG chains and their cytotoxicity. In addition, a positive relationship between the cytotoxicity of **71**, **72**, and **73** against Jurkat cells and mitochondrial Ca^{2+} overload was observed (Figure 4-8) and 4-9).
- (2) It is previously reported that cytotoxicity of IPHs having $Ir(tpy)$ ₃ core such as **70a-f** is dependent on the cationic charge and the peptide sequence as well as the linker length.^{27a} In this work, however, the number of KKKGG sequence and the total cationic charge of **71**, **72**, and **73** are not separable. Therefore, we conclude that the cytotoxicity of **71**, **72**, and **73** is dependent on the number of KKKGG units and their net positive charge (+12 for **71**, +8 for **72**, and +4 for **73**).
- (3) It is very likely that IPHs **71** and **72** induce paraptotic cell death in Jurkat cells, which is a newly recognized type of PCD,⁶⁶ as evidenced by cytoplasmic vacuolization (Figure 4-6) together with our previous results. Although metal complexes such as rhenium, copper, ruthenium, and titanium have been reported to induce paraptotic cell death in cancer cell lines,⁷⁵⁻⁸¹ our IPHs are the first example to induce the direct transfer of Ca^{2+} from the ER to mitochondria for the induction of paraptotic cell death in such a short incubation time (1 h). Very recently, the positive correlation between the increase in the intracellular uptake of IPHs and emission enhancement of IPHs in dead cells was discovered.^{27f,j}
- (4) IPHs 71 and 72 enhance the transfer of Ca^{2+} into mitochondria rather than to the cytosol possibly from the ER, which is an intracellular Ca^{2+} -storing organelle, at the early stage of cell death, as confirmed by confocal microscopic observations and flow cytometric analysis

using mitochondrial and cytosolic Ca^{2+} probes (Rhod-2 and Rhod-4, respectively) (Figures 4-8 and 4-9). In contrast, **73** induces only a negligible overload of mitochondrial Ca^{2+} . It was previously reported that the change in Ca^{2+} concentration in a culture medium negligibly changes the cell death induced by IPHs.^{27a,g} These results suggest that the influx of Ca^{2+} into mitochondria possibly from the ER is one of the critical events for the induction of cell death in Jurkat cells.

- (5) Celastrol, a naturally-occurring triterpenoid, also induces paraptosis (Figures 4-4 and 4-5). However, the paraptotic mechanism induced by celastrol is different from that by **71** and **72** (Figure 4-9). This difference may have some relationship with different cytotoxicity profiles of IPHs and celastrol against Jurkat, HeLa S3, A549, and IMR90 cells (Table 4-3).
- (6) The findings of this study indicate that **71** and **72** are localized mainly in mitochondria (Figure 4-10) and induce the degradation (structural breakdown or disappearance) of the ER (Figure 4-11), as confirmed by co-staining experiments with MitoTracker Red and ER-Tracker Red, respectively. The relationship of mitochondria and ER with programmed cell death has been extensively discussed in the past.82-86 We assume that **71** and **72** interact with biomolecules in/on mitochondria, the ER membrane or mitochondria-associated membranes (MAMs) to activate (or inactivate) some signaling pathways for the induction of mitochondria- and ER-related cell death.
- (7) IPHs **71–73** induce the degradation of lysosomes during the cell death process (Figure 4- 12). In our previous paper, we concluded that autophagy may not be a major pathway for the induction of paraptosis-like cell death, although lysosomal degradation and the upregulation of autophagy markers such as LC3, Beclin-1, and Atg-12 are induced by IPHs.^{27f} Therefore, autophagy may not be a major cause for the induction of paraptosislike cell death that is induced by IPHs **71** and **73**.

Figure 4-13. Proposed cell death mechanism for IPHs in Jurkat cancer cells.

4-3 Conclusions

In this study, bis(peptide) and mono(peptide) conjugates (**72** and **73**) of cyclometalted Ir(III) complexes were synthesized in addition to our previous tris(peptide) hybrid **71** and the effect of the number of H2N-KKKGG chains in IPHs on their anticancer activity was examined. The new Ir complexes were extensively characterized by various spectroscopic and analytical methods including X-ray single crystal structure analysis of the synthetic intermediates. The prepared IPHs show a green emission with lifetimes are in the order of μ s, which are due to phosphorescence from the excited triplet state.

 The *in vitro* anticancer activity of IPHs, **71**, **72**, and **73**, against a panel of three cancer cell lines (Jurkat, HeLa S3, and A549) and one non-cancerous (IMR90) cell line was evaluated. The IPHs **71** and **72** exhibit potential anticancer activity against Jurkat cells and dead cells show a strong green emission. Complexes **71** and **72** show more potent activity towards Jurkat cancer cells, indicative of a positive relationship between the number of the KKKGG peptide chains and hence net cationic charges and their anticancer activity. Mechanistic studies of cell death suggest that **71**, **72**, and celastrol induce paraptotic cell death in Jurkat cells with the vacuolization of the cytoplasm, mitochondria, the ER, and lysosomes, as confirmed by TEM images of Jurkat cells, although the cell death mechanism between IPHs and celastol is different. The findings reported herein also strongly suggested that the direct transfer of Ca^{2+} from the ER to mitochondria is essential for the induction of cell death in Jurkat cells, as confirmed by fluorescent microscopic observations and flow cytometric analysis by using intracellular Ca^{2+} probes. The results of co-staining assays of **71**–**73** with probes that are specific for different intracellular organelles suggest that these IPHs are localized in mitochondria for the induction of paraptotic cell death in Jurkat cells. More detailed mechanistic studies regarding direct Ca^{2+} transfer from ER to mitochondria are currently underway.

These results provide useful information for the design and synthesis of metal complexes and other organic molecules containing a greater number of peptide units and their analogs as potential anticancer agents in medicinal chemistry, bioinorganic chemistry, bioorganometallic chemistry, biomedical science, and other related research fields.

Chapter 5

Concluding Remarks

In this thesis, the efficient synthesis and photophysical properties of *tris-heteroleptic* Ir complexes represented by IrLL'A or IrLL'L'' and the biological applications of newly synthesized bis-heteroleptic Ir complexes $IrL₂L'$ compared with homoleptic IrL₃ complexes are also reported.

In Chapter 2, the degradation reactions of tris-cyclometalated Ir complexes (IrL₃, L: cyclometalating ligand) promoted by Brønsted acids such as HCl/1,4-dioxane and Lewis acids such as ZnX_2 (X = Br or Cl), TMSCl, and AlCl₃ were reported. Among the various Lewis acids, ZnBr₂ exhibited a good reactivity for $Ir(ppy)$ ³ **1**, $Ir(tpy)$ ³ **2**, and $Ir(mppy)$ ³ **12** to provide the corresponding halogen-bridged Ir dimers (μ -complexes). It was also found that the reactivity of tris-cyclometalated Ir complexes containing electron-withdrawing groups such as fluorine atoms, nitro or CF³ groups on the ligands is quite low. This reaction was applied to the selective degradation of *mer*-tris-cyclometalated Ir complexes such as *mer*-Ir(tpy)₂(F₂ppy) (*mer*-20). The reaction of *mer***-20** with ZnBr² gave a mixture of a halogen-bridged Ir dimer **15b** and **34b**, and the tris-heteroleptic Ir complex **25** (IrLL'A, A: ancillary ligand) was then obtained by treatment with acetylacetone. Furthermore, we successfully isolated a novel tris-heteroleptic Ir complex **42a** having a non-symmetric ancillary ligand after careful separation of its diastereomers. Mechanistic studies suggest that the formation of different products from some Ir complexes by H^+ , TMSCl, and AlCl₃ and ZnX_2 is due to the hardness and softness of these Brønsted and Lewis acids used. The reactivity and selectivity in the ZnX2-promoted degradation can be explained by the interaction of the HOMO of Ir complexes with the LUMO of ZnX² and by the difference of electronegativity of iridium and carbon in the ligand part. Further mechanistic study is now underway.

In Chapter 3, we report on the stereospecific synthesis of two single isomers of trisheteroleptic tris-cyclometalated iridium(III) (Ir(III)) complexes consisting of three different nonsymmetric cyclometalating ligands via heteroleptic halogen-bridged Ir dimers $[\text{Ir(tpy)}(F_2ppy)(\mu-Br)]_2$ **34b** and $[\text{Ir(mpiq)}(F_2ppy)(\mu-Br)]_2$ **67b** (tpy: (2-(4'-tolyl)pyridine) and F2ppy: (2-(4',6'-difluorophenyl)pyridine), and mpiq: (1-(4'-methylphenyl)isoquinoline)) prepared by Zn^{2+} -promoted degradation reactions of Ir(tpy)₂(F₂ppy) **20** and Ir(mpiq)₂(F₂ppy) **23**, as reported by us. Subsequently, **34b** and **67b** was converted to the tris-heteroleptic triscyclometalated Ir complexes $Ir(tpy)(F_2py)(mpiq)$ 66 consisting of tpy, F_2ppy , and mpiq, as confirmed by spectroscopic and X-ray crystal structure analysis. The important findings of this work are that: i) the specific isomers of tris-heteroleptic tris-cyclometalated such as **66a,b** and **66c,d** can be prepared from the different halogen-bridged Ir dimers **34b** and **67b**, while all the stereoisomers are composed of the same ligands. The mechanism of the formation of only two diastereomers from the µ-complex is discussed based on the X-ray crystal structures of **34b** and the product **66c**. The mechanism of the formation of only two diastereomers from the µcomplex is discussed based on the X-ray crystal structures of **34b** and the product **66c**. ii) each **66** isomers have different stability (in the presence of silica gel, in this work). iii) different stereoisomers of **66** exhibit different emission spectra. Namely, one stereoisomer **66a** exhibits single broad emission from *ca*. 550 nm to *ca*. 650 nm (orange emission), while another stereoisomer **66c** emits dual emission at *ca*. 509 nm and *ca*. 600 nm (pale pink emission). To the best of our knowledge, this is the first report on the selective and efficient synthesis, different stability and different photophysical properties of tris-heteroleptic tis-cyclometalated Ir(III) complexes.

In Chapter 4, bis(peptide) and mono(peptide) conjugates (**72** and **73**) of bis-heteroleptic cyclometalted Ir(III) complexes were synthesized in addition to our previous tris(peptide) hybrid **71** and the effect of the number of H2N-KKKGG chains in IPHs on their anticancer activity was examined. The new Ir complexes were extensively characterized by various spectroscopic and analytical methods including X-ray single crystal structure analysis of the synthetic intermediates. The prepared IPHs show a green emission with lifetimes are in the order of μ s, which are due to phosphorescence from the excited triplet state.

 The *in vitro* anticancer activity of IPHs, **71**, **72**, and **73**, against a panel of three cancer cell lines (Jurkat, HeLa S3, and A549) and one non-cancerous (IMR90) cell line was evaluated. The IPHs **71** and **72** exhibit potential anticancer activity against Jurkat cells and dead cells show a strong green emission. Complexes **71** and **72** show more potent activity towards Jurkat cancer cells, indicative of a positive relationship between the number of the KKKGG peptide chains and hence net cationic charges and their anticancer activity. Mechanistic studies of cell death suggest that **71**, **72**, and celastrol induce paraptotic cell death in Jurkat cells with the vacuolization of the cytoplasm, mitochondria, the ER, and lysosomes, as confirmed by TEM images of Jurkat cells, although the cell death mechanism between IPHs and celastol is different. The findings reported herein also strongly suggested that the direct transfer of Ca^{2+} from the ER to mitochondria is essential for the induction of cell death in Jurkat cells, as confirmed by fluorescent microscopic observations and flow cytometric analysis by using intracellular Ca^{2+} probes. The results of co-staining assays of **71**–**73** with probes that are specific for different intracellular organelles suggest that these IPHs are localized in mitochondria for the induction of paraptotic cell death in Jurkat cells. More detailed mechanistic studies regarding direct Ca^{2+} transfer from ER to mitochondria are currently underway.

In conclusion, we have succeeded in the efficient synthesis and the measurements of photophysical properties of *tris-heteroleptic* Ir complexes represented by IrLL'A or IrLL'L'' and the biological applications of newly synthesized bis-heteroleptic Ir complexes $IrL₂L$ ^t to compare with these properties with those of homoleptic IrL₃ complexes. These information of tris-heteroleptic tris-cyclometalated Ir complexes represents a potentially useful synthetic method for preparing not only Ir complexes but also other metal complexes. Moreover, the aforementioned results provide useful information for the design and synthesis of metal complexes and other organic molecules containing a greater number of peptide units and their analogs as potential anticancer agents in medicinal chemistry, bioinorganic chemistry, bioorganometallic chemistry, biomedical science, and other related research fields.

Experimental Section

General Information.

IrCl₃·3H₂O was purchased from KANTO CHEMICAL Co. Glycerol, AgOTf, and Trypsin was purchased from NACALAI TESQUE, INC. Anhydrous 1,2-dichloroethane was obtained by distillation from calcium hydride and 1,2-dichloroethane-*d⁴* was purchased from SIGMA-ALDRICH. Anhydrous *N*,*N*-dimethylformamide (DMF) was obtained by distillation from calcium hydride. All aqueous solutions were prepared using deionized and distilled water. MTT (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2*H*-tetrazolium bromide) was purchased from Dojindo. ¹H NMR spectra (300 MHz and 400 MHz) were recorded on a JEOL Always 300 spectrometer, a JEOL Lambda 400 spectrometer and IR spectra were recorded with a Perkin-Elmer FTIR Spectrum 100 (ATR). Luminescence imaging studies were performed using fluorescent microscopy (Biorevo, BZ-9000, Keyence). Electrospray ionization (ESI) mass spectra were recorded on a Varian 910-MS spectrometer. Mass of some tris-cyclometalated Ir complexes are observed as $[M]^+$ (rather than $[M^+H]^+$) in ESI mode (Varian TQ-FT). Lyophilization was performed with freeze dryer FD-5N (EYELA). Thin-layer (TLC) and silica gel column chromatographies were performed using Merck 5554 (silica gel) TLC plates and Fuji Silysia Chemical FL-100D, respectively. Emission lifetimes were determined using a TSP-1000 spectrometer (Unisoku Co., Ltd.). Commercially available DMSO (spectrophotometric grade, WAKO CHEMICAL Co) was used for the measurement of photophysical data. Elemental analyses were performed on a Perkin-Elmer CHN 2400 series II CHNS/O analyzer. Elemental analyses of the Ir complexes, except for *mer***-10** and **36** were not carried out because the metal and/or halogen contents of these compounds are >25%. GPC experiments were carried out using a system consisting of a POMP P-50 (Japan Analyrical Industry Co., Ltd.), a UV/VIS DETECTOR S-3740 (Soma, Japan), a Manual Sample Injector 7725i (Rheodyne, USA), a MDL-101 1 PEN RECORDER (Japan Analyrical Industry Co., Ltd.), equipped with two preparative GPC, JAIGEL-1H and JAIGEL-2 (Japan Analyrical Industry

Co., Ltd.) (20 ϕ x 600 mm, No. A605201 and A605204) were used. HPLC experiments were carried out using a system consisting of two PU-980 intelligent HPLC pumps (JASCO, Japan), a UV-970 intelligent UV-visible detector (JASCO), a Rheodine injector (Model No. 7125) and a Chromatopak C-R6A (Shimadzu, Japan). For analytical HPLC, a SenshuPak Pegasil ODS column (Senshu Scientific Co., Ltd.) $(4.6\varphi \times 250 \text{ mm}, \text{No. } 07051001)$ was used. For preparative HPLC, a SenshuPak Pegasil ODS SP100 column (Senshu Scientific Co., Ltd.) $(20\varphi \times 250 \text{ mm}, \text{No. } 1302014 \text{ G})$ was used. Density functional theory (DFT) calculations were also carried out using the Gaussian09 program⁵⁴ (B3LYP, the LanL2DZ basis set for a Ir and Br atoms and the 6-31G basis set for H, C, F, S, O, N atoms).⁸⁷ Time-dependent DFT (TD-DFT) calculations were carried out based on all the ground state geometries.⁸⁸ The chromaticity values obtained from emission spectra of Ir complexes were plotted on a color diagram using a ColorAC software.

Synthesis.

fac-1,⁸ mer-1,⁸ fac-2,⁸ mer-2,⁸ fac-3,⁸ mer-3,⁸ fac-12,^{27b} fac-16,^{27a} fac-17,^{28a} fac-18,^{27a} fac-**19**,^{28a} **6**,^{55b} and **29b**^{55b} were prepared in the form of a racemic mixture of Λ and Δ forms and their spectral data were in good agreement with reported data.

15b: A solution of *fac***-2** (30 mg, 43 μmol) and ZnBr2 (0.50 g, 2.2 mmol) in 1,2 dichloroethane (2.0 mL) was refluxed for 3 h. After cooling to room temperature, the insoluble materials were removed by filtration and the filtrate was concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography using CHCl₃ as the eluent to give 15b as a yellow powder (24 mg, 93% yield). mp > 300 °C. IR (ATR): $v = 1604$, 1587, 1560, 1476, 1462, 1426, 1152, 1065, 769, 747, 664, 427 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.43 (d, *J* = 5.7 Hz, 4H), 7.82 (d, *J* = 6.9 Hz, 4H), 7.74 (td, *J* = 6.9, 1.2 Hz, 4H), 7.36 (d, *J* = 8.1 Hz, 4H), 6.81 (td, *J* = 6.3, 1.8 Hz, 4H), 6.55 (d, *J* = 7.2 Hz, 4H), 5.72 $(s, 4H)$, 1.93 $(s, 12H)$. ESI-MS (m/z) . Calcd for C₄₈H₄₀N₄⁸¹Br₂¹⁹¹Ir₂ [M]⁺: 1216.0782. Found: 1216.0841.

 d_{12} **-15a:** A solution of **15a** (0.43 g, 0.38 mmol) in a mixture of 1,2-dichloroethane (8.0 mL) and DCl/D_2O (2.0 mL) was refluxed for 16 h. After cooling to room temperature, the solvent was removed imcompletely under reduced pressure and the insoluble materials was filtrated to give *d12*-**15a** as a greenish yellow powder (0.30 g, 70% yield and deuteration rate is 92% determined by ¹H-NMR). mp > 300 °C. IR (ATR): $v = 3074$, 2913, 2236, 2040, 1966, 1603, 1557,1471, 1440, 1397, 1372, 1336, 1304, 1264, 1237, 1160, 1035, 995, 967, 944, 887, 800, 764, 750, 717, 703, 667, 631, 565, 523, 505, 490, 456, 429, 416 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.16 (d, *J* = 5.7 Hz, 4H), 7.80 (d, *J* = 8.1 Hz, 4H), 7.70 (t, *J* = 7.2 Hz, 4H), 6.71 (t, *J* = 5.7 Hz, 4H), 1.93 (s, 12H).

*mer***-17:** A mixture of chloro-bridged Ir dimer $[\{Ir(mpiq)z(\mu-Cl)\}z]$ (26a) (prepared from 1-(4'-methylphenyl)isoquinoline (mpiq) and IrCl3∙3H2O as shown in Chart 6) (0.20 g, 0.18 mmol), K_2CO_3 (0.56 g, 4.0 mmol) and mpiq (0.26 mg, 0.12 mmol) in glycerol (5.0 mL) was stirred at 150 °C for 8 h. After cooling to room temperature, water was added to the reaction mixture and extracted with CHCl₃. The combined organic layer was dried over Na₂SO₄ and filtered to obtain the filtrate, to which hexanes and CH_2Cl_2 were added to obtain *mer***-17** as a red powder (0.13 g, 52% yield). mp 219-220 °C. IR (ATR): $v = 2917, 1580, 1499, 1438, 1347,$ 1311, 1269, 1206, 1146, 1120, 1040, 868, 811, 737, 672, 581, 418 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.99-8.87 (m, 3H), 8.20 (d, *J* = 8.4 Hz, 1H), 8.09 (t, *J* = 6.6 Hz, 3H), 7.70-7.56 (m, 10H), 7.52 (d, *J* = 6.3 Hz, 1H), 7.10 (d, *J* = 6.3 Hz, 1H), 6.97 (d, *J* = 6.9 Hz, 1H), 6.90 (d, *J* = 6.6 Hz, 1H), 6.85 (d, *J* = 8.1 Hz, 2H), 6.78 (d, *J* = 8.7 Hz, 1H), 6.74 (s, 1H), 6.47 (s, 1H), 6.32 (s, 1H), 2.13 (s, 3H), 2.11 (s, 3H), 2.09 (s, 3H). ESI-MS (m/z). Calcd for C₄₈H₃₆N₃¹⁹¹Ir $[M]^+$: 845.2510. Found: 845.2514. Anal. Calcd for C₄₈H₃₆N₃Ir: C, 68.06; H, 4.28; N, 4.96. Found: C, 67.70; H, 4.19; N, 4.89.

fac-20: A solution of the F_2 ppy ligand (0.15 g, 0.76 mmol), the chloro-bridged Ir dimer $[\{Ir(tpy)_2(\mu-Cl)\}_2]$ **15a** (0.20 g, 0.18 mmol), and AgOTf (90 mg, 0.35 mmol) in 2-ethoxyethanol (3.0 mL) was refluxed for 3 h. The yellow precipitate was filtrated on a filter and dried for 1 h. The combined bright yellow powder was purified by silica gel column chromatography using hexanes/CH₂Cl₂ (2/1) as the eluent to afford *fac*-20 as a yellow powder (0.12 g, 46% yield). mp $>$ 300 °C. IR (ATR): $v = 1597, 1554, 1470, 1397, 1264, 1237, 1159, 1098, 982, 829, 808, 769,$ 748, 725, 717, 567, 523, 429 cm-1 . ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 8.26 (d, *J* = 8.4 Hz, 1H), 7.83 (dd, *J* = 7.8, 3.3 Hz, 2H), 7.62-7.50 (m, 6H), 7.44 (d, *J* = 5.4 Hz, 1H), 7.39 (d, *J* = 5.4 Hz, 1H), 6.88-6.79 (m, 3H), 6.73 (td, *J* = 7.2, 1.8 Hz, 2H), 6.64 (s, 1H), 6.61 (s, 1H), 6.39-

6.30 (m, 2H), 2.17 (s, 3H), 2.12 (s, 3H). ESI-MS (m/z). Calcd for C₃₅H₂₆N₃F₂¹⁹¹Ir [M]⁺: 717.1695. Found: 717.1702.

mer-20: A mixture of 20a (0.20 g, 0.18 mmol), K_2CO_3 (0.25 g, 1.8 mmol) and the F₂ppy ligand (0.14 g, 0.72 mmol) in glycerol (5.0 mL) was stirred at 150 °C for 24 h. After cooling to room temperature, water was added to the reaction mixture and extracted with CHCl3. The combined organic layer was dried over Na2SO4, filtered, and the resulting filtrate was concentrated under reduced pressure and then purified by silica gel column chromatography using hexanes/CHCl₃ (2/1) as the eluent to afford **mer-20** as a yellow powder (0.19 g, 74%) yield). mp > 300 °C. IR (ATR): $v = 1588$, 1552, 1466, 1420, 1392, 1281, 1262, 1231, 1158, 1095, 1054, 977, 872, 806, 769, 750, 717, 569, 522, 426 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/Si(CH₃)₄) δ = 8.29 (d, *J* = 8.4 Hz, 1H), 8.00 (d, *J* = 6.0 Hz, 1H), 7.94 (d, *J* = 5.2 Hz, 1H), 7.75 (d, *J* = 8.0 Hz, 2H), 7.62-7.46 (m, 6H), 6.88 (t, *J* = 6.4 Hz, 1H), 6.78-6.68 (m, 4H), 6.39- 6.33 (m, 3H), 6.18 (s, 1H), 2.14 (s, 3H), 2.12 (s, 3H). ESI-MS (m/z). Calcd for C₃₅H₂₆N₃F₂¹⁹¹Ir [M]⁺: 717.1695. Found: 717.1696.

*fac***-21:** A solution of the tpy ligand (40 mg, 0.24 mmol), **30a** (0.10 g, 82 μmol), and AgOTf (78 mg, 0.30 mmol) in 2-ethoxyethanol (2.0 mL) was refluxed for 11 h. The yellow precipitate was dissolved in CHCl₃ and washed with water. The combined organic layer was dried over Na2SO4. After filtration, the solvent was concentrated under reduced pressure and the resulting residue was purified by silica gel chromatography using hexanes/CHCl₃ (10/1) as the eluent to give *fac*-21 as a yellow powder (74 mg, 61% yield), IR (ATR): $v = 1596$, 1565, 1555, 1471, 1396, 1282, 1236, 1158, 1098, 982, 823, 771, 746, 717, 566, 524, 419 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.29-8.27 (m, 2H), 7.85 (d, J = 8.7 Hz, 1H), 7.68-7.58 (m, 3H), 7.54 (d, *J* = 8.1 Hz, 1H), 7.50 (d, *J* = 5.7 Hz, 1H), 7.46 (d, *J* = 5.7 Hz, 1H), 7.34 (d, *J* = 6.6 Hz, 1H), 6.936.83 (m, 3H), 6.74 (dd, *J* = 8.1, 1.2 Hz, 1H), 6.58 (s, 1H), 6.42-6.26 (m, 4H), 2.15 (s, 3H). ESI-MS (*m/z*). Calcd for C₃₄H₂₂N₃F₄¹⁹¹Ir [M]⁺: 739.1350. Found: 739.1351.

*mer***-21:** A mixture of AgOTf (0.32 g, 1.2 mmol), chrolo-bridged Ir dimer $[\{Ir(F_2ppy)_{2}(\mu -$ Cl) $\{2, 0.30 \text{ g}, 0.25 \text{ mmol}\}$, the tpy ligand (0.12 g, 0.75 mmol) and triethylamine (0.14 g, 0.75 mmol) in 1,2-dichloroethane (50 mL) was refluxed for 2 h. The mixture was then cooled to room temperature, and the precipitate was removed by filtration. The filtrate was evaporated under reduced pressure and purified by silica gel column chromatography using CHCl₃ as the eluent to afford *mer*-21 as a yellow powder (0.26 g, 72% yield), mp > 300 °C. IR (ATR): $v =$ 1595, 1564, 1551, 1469, 1397, 1285, 1237, 1157, 1096, 981, 834, 807, 772, 749, 711, 566, 524 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/Si(CH₃)₄) δ = 8.21-8.19 (m, 2H), 8.09 (d, *J* = 5.1 Hz, 1H), 7.89 (d, *J* = 8.1 Hz, 1H), 7.82 (d, *J* = 4.8 Hz, 1H), 7.65-7.50 (m, 5H), 6.89 (td, *J* = 6.0, 1.2Hz, 1H), 6.83 (dd, *J* = 7.5, 1.5 Hz, 1H), 6.75 (q, *J* = 7.5 Hz, 2H), 6.69 (s, 1H), 6.46-6.30 (m, 2H), 5.98 (dd, *J* = 6.9, 2.1 Hz, 1H), 5.81 (dd, *J* = 9.6, 2.1 Hz, 1H), 2.17 (s, 3H). ESI-MS (*m/z*). Calcd for C₃₄H₂₂N₃F₄¹⁹¹Ir [M]⁺: 739.1350. Found: 739.1358.

*mer***-22:** *mer***-22** was obtained as a yellow powder (67 mg, 17% yield) from $[\{Ir(mpy)_{2}(\mu -$ Cl) $\{2\}$ (0.30 g, 0.25 mmol), K₂CO₃ (0.35 g, 2.5 mmol), glycerol (15 ml) and F₂ppy ligand (0.14 g, 0.76 mmol) using a precedure similar to that for *mer***-20**. The product was purified by silica gel column chromatography with CHCl₃ as the eluent. mp > 300 °C. IR (ATR): $v = 1579$, 1508, 1476, 1427, 1399, 1278, 1213, 1158, 1101, 1042, 984, 835, 768, 589, 567, 525, 420 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.30 (dd, *J* = 8.1, 2.1 Hz, 1H), 7.98-7.96 (m, 2H), 7.68-7.43 (m, 8H), 6.89 (t, *J* = 6.9 Hz, 1H), 6.71-6.63 (m, 2H), 6.56-6.34 (m, 4H), 6.10 (d, *J* = 2.7 Hz, 1H), 5.91 (d, *J* = 2.7 Hz, 1H), 3.62 (s, 3H), 3.58 (s, 3H). ESI-MS (*m/z*). Calcd for $C_{35}H_{26}N_3O_2F_2^{191}$ Ir [M]⁺: 749.1593. Found: 749.1578.

*mer***-23:** *mer***-23** was prepared as a red powder (0.20 g, 81% yield) from **25a** (0.20 g, 0.15 mmol), K_2CO_3 (0.42 g, 3.0 mmol), glycerol (5 mL) and F_2 ppy ligand (0.22 g, 0.90 mmol) using a precedure similar to that for *mer***-20**. The product was purified by silica gel column chromatography with hexanes/CH₂Cl₂ (2/1) as the eluent. mp 252-253 °C. IR (ATR): $v = 1583$, 1554, 1500, 1470, 1440, 1391, 1348, 1264, 1231, 1159, 1148, 1122, 1096, 1043, 978, 867, 810, 751, 737, 717, 672, 579, 567, 522 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.98-8.89 (m, 2H), 8.31 (d, *J* = 8.1 Hz, 1H), 8.16 (d, *J* = 8.4 Hz, 1H), 8.11 (d, *J* = 8.1 Hz, 1H), 7.95 (d, *J* = 6.3 Hz, 1H), 7.78-7.73 (m, 3H), 7.66-7.57 (m, 5H) 7.43 (d, *J* = 6.3 Hz, 1H), 7.07-7.01 (m, 2H), 6.85-6.77 (m, 3H), 6.45-6.24 (m, 4H), 2.09 (s, 3H), 2.08 (s, 3H). ESI-MS (*m/z*). Calcd for $C_{43}H_{30}N_3F_2^{191}$ Ir [M]⁺: 817.2008. Found: 817.2008.

*fac***-24:** A solution of the mpiq ligand (0.56 g, 2.6 mmol), **30a** (0.41 g, 0.34 mmol), and AgOTf (0.16 g, 0.63 mmol) in 2-ethoxyethanol (6.0 mL) was refluxed for 18 h. The orange precipitate was filtrated on a filter and dried for 1 h. The combined orange powder was purified by silica gel column chromatography using hexanes/CHCl³ (1/1) as the eluent to afford *fac***-30** as an orange powder (0.40 g, 75% yield). mp > 300 °C. IR (ATR): $v = 3081$, 1972, 1599, 1567, 1556, 1504, 1472, 1445, 1431, 1397, 1351, 1283, 1265, 1238, 1160, 1100,981, 821, 807, 787, 750, 740, 717, 675, 578, 568, 525 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.95-8.92 (m, 1H), 8.29 (t, *J* = 9.3 Hz, 2H), 8.08 (d, *J* = 7.5 Hz, 1H), 7.78-7.75 (m, 1H), 7.69-7.58 (m, 4H), 7.47 (d, *J* = 5.4 Hz, 1H), 7.35 (d, *J* = 5.4 Hz, 1H), 7.29 (d, *J* = 6.0 Hz, 1H), 7.18 (d, *J* = 6.3 Hz, 1H), 6.92 (t, *J* = 6.0 Hz, 1H), 6.85-6.77 (m, 3H), 6.43-6.29 (m, 3H), 6.18 (dd, *J* = 9.6, 1.8 Hz, 1H), 2.21 (s, 3H). ESI-MS (m/z). Calcd for C₃₈H₂₄N₃F₄¹⁹¹Ir [M]⁺: 789.1506. Found: 789.1511

mer-24: The chloro-bridged Ir dimer $[\{Ir(F_2ppy)_2(\mu-Cl)\}_2]$ 30a (0.10 g, 78 μ mol), AgOTf $(0.10 \text{ g}, 0.39 \text{ mmol})$, triethylamine $(23 \text{ mg}, 0.24 \text{ mmol})$, the mpiq ligand $(52 \text{ mg}, 0.24 \text{ mmol})$ were reacted in 1,2-dichloroethane (15 mL) using a precedure similar to that for *mer***-24**. The crude product was purified by silica gel column chromatography with hexanes/CHCl₃ $(4/1)$ as the eluent to give *mer*-24 as an orange powder (60 mg, 48% yield). mp > 300 °C. IR (ATR): v $= 1594, 1567, 1552, 1471, 1397, 1286, 1237, 1157, 1112, 1097, 1039, 981, 807, 783, 752, 740,$ 711, 676, 567, 524, 383 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.94-8.91 (m, 1H), 8.21-8.19 (m, 2H), 8.10 (d, *J* = 8.1 Hz, 1H), 8.06 (d, *J* = 5.7 Hz, 1H), 7.80-7.76 (m, 2H), 7.70- 7.63 (m, 2H), 7.56-7.49 (m, 3H), 7.21 (d, *J* = 5.7 Hz, 1H), 6.90 (dd, *J* = 7.8, 1.5 Hz, 1H), 6.84 (s, 1H), 6.75-6.65 (m, 2H), 6.48-6.37 (m, 2H), 5.98 (dd, *J* = 8.1, 2.4 Hz, 1H), 5.81 (dd, *J* = 8.1, 2.4 Hz, 1H), 2.22 (s, 3H). ESI-MS (m/z). Calcd for C₃₈H₂₄N₃F₄¹⁹¹Ir [M]⁺: 789.1507. Found: 789.1503.

*d6***-6**: a mixture of *d12*-**15a** (10 mg, 8.6 μmol), tetrabutylammonium hydroxide (TBAH) (50 mg, 77 μ mol) and acetylacetone (6.0 mg, 56 μ mol) in CH₂Cl₂/MeOH (10/1, 1.1 mL) was refluxed for 1 h. After cooling to room temperature, water was added to the reaction mixture and the resulting solution was extracted with CHCl3. The combined organic layer was dried over Na2SO4. After filtration, the solvent was concentrated under reduced pressure and the resulting residue was purified by silica gel chromatography using hexanes/ CH_2Cl_2 (4/1) as the eluent to give d_6 **-6** as a yellow powder (7.6 mg, 69%). IR (ATR): $v = 2918$, 2161, 1723, 1606, 1574, 1517, 1473, 1423, 1395, 1358, 1308, 1259, 1223, 1202, 1156, 1068, 1042, 1017, 926, 871, 775, 750, 604, 586, 426 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.47 (d, *J* = 6.0 Hz, 2H), 7.79 (d, *J* = 7.8 Hz, 2H), 7.70 (td, *J* = 7.2, 1.5 Hz, 2H), 7.08 (td, *J* = 6.9, 1.5 Hz, 2H), 5.18 (s, 1H), 2.04 (s, 6H), 1.76 (s, 6H). ESI-MS (m/z). Calcd for C₂₉¹H₂₁²H₆N₂O₂F₂¹⁹¹Ir [M]⁺: 632.2049. Found: 632.2046.

25: A solution of *mer***-20** (20 mg, 28 μmol) and ZnBr2 (0.31 g, 1.4 mmol) in 1,2 dichloroethane (2.0 mL) was refluxed for 3 h. After cooling to room temperature, the insoluble materials were removed by filtration washing with CHCl₃ and the filtrate was concentrated under reduced pressure. The crude product was purified by silica gel column chromatography using CHCl₃ as the eluent to give the mixture of **15b** and **34b** (18 mg, quant. assuming that only **34b** was obtained) as a yellow powder.

As reported by Baranoff and co-workers,^{36c} a mixture of 15b and 34b obtained above (18) mg, 14 μmol), tetrabutylammonium hydroxide (TBAH) (30 mg, 42 μmol) and acetylacetone $(6.0 \text{ mg}, 56 \text{ µmol})$ in CH₂Cl₂/MeOH $(10/1, 1.1 \text{ mL})$ was refluxed for 1 h. After cooling to room temperature, water was added to the reaction mixture and the resulting solution was extracted with CHCl₃. The combined organic layer was dried over Na₂SO₄. After filtration, the solvent was concentrated under reduced pressure and the resulting residue was purified by silica gel chromatography using hexanes/ CH_2Cl_2 (4/1) as the eluent to give 25 as a yellow powder (10) mg, 63% yield from **mer-20**). mp > 300 °C. IR (ATR): $v = 2923$, 1579, 1508, 1477, 1399, 1101, 985, 836, 768, 587, 567, 422, 413 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/Si(CH₃)₄) δ = 8.51 (dd, *J* $= 6.4, 0.8$ Hz, 1H), 8.40 (d, $J = 5.6$ Hz, 1H), 8.22 (d, $J = 8.0$ Hz, 1H), 7.80-7.68 (m, 3H), 7.43 (d, *J* = 7.6 Hz, 1H), 7.14 (td, *J* = 6.8, 1.2 Hz, 1H), 7.10 (td, *J* = 5.2, 1.2 Hz, 1H), 6.66 (dd, *J* = 8.0, 1.2 Hz, 1H), 6.28 (td, *J* = 10.8, 2.4 Hz, 1H), 6.00 (s, 1H), 5.71 (dd, *J* = 9.0, 2.4 Hz, 1H), 5.22 (s, 1H), 2.06 (s, 3H), 1.81 (s, 3H), 1.79 (s, 3H). ESI-MS (m/z). Calcd for C₂₈H₂₃N₂O₂F₂¹⁹¹Ir [M]⁺: 648.1328. Found: 648.1327.

26b: 26b was obtained as a red powder (25 mg, 98% yield) from *fac***-17** (30 mg, 35 μmol) and ZnBr² (0.38 g, 1.7 mmol) using a precedure similar to that for **15b** (2.0 mL of 1,2 dichloroethane was used as the solvent). mp > 300 °C. IR (ATR): $v = 1585$, 1501, 1444, 1378, 1348, 1271, 1148, 1124, 1045, 867, 812, 782, 738, 671, 579, 483, 414 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.22 (d, *J* = 6.3 Hz, 4H), 8.93 (d, *J* = 8.4 Hz, 4H), 8.00-7.93 (m, 8H), 7.82 (t, *J* = 7.2 Hz, 4H), 7.76 (t, *J* = 7.2 Hz, 4H), 6.70 (d, *J* = 6.0 Hz, 4H), 6.62 (d, *J* = 6.6 Hz, 4H), 5.85 (s, 4H), 1.86 (s, 12H). ESI-MS (m/z). Calcd for C₆₄H₄₈N₄⁸¹Br₂¹⁹¹Ir₂ [M]⁺: 1416.1406. Found: 1416.1514.

 $fac-27$: A solution of $fac-2$ (30 mg, 43 µmol) and $FeCl₃$ (58 mg, 2.2 mmol) in 1,2dichloroethane (2.0 mL) was stirred at room temperature for 2 h. The solvent was removed by evaporation. The crude residue was purified by silica gel column chromatography using CHCl³ as the eluent to give *fac*-27 as a yellow powder $(3.1 \text{ mg}, 9\% \text{ yield})$. mp $> 300 \degree \text{C}$. IR (ATR): v $=$ 1598, 1467, 1421, 1255, 1063, 1043, 882, 779, 741, 614 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 7.79 (d, *J* = 7.8 Hz, 3H), 7.59 (t, *J* = 9.0 Hz, 3H), 7.58 (s, 3H), 7.42 (d, *J* $= 5.7$ Hz, 3H), 6.86 (t, $J = 5.7$ Hz, 3H), 6.63 (s, 3H), 2.16 (s, 9H). ESI-MS (m/z). Calcd for $C_{36}H_{27}N_3{}^{35}Cl_3{}^{191}Ir$ [M]⁺: 797.0871. Found: 797.0871.

30b: 30b was obtained as a yellow powder (12 mg, 55% yield) from *mer***-3** (25 mg, 33 μmol) and ZnBr² (0.46 mg, 2.0 mmol) using a precedure similar to that for **15b** (2.0 mL of 1,2 dichloroethane was used as the solvent). mp > 300 °C. IR (ATR): $v = 1599$, 1572, 1557, 1478, 1402, 1293, 1248, 11660, 1102, 988, 829, 782, 753, 715, 708, 568, 527 cm⁻¹. ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 9.40 (d, *J* = 5.1 Hz, 4H), 8.33 (d, *J* = 8.4 Hz, 4H), 7.87 (td, *J* = 8.1, 1.8 Hz, 4H), 6.91 (td, *J* = 6.9, 0.9 Hz, 4H), 6.33 (td, *J* = 11.1, 2.1 Hz, 4H), 5.28 (dd, *J* = 9.3, 2.1 Hz, 4H). ESI-MS (*m*/z). Calcd for C₄₄H₂₄N₄F₈⁸¹Br₂¹⁹¹Ir₂ [M]⁺: 1303.9406. Found: 1303.9467.

31b: 31b was obtained as a yellow powder (13 mg, 50% yield) from *fac***-1** (30 mg, 46 μmol) and ZnBr² (0.51 g, 2.3 mmol) using a precedure similar to that for **15b** (2.0 mL of 1,2 dichloroethane was used as the solvent). mp > 300 °C. IR (ATR): $v = 1605$, 1580, 1477, 1421, 1157, 1030, 754, 740, 732, 724, 670, 629, 420 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.51 (d, *J* = 5.4 Hz, 4H), 7.89 (d, *J* = 7.8 Hz, 4H), 7.78 (td, *J* = 7.2, 1.2 Hz, 4H), 7.48 (d, *J* = 7.8 Hz, 4H), 6.86 (td, *J* = 6.6, 1.5 Hz, 4H), 6.74 (td, *J* = 6.9, 1.2 Hz, 4H), 6.58 (td, *J* = 5.1, 1.5 Hz, 4H), 5.92 (d, $J = 7.8$ Hz, 4H). ESI-MS (m/z). Calcd for C₄₄H₃₂N₄⁸¹Br₂¹⁹¹Ir₂ [M]⁺: 1160.0158. Found: 1160.0210.

*d4***-31a:** A solution of **31a** (37 mg, 34 μmol) in a mixture of 1,2-dichloroethane (2.0 mL) and $DCI/D₂O$ (0.5 mL) was refluxed for 2 h. After cooling to room temperature, the solvent was removed incompletely under reduced pressure and the insoluble materials was filtrated to give *d4*-**31a** as a greenish yellow powder (22 mg, 61% yield and deuteration rate is 99% determined by ¹H-NMR). mp > 300 °C. IR (ATR): $v = 3041, 2250, 1951, 1604, 1561, 1475, 1425, 1407$, 1365, 1287, 1267, 1192, 1158, 1086, 1057, 1020, 969, 873, 804,752, 725, 662, 648, 624, 561 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.24 (d, *J* = 5.7 Hz, 4H), 7.86 (d, *J* = 7.2 Hz, 4H), 7.73 (t, *J* = 6.6 Hz, 4H), 6.79-6.73 (m, 8H), 6.56 (t, *J* = 7.5 Hz, 4H), 5.93 (t, *J* = 7.5 Hz, 4H).

32b: 32b was obtained as a yellow powder (20 mg, 80% yield) from *fac***-12** (30 mg, 40 μmol) and ZnBr² (0.45 g, 2.0 mmol) using a precedure similar to that for **15b** (2.0 mL of 1,2 dichloroethane was used as the solvent). mp > 300 °C. IR (ATR): $v = 1581$, 1547, 1459, 1425, 1277, 1209, 1158, 1033, 769, 748, 586 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.44 (d, *J* = 6.3 Hz, 4H), 7.76-7.68 (m, 8H), 7.41 (d, *J* = 8.7 Hz, 4H), 6.77 (td, *J* = 6.3, 1.8 Hz, 4H), 6.35
(dd, *J* = 8.4, 2.4 Hz, 4H), 5.43 (d, *J* = 2.7 Hz, 4H), 3.41 (s, 12H). ESI-MS (*m/z*). Calcd for $C_{48}H_{40}N_4O_4^{81}Br_2^{191}Ir_2 [M]^+$: 1280.0578. Found: 1280.0628.

*d12***-32a:** A solution of **32a** (36 mg, 25 μmol) in a mixture of 1,2-dichloroethane (2.0 mL) and DCl/D₂O (0.5 mL) was refluxed for 6 h. After cooling to room temperature, the solvent was removed incompletely under reduced pressure and the insoluble materials was filtrated to give *d4*-**27a** as a greenish yellow powder (28 mg, 93% yield and deuteration rate is 99% determined by ¹H-NMR). mp > 300 °C. IR (ATR): $v = 2931, 2853, 2581, 2278, 2161, 2038, 1604, 1559$, 1528, 1474, 1459, 1389, 1345, 1312, 1280, 1265, 1200, 1152, 1031, 977, 887, 848, 800, 772, 752, 690, 573 cm-1 . ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 9.17 (d, *J* = 5.7 Hz, 4H), 7.74- 7.60 (m, 8H), 6.68 (t, *J* = 7.5 Hz, 4H), 3.40 (s, 12H).

33: 33 was obtained as a yellow powder (20 mg, 83% yield) from *fac***-16** (30 mg, 32 μmol) and ZnBr² (0.36 g, 1.6 mmol) using a precedure similar to that for **15b** (2.0 mL of 1,2 dichloroethane was used as the solvent). mp > 300 °C. IR (ATR): $v = 1606$, 1574, 1558, 1471, 1422, 1358, 1267, 1221, 1156, 1068, 1039, 1016, 866, 776, 748, 604, 422 cm⁻¹. ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 9.41 (d, *J* = 5.7 Hz, 4H), 7.82-7.77 (m, 8H), 7.62 (s, 4H), 6.89-6.85 $(m, 4H)$, 5.69 (s, 4H), 1.98 (s, 12H). ESI-MS (m/z) . Calcd for C₄₈H₃₆N₄⁷⁹Br₄⁸¹Br₂¹⁹¹Ir₂ [M]⁺: 1527.7238. Found: 1527.7202.

34b: A solution of 20 (30 mg, 42 μ mol) and ZnBr₂ (0.58 g, 2.6 mmol) in ClCH₂CH₂Cl (2.0) mL) was refluxed for 3 h. After cooling to room temperature, the solvent was concentrated under reduced pressure. The acetone and water were added to the crude reside and sonicated for 10 min. The yellow precipitate was filtered on and dried, to which was added a mixture of hexanes and CH₂Cl₂ to give **34b** as a yellow crystaline powder (18 mg, 68% yield). mp > 300 $^{\circ}$ C. IR (ATR): $v = 1602$, 1569, 1557, 1477, 1463, 1427, 1162, 1104, 986, 835, 751, 566, 527, 429 cm^{-1} . ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.48 (m, 2H), 9.36 (m, 2H), 8.28 (d, *J* = 6.9 Hz, 4H), 7.87-7.75 (m, 6H), 7.40 (d, *J* = 3.9 Hz, 1H), 7.37 (d, *J* = 3.9 Hz, 1H), 6.90-6.84 (m, 4H), 6.63 (d, *J* = 3.9 Hz, 1H), 6.60 (d, *J* = 2.1 Hz, 1H), 6.27 (tt, *J* = 10.5, 2.7 Hz, 2H), 5.63 (s, 1H), 5.62 (s, 1H), 5.37 (dt, *J* = 9.3, 3.0 Hz, 2H), 1.96 (s, 3H), 1.95 (s, 3H). ESI-MS (*m/z*). Calcd for $C_{46}H_{32}N_4F_4^{81}Br_2^{191}Ir_2$ [M]⁺: 1260.0094. Found: 1260.0149.

35 and **36**: **35** (yellow powder, 9.1 mg, 54% yield) and **36** (yellow powder, 2.0 mg, 13% yield) was obtained from *mer***-22** (20 mg, 27 μmol), ZnBr² (0.30 g, 1.3 mmol), acetylacetone (6.0 mg, 54 μmol) and tetrabutylammonium hydroxide (TBAH) (25 mg, 40 μmol) using a procedure similar to that for **25**. The product was purified by silica gel column chromatography with hexanes/ $CH_2Cl_2(4/1)$ as the eluent.

35: mp 261-263 °C. IR (ATR): $v = 2925$, 1579, 1508, 1476, 1399, 1278, 1213, 1158, 1101, 1042, 984, 835, 768, 588, 567, 525, 420, 412 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.51 (dd, *J* = 5.4, 1.2 Hz, 1H), 8.36 (d, *J* = 5.7 Hz, 1H), 8.22 (d, *J* = 7.8 Hz, 1H), 7.75-7.69 (m, 3H), 7.50 (d, *J* = 8.7 Hz, 1H), 7.14 (td, *J* = 6.3, 1.2 Hz, 1H), 7.08 (td, *J* = 6.3, 2.1 Hz, 1H), 6.66 (dd, *J* = 8.0, 1.2 Hz, 1H), 6.28 (td, *J* = 10.8, 2.4 Hz, 1H), 5.72 (dd, *J* = 8.7, 2.7 Hz, 1H), 5.69 (d, *J* = 2.7 Hz, 1H), 5.20 (s, 1H), 3.55 (s, 3H), 1.80 (s, 3H), 1.79 (s, 3H). ESI-MS (*m/z*). Calcd for $C_{28}H_{23}N_2O_3F_2^{191}$ Ir [M]⁺: 664.1277. Found: 664.1282.

36: mp > 300 °C. IR (ATR): $v = 2923$, 1579, 1508, 1477, 1399, 1101, 985, 836, 768, 587, 567, 422, 413 cm-1 . ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 8.43 (d, *J* = 5.4 Hz, 2H), 7.72- 7.63 (m, 4H), 7.47 (d, *J* = 8.7 Hz, 2H), 7.04 (td, *J* = 6.3, 1.8 Hz, 2H), 6.40 (dd, *J* = 8.7, 2.4 Hz, 2H), 5.77 (d, *J* = 2.7 Hz, 2H), 5.20 (s, 1H), 3.53 (s, 6H), 1.78 (s, 6H). ESI-MS (*m/z*). Calcd for $C_{29}H_{27}N_2O_4^{191}$ Ir [M]⁺: 658.1571. Found: 658.1578.

37: **37** was prepared as an orange powder (12 mg, 67% yield from *mer***-23**) from *mer***-23** (20 mg, 24 μmol), $ZnBr_2$ (0.27 g, 1.2 mmol), acetylacetone (6.0 mg, 54 μmol) and tetrabutylammonium hydroxide (TBAH) (25 mg, 40 μmol) using a precedure similar to that for **25**. The product was purified by silica gel column chromatography with hexanes/ $CH_2Cl_2(2/1)$ as the eluent. mp > 300 °C. IR (ATR): $v = 1566$, 1557, 1515, 1391, 1290, 1265, 1245, 1100, 985, 835, 813, 782, 753, 737, 714, 672, 588, 574, 567, 526 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.98-8.94 (m, 1H), 8.52 (d, *J* = 5.4 Hz, 1H), 8.33 (d, *J* = 6.3 Hz, 1H) 8.27 (d, *J* = 8.4 Hz, 1H), 8.10 (d, *J* = 8.1 Hz, 1H), 7.93-7.90 (m, 1H), 7.77 (t, *J* = 7.5 Hz, 1H), 7.72- 7.69 (m, 2H), 7.44 (d, *J* = 6.0 Hz, 1H), 7.18 (td, *J* = 6.6, 1.2 Hz, 1H), 6.76 (dd, *J* = 7.8, 2.1 Hz, 1H), 6.28 (td, *J* = 10.2, 2.1 Hz, 1H), 6.15 (s, 1H), 5.70 (dd, *J* = 8.7, 2.4 Hz, 1H), 5.21 (s, 1H), 2.12 (s, 3H), 1.82 (s, 3H), 1.73 (s, 3H). ESI-MS (m/z). Calcd for C₃₂H₂₅N₂O₂F₂¹⁹¹Ir [M]⁺: 698.1484. Found: 698.1476.

38: **38** was prepared for the characterization of the products of degradation reaction of *mer***-23** (to prove that only **37** was produced, not **38**, from *mer***-23**). A solution of **26b** (10 mg, 7.0 μmol), tetrabutylammonium hydroxide (TBAH) (13 mg, 21 μmol) and acetylacetone (5 mg, 50 μ mol) in CH₂Cl₂/MeOH (10/1, 1.1 mL) was refluxed for 14 h. After cooling to room temperature, water was added to the reaction mixture and extracted with CHCl3. The combined organic layer was dried over Na2SO4. After filtration, the filtrate was evaporated under reduced pressure and the resulting residue was purified by silica gel chromatography using hexanes/CH₂Cl₂ (5/1) as the eluent to give 38 as a red powder (8.9 mg, 89% yield). mp $>$ 300 $^{\circ}$ C. IR (ATR): $v = 3021, 1576, 1513, 1502, 1437, 1395, 1373, 1350, 1271, 1252, 1148, 1123,$

1049, 1014, 869, 813, 741, 733, 671, 584, 576, 415, 396 cm⁻¹. ¹H NMR (300 MHz, CDCl3/Si(CH3)4) δ = 9.00-8.97 (m, 2H), 8.41 (d, *J* = 6.3 Hz, 2H), 8.10 (d, *J* = 8.4 Hz, 2H), 7.93- 7.90 (m, 2H), 7.73-7.68 (m, 4H), 7.42 (d, *J* = 6.6 Hz, 2H), 6.71 (dd, *J* = 9.0, 1.8 Hz, 2H), 6.23 $(s, 2H)$, 5.16 $(s, 1H)$, 2.02 $(s, 6H)$, 1.73 $(s, 6H)$. ESI-MS (m/z) . Calcd for C₃₇H₃₁N₂O₂¹⁹¹Ir [M]⁺: 726.1986. Found: 726.1979.

42a: A solution of *mer-***20** (10 mg, 14 μmol) and ZnBr2 (0.20 g, 0.89 mmol) in 1,2 dichloroethane (1.0 mL) was refluxed for 4 h. After the precipitate was filtered off and the filtrate was concentrated under reduced pressure, the resulting residue was purified by silica gel column chromatography using CHCl³ as the eluent to give the mixture of **15b** and **34b** as a yellow powder (6.4 mg, 72% yield assuming that only **34b** was obtained).

The next step was carried out according to our previous report.¹³ The obtained mixture of **15b** and $34b$ (6.4 mg, 5.2 µmol) was dissolved in a mixture of CH₂Cl₂/EtOH (4/1, 1.3 mL), and 8-benzenesulfonylamidoquinoline (5.0 mg, 18 μmol) and triethylamine (38 mg, 0.37 mmol) were then added. The reaction mixture was refluxed at 80 $^{\circ}$ C for 1 h. The solvent was evaporated under reduced pressure and the residue purified by silica gel chromatography with CHCl₃ as the eluent to give the stereoisomer mixture of **37** as a brown powder (4.1 mg, 36% yield from *mer***-20**). The stereoisomer mixture of **42** was subjected to further purification by silica gel column chromatography eluted with hexanes/CHCl3/THF (4/1/1) to afford **42a** as a brown powder (1.0 mg, 10% yield from *mer*-20). mp > 300 °C. IR (ATR): $v = 2923$, 1579, 1508, 1477, 1399, 1101, 985, 836, 768, 587, 567, 422, 413 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.20 (dd, *J* = 5.7, 0.6 Hz, 1H), 8.22-8.16 (m, 2H), 8.06 (dd, *J* = 8.7 1.5 Hz, 1H) 7.75-7.64 (m, 3H), 7.54-7.52 (m, 1H), 7.46 (t, *J* = 8.1 Hz, 1H), 7.36 (d, *J* = 8.1 Hz, 1H), 7.25-7.12 (m, 6H), 7.04-6.96 (m, 3H), 6.71-6.68 (m, 2H), 6.41 (td, *J* = 10.8, 2.4 Hz, 1H), 6.05 (s, 1H), 5.64 (dd, *J*

 $= 8.7, 2.1$ Hz, 1H), 2.12 (s, 3H). ESI-MS (m/z). Calcd for C₃₈H₂₇N₄O₂F₂S¹⁹¹Ir [M]⁺: 832.1423. Found: 832.1426.

43: The authentic sample of **43** was prepared as described in our previous report to characterize the products of the degradation reactions of *mer***-20**. 13 A solution of the chlorobridged Ir dimer $[\{Ir(tpy)_2(\mu-Cl)\}_2]$ 15a (52 mg, 45 μmol), 8-benzenesulfonylamidoquinoline (28 mg, 97 µmol), and triethylamine (0.20 mg, 2.0 mmol)in $CH_2Cl_2/EtOH$ (4/1, 2.5 mL) was stirred at room temperature for 10 min. The solvent was evaporated under reduced pressure and the residue was purified by silica gel chromatography with CHCl³ as the eluent to give **43** as a brown powder (68 mg, 95% yield), mp > 300 °C. IR (ATR): $v = 1588$, 1562, 1475, 1459, 1381, 1311, 1138, 1109, 1086, 942, 849, 822, 746, 689, 580, 566, 528, 427 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.12 (dd, *J* = 4.8, *J* = 0.9 Hz, 1H), 8.15 (dd, *J* = 7.8, 1.2 Hz, 1H), 8.00 (dd, *J* = 8.4, 1.2 Hz, 1H), 7.73-7.66 (m, 3H), 7.61-7.52 (m, 2H), 7.46-7.36 (m, 3H), 7.29 (d, *J* = 6.6 Hz, 1H), 7.19-7.09 (m, 5H), 7.00-6.91 (m, 3H), 6.74-6.64 (m, 3H), 6.13 (s, 1H), 5.97 (s, 1H), 2.10 (s, 3H), 2.08 (s, 3H). ESI-MS (m/z). Calcd for C₃₉H₃₁N₄O₂S¹⁹¹Ir [M]⁺: 810.1768. Found: 810.1759. Anal. Calcd for C₃₉H₃₁N₄O₂SIr⋅0.1CHCl₃: C, 57.00; H, 3.80; N, 6.80. Found: C, 56.73; H, 3.78; N, 6.56.

*mer***-Ir(** d_3 **-tpy)2**(ppy): A mixture of d_{12} **-15a** (0.13 g, 0.12 mmol), K₂CO₃ (0.22 g, 1.6 mmol) and ppy $(0.11 \text{ g}, 0.70 \text{ mmol})$ in glycerol (1.0 mL) was stirred at 150 °C for 16 h. After cooling to room temperature, water was added to the reaction mixture and extracted with CHCl3. The combined organic layer was dried over Na2SO⁴ and filtered to obtain the filtrate, to which hexanes and AcOEt were added to obtain *mer*-Ir(d_3 -tpy)₂(ppy) as a yellow powder (0.13 g, 55% yield). IR (ATR): $v = 3322, 3061, 3039, 3018, 2918, 2859, 2255, 2217, 2030, 1973, 1863, 1733,$ 1596, 1574, 1556, 1465, 1431, 1409, 1391, 1297, 1256, 1237, 1221, 1159, 1115, 1054, 1034,

1025, 993, 932, 890, 880, 798, 786, 753, 733, 669, 649, 636, 629, 557, 522, 513, 410 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.06 (d, *J* = 6.0 Hz, 1H), 7.89-7.88 (m, 2H), 7.74-7.71 (m, 3H), 7.61-7.39 (m, 4H), 6.96-6.84 (m, 4H), 6.68-6.62 (m, 2H), 2.14 (s, 3H), 2.13 (s, 3H). ESI-MS (*m/z*). Calcd for C₄₈H₃₆N₃¹⁹¹Ir [M]⁺: 845.2510. Found: 845.2514.

*mer***-50:** A solution of *mer***-20** (11 mg, 15 µmol) and NBS (3.0 mg, 17 µmol) in CH₂Cl₂ (2) mL) was stirred at room temperature for 1 h. The solvent was removed under reduced pressure and the resulting residue was purified by silica gel chromatography using hexanes/CHCl₃ $(4/1)$ as the eluent to give **mer-50** as a yellow powder (7.1 mg, 64% yield), mp > 300 °C. IR (ATR): $v = 1581, 1552, 1469, 1420, 1391, 1261, 1233, 1157, 1096, 980, 874, 771, 748, 603, 568, 522,$ 428 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.30 (d, *J* = 11 Hz, 1H), 7.98 (dd, *J* = 5.7, 0.9 Hz, 1H), 7.93 (dd, *J* = 5.7, 0.9 Hz, 1H), 7.75 (t, *J* = 5.7 Hz, 3H), 7.66-7.49 (m, 5H), 6.88 (td, *J* = 6.6, 1.2 Hz, 1H), 6.81-6.71 (m, 3H), 6.42-6.34 (m, 2H), 6.30 (s, 1H), 6.23 (s, 1H), 2.16 $(s, 3H)$, 2.14 $(s, 3H)$. ESI-MS (m/z) . Calcd for C₃₅H₂₅N₃F₂⁷⁹Br¹⁹¹Ir [M]⁺: 795.0800. Found: 795.0800.

67a: 67a was obtained as an orange crystaline powder (19 mg, 77% yield) from **23** (33 mg, 40 μmol), ZnBr_2 (0.33 g, 2.4 mmol) and ClCH₂CH₂Cl (2.0 mL) using a precedure similar to that for **34b**. mp > 300 °C. IR (ATR): $v = 1709$, 1602, 1571, 1557, 1478, 1403, 1292, 1270, 1245, 1220, 1163, 1147, 1104, 1053, 1042, 989, 869, 834, 815, 785, 753, 737, 713, 674, 567, 528, 418 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.30 (d, *J* = 4.8 Hz, 1H), 9.08 (d, *J* = 6.6 Hz, 1H), 9.00 (d, *J* = 5.4 Hz, 1H), 8.94 (t, *J* = 8.4 Hz, 2H), 8.77 (d, *J* = 6.3 Hz, 1H), 8.30- 8.27 (m, 2H), 8.04 (d, *J* = 7.8 Hz, 1H), 8.00 (d, *J* = 8.1 Hz, 1H), 7.95 (d, *J* = 8.1 Hz, 1H), 7.85- 7.70 (m, 6H), 7.05 (d, *J* = 6.6 Hz, 1H), 6.85 (t, *J* = 6.9 Hz, 1H), 6.73-6.66 (m, 2H), 6.36 (d, *J* = 6.3 Hz, 1H), 6.31-6.27 (m, 4H), 5.81 (s, 1H), 5.69 (s, 1H), 5.43 (dd, *J* = 6.3, 1.8 Hz, 1H), 5.35

(dd, $J = 6.0$, 1.8 Hz, 1H), 2.00 (s, 3H), 1.95 (s, 3H). ESI-MS (m/z). Calcd for C₅₄H₃₆N₄F₄Cl₂Ir₂ [M]⁺: 1272.15061. Found: 1272.1489.

67b: 67b was obtained as an orange crystaline powder (24 mg, 93% yield) from **15** (31 mg, 38 μ mol), ZnBr₂ (0.53 g, 2.3 mmol) and ClCH₂CH₂Cl (2.0 mL) using a precedure similar to that for **34b**. mp > 300 °C. IR (ATR): $v = 1601$, 1588, 1572, 1556, 1500, 1478, 1446, 1430, 1402, 1381, 1349, 1311, 1294, 1270, 1246, 1162, 1148, 1126, 1103, 986, 870, 843, 833, 814, 783, 754, 737, 710, 688, 672, 568, 527 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 9.57 (d, *J* = 5.4 Hz, 1H), 9.34 (d, *J* = 6.6 Hz, 1H), 9.29 (d, *J* = 4.8 Hz, 1H), 9.06 (d, *J* = 6.3 Hz, 1H), 8.92 (t, *J* = 9.0 Hz, 2H), 8.30 (d, *J* = 7.2 Hz, 2H), 8.03-7.99 (m, 3H), 7.90-7.74 (m, 7H), 7.13 (d, *J* = 7.2 Hz, 1H), 6.95-6.89 (m, 1H), 6.72-6.64 (m, 3H), 6.53 (t, *J* = 6.3 Hz, 1H), 6.31-6.23 (m, 2H), 5.79 (s, 1H), 5.68 (s, 1H), 5.42 (dd, *J* = 9.3, 1.5 Hz, 1H), 5.33 (dd, *J* = 9.3, 1.5 Hz, 1H), 2.00 (s, 3H), 1.95 (s, 3H). ESI-MS (m/z). Calcd for C₅₄H₃₇N₄F₄Br₂Ir₂ [M + H]⁺: 1361.0574. Found: 1361.0543.

66a: A mixture of AgOTf (21 mg, 82 μmol), **34b** (14 mg, 12 μmol), the mpiq ligand (25 mg, 0.12 mmol) and triethylamine (15 mg, 0.15 mmol) in ClCH₂CH₂Cl (1.0 mL) was refluxed for 1.5 h. The mixture was then cooled to room temperature, and the precipitate was removed by filtration. The filtrate was evaporated under reduced pressure and purified by silica gel column chromatography using hexanes/CHCl³ as the eluent to afford single stereoisomer **66a** as an orange powder (2.5 mg, 13% yield). mp 206-207 °C. IR (ATR): $v = 1599$, 1580, 1564, 1551, 1470, 1397, 1285, 1263, 1158, 1097, 981, 816, 772, 675, 666, 578, 567, 524, 419 cm⁻¹. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3/\text{Si}(\text{CH}_3)_4)$ $\delta = 8.94 - 8.91$ (m, 1H), 8.19 (d, $J = 7.2$ Hz, 1H), $8.10 - 8.07$ (m, 1H), 7.97 (dd, *J* = 5.1, 0.9 Hz, 1H), 7.82-7.12 (m, 3H), 7.65-7.61 (m, 2H), 7.58-7.54 (m, 1H), 7.51- 7.44 (m, 3H), 7.15 (d, *J* = 6.0 Hz, 1H), 6.89-6.81 (m, 3H), 6.69-6.62 (m, 2H), 6.40-6.32 (m,

2H), 5.88 (dd, *J* = 9.6, 2.4 Hz, 1H), 2.22 (s, 3H), 2.17 (s, 3H). ESI-MS (*m/z*). Calcd for $\rm{C}_{39}\rm{H}_{28}\rm{N}_{3}\rm{F}_{2}^{191}\rm{Ir}$ [M]⁺: 767.1851. Found: 767.1847.

66c: A mixture of AgOTf (0.15 g, 0.58 mmol), **67b** (0.17 g, 0.13 mmol), the tpy ligand (0.16 g, 0.94 mmol) and triethylamine (89 mg, 0.88 mmol) in ClCH₂CH₂Cl (10 mL) was refluxed for 30 min. The mixture was then cooled to room temperature, and the precipitate was removed by filtration. The filtrate was evaporated under reduced pressure and purified by silica gel column chromatography using hexanes/CHCl₃ as the eluent to afford the stereoisomer mixture of 66c and **66d** as a red crystaline powder (0.16 g, 88% yield).

The obtained stereoisomer mixture of **66c** and **66d** (15 mg, 0.19 μmol) was dissolved in $CICH_2CH_2Cl$ (1.0 mL) and silica gel (21 mg) was added. The reaction mixture was reflux for 10 min. The solvent was evaporated under reduced pressure and the residue was purified by silica gel column chromtography with hexanes/CHCl³ as the eluent to afford single isomer **66c** as a red crystaline powder (7.6 mg, 50% yield from the stereoisomers mixture **66c** and **66d**). mp 263-264 °C. IR (ATR): $v = 2915$, 1583, 1565, 1552, 1500, 1468, 1421, 1441, 1395, 1348, 1310, 1285, 1261, 1236, 1208, 1158, 1097, 1038, 979, 808, 772, 749, 739, 713, 671, 569, 523 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.94 (d, *J* = 7.2 Hz, 1H), 8.21-8.15 (m, 3H), 7.89 (d, *J* = 8.7 Hz, 1H), 7.83 (d, *J* = 5.4 Hz, 1H), 7.70-7.65 (m, 1H), 7.62-7.54 (m, 5H), 7.41 (d, *J* = 6.0 H, 1H), 6.97 (d, *J* = 6.3 H, 1H), 6.89 (t, *J* = 5.7 Hz, 2H), 6.84 (d, *J* = 5.1 H, 1H), 6.78-6.76 (m, 2H), 6.53 (s, 1H), 6.39-6.31 (m, 2H), 5.90 (dd, *J* = 7.5, 2.1 Hz, 1H), 2.22 (s, 3H), 2.09 (s, 3H). ESI-MS (*m/z*). Calcd for C₃₉H₂₈N₃F₂¹⁹¹Ir [M]⁺: 767.1851. Found: 767.1860.

Ir(tpy)(F2ppy)(Br-mpiq): The obtained stereoisomer mixture of **66c** (44 mg, 57 μmol) was dissolved in CH_2Cl_2 (12 mL) and NBS (11 mg, 63 µmol) was added. The reaction mixture was stirred at room temperature for 10 min. The solvent was evaporated under reduced pressure

and the residue was purified by silica gel column chromtography with hexanes/CHCl₃ as the eluent to afford **70** as a red powder (43 mg, 87% yield). mp 248-249 °C. IR (ATR): $v = 2920$. 1595, 1469, 1396, 1286, 1262, 1237, 1159, 1098, 1059, 980, 809, 773, 749, 732, 569, 523, 415 cm⁻¹. ¹H NMR (300 MHz, CDCl₃/Si(CH₃)₄) δ = 8.88 (d, *J* = 8.1 Hz, 1H), 8.44 (s, 1H), 8.21 (d, *J* = 8.1 Hz, 1H), 8.13 (d, *J* = 6.0 Hz, 1H), 7.89 (d, *J* = 7.8 Hz, 1H), 7.82 (d, *J* = 5.7 Hz, 1H), 7.71 (t, *J* = 9.0 Hz, 2H), 7.65-7.55 (m, 4H), 7.43 (d, *J* = 6.3 Hz, 1H), 7.01 (d, *J* = 6.3 Hz, 1H), 6.90 (t, *J* = 7.2 Hz, 1H), 6.80-6.75 (m, 2H), 6.57 (s, 1H), 6.42-6.34 (m, 2H), 5.89 (ddd, *J* = 7.5, 2.1, 1.2 Hz, 1H), 2.25(s, 3H), 2.11 (s, 3H). ESI-MS (m/z). Calcd for C₃₉H₂₇N₃F₂⁷⁹Br¹⁹¹Ir [M]⁺: 845.0956. Found: 845.0961.

 74: Phosphorus oxychloride (0.5 mL) was added dropwise to DMF (4.8 mL), and the resulting mixture was stirred at room temperature for 30 min, after which *fac***-20** (140 mg, 0.16 mmol) was added to produce a yellow solution. After stirring overnight at 80° C, the deep-red colored reaction mixture was allowed to cool at 0° C, and 3 M aqueous NaOH (10 mL) was then added. After stirring at room temperature for 1 h, the yellow solid was isolated by filtration and washed with water to afford **74** as a bright yellow powder (60 mg, 53%). IR (ATR): $v = 1671$, 1577, 1559, 1524, 1472, 1421, 1398, 1286, 1240, 1205, 1162, 1099, 1066, 1033, 1021, 983, 929, 887, 859, 829, 783, 746, 718, 665, 566, 523 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/TMS): δ = 10.20 (s, 1H), 10.17 (s, 1H), 8.30 (d, *J* = 8.8 Hz, 1H), 8.07 (d, *J* = 7.6 Hz, 2H), 8.01 (d, *J* = 8.4 Hz, 2H), 7.71-7.67 (m, 3H), 7.46-7.41 (m, 3H), 7.09-6.92 (m, 3H), 6.78 (s, 1H), 6.65 (s, 1H), 6.43-6.38 (m, 1H), 6.23 (dd, *J* = 9.0, 2.1 Hz, 1H), 2.48 (s, 3H), 2.41 (s, 3H). ¹³C NMR (100 MHz, CDCl3/TMS): *δ* = 173.6, 166.0, 165.8, 163.7, 147.5, 143.5, 142.6, 140.6, 137.8, 129.0, 128.8, 126.5, 123.8, 123.3, 123.3, 122.8, 120.0, 118.7, 97.28, 19.6, 19.5. ESI-MS (*m/z*). Calcd for $C_{37}H_{26}N_3O_2F_2^{191}$ Ir [M]⁺: 775.1622; found: 775.1680.

75: A mixture of NaClO₂ (430 mg, 4.7 mmol) and NaH₂PO₄·2H₂O (465 mg, 2.9 mmol) in water (12 mL) was added dropwise to a solution of **74** (82 mg, 106 µmol) and 2-methyl-2butene (500 µL, 4.7 mmol) in DMSO (3 mL) at room temperature. After stirring at room temperature for 5 h, the mixture was concentrated under reduced pressure, and 1 M aqueous HCl was then added until pH 1. The resulting solid was filtered off and washed with water and CHCl₃ to afford **75** as a yellow powder (58 mg, 69% yield). IR (ATR): $v = 2916$, 1671, 1597, 1583, 1559, 1524, 1472, 1421, 1400, 1284, 1240, 1160, 1098, 1069, 1032, 1017, 984, 910, 883, 861, 827, 778, 750, 718, 626, 567, 524 cm-1 . ¹H NMR (400 MHz, *d6*-DMSO/TMS): *δ* = 12.31 (brs, 2H), 8.31-8.17 (m, 5H), 7.95-7.85 (m, 3H), 7.53 (d, *J* = 6.3 Hz, 2H), 7.44 (d, *J* = 5.7 Hz, 1H), 7.25-7.17 (m, 3H), 6.66 (t, *J* = 9.3 Hz, 1H), 6.54 (s, 1H), 6.52 (s, 1H), 6.13 (dd, *J* = 9.3, 2.1 Hz, 1H), 2.28 (s, 3H), 2.23 (s, 3H). ¹³C NMR (100 MHz, d_6 -DMSO/TMS): δ = 169.6, 167.8, 167.4, 164.7, 164.5, 161.8, 147.8, 147.4, 142.8, 140.5, 139.5, 138.6, 128.0, 126.5, 123.9, 123.4, 122.9, 122.7, 120.0, 117.6, 96.6, 22.8, 22.7. ESI-MS (m/z) . Calcd for C₃₇H₂₆N₃O₄F₂¹⁹¹Ir [M]⁺: 807.1521; found: 807.1513.

 76: DIEA (0.10 g, 0.74 mmol), PyBOP (0.29 g, 0.56 mmol), and mono-Boc-protected octamethylenediamine (0.11 g, 0.42 mmol) were added to a solution of **75** (76 mg, 95 μmol) in freshly distilled DMF (1 mL). The reaction mixture was stirred overnight at room temperature, and then concentrated under reduced pressure. The remaining residue was purified by silica gel column chromatography (CHCl₃) and gel permeation chromatography (CHCl₃) to afford the Boc protected **76** (40 mg, 33%) as a yellow powder. R_f value of TLC (CHCl₃): 0.58. IR $(ATR): v = 2926, 2855, 1693, 1634, 1598, 1556, 1512, 1471, 1424, 1398, 1364, 1248, 1161,$ 1109,1097, 1068, 1029, 1012, 982, 887, 863, 828, 809, 780, 749, 718, 694, 672, 566, 524 cm-¹. ¹H NMR (400 MHz, CDCl₃/TMS): δ = 8.25 (d, *J* = 9.0 Hz, 1H), 7.87 (t, *J* =5.4 Hz, 2H), 7.69-7.62 (m, 5H), 7.45 (d, *J* = 5.1 Hz, 1H), 7.41 (d, *J* = 5.1 Hz, 1H), 7.38 (d, *J* = 5.1 Hz, 1H), 6.92-6.88 (m, 3H), 6.64 (s, 1H), 6.63 (s, 1H), 6.36-6.32 (m, 2H), 4.52 (brs, 2H), 3.49-3.40 (m,

4H), 3.10-3.08 (m, 4H), 2.26 (s, 3H), 2.22 (s, 3H), 1.46 (s, 18H), 1.31-1.20 (m, 24H). ¹³C NMR (100 MHz, CDCl3/TMS): *δ* = 172.1, 166.5, 164.1, 156.8, 147.7, 142.2, 139.9, 137.5, 137.1, 129.6, 128.1, 123.4, 122.6, 96.6, 79.4, 40.7, 40.0, 30.1, 29.8, 29.3, 28.5, 27.1, 26.8, 20.6, 20.4. ESI-MS (m/z). Calcd for C₆₃H₇₉N₇O₆F₂¹⁹¹Ir [M + H]⁺: 1260.5689; found: 1260.5696.

 77: A mixture of TMSCl (130 µL, 1.03 mmol) and NaI (154 mg, 1.03 mmol) in CH3CN (2.0 mL) was added to a suspension of Boc protected **76** (65 mg, 0.05 mmol) in CH₃CN (3.0 mL). After stirring the mixture at room temperature for 20 min, it was sonicated for 2 min. The insoluble compound was isolated by centrifugation and washed with CH3CN to give **77** as the HI salt (49 mg, 72%). IR (ATR): $v = 2927, 2856, 1598, 1555, 1529, 1471, 1424, 1398, 1285$ 1264, 1240, 1160, 1097, 1068, 983, 863, 830, 784, 750, 720, 567, 517 cm⁻¹. ¹H NMR (400 MHz, CD3OD/TMS): *δ* = 8.30 (dd, *J* = 12.6, 5.2 Hz, 1H), 8.09-8.01 (m, 2H), 7.83-7.72 (m, 5H), 7.58-45 (m, 3H) 7.08-6.95 (m, 3H), 6.68 (s, 1H), 6.61 (s, 1H), 6.45-6.38 (m, 1H), 6.35-6.29 (m, 1H), 2.91 (t, *J* = 4.0 Hz, 4H), 2.18 (s, 3H), 2.12 (s, 3H), 1.69-1.61 (m, 10H), 1.47-1.43 (m, 18H). ESI-MS (m/z) . Calcd for C₅₃H₆₃N₇O₂F₂¹⁹¹Ir [M + H]⁺: 1060.4641; found: 1060.4630. Anal. Calcd for $C_{85}H_{114}F_{4}IrN_{13}O_{14}$: C, 60.08; H, 6.24; N, 7.78%; found: C, 60.23; H, 6.42; N, 7.97%.

 72: DIEA (44 μL, 0.25 mmol), PyBop (113 mg, 0.21 mmol), and the protected KKKGG peptide (54 mg, 59 μmol) were added to a solution of **77** (26 mg, 20 μmol) in dist. DMF (1 mL). The reaction mixture was stirred overnight at room temperature and then concentrated under reduced pressure and purified by silica gel column chromatography $\text{CHCl}_3/\text{MeOH} = 1/0$ to 10/1) to afford the protected Ir complex R_f value of TLC (CHCl₃/MeOH (10/1)): 0.70. To the protected Ir complex in CH₂CN (2.0 mL) was added a mixture of TMSCl (62 μ L, 0.49 mmol) and NaI (73 mg, 0.49 mmol) in CH₃CN (3.0 mL). The reaction mixture was stirred at room temperature for 20 min and then sonicated for 2 min. The insoluble compound was isolated by

centrifugation and washed with CH₃CN to give 69 as the HI salt. The product was purified by preparative RP-HPLC [H₂O (0.1% TFA)/CH₃CN (0.1% TFA) = 80/20 to 40/60 (30 min), t_r = 17 min, 8.0 mL/min], lyophilized to give **72** as a yellow powder (6.2 mg, 29% as 8TFA salt). IR (ATR): = 3061, 2929, 1662, 1599, 1565, 1553, 1470, 1420, 1397, 1282, 1267, 1236, 1196, 1183, 1132, 1095, 1067, 1035, 980, 863, 837, 826, 798, 785, 750, 723, 568, 523, 419, 405 cm-¹. ¹H NMR (400 MHz, CD₃OD/TMS): δ = 8.19 (d, *J* = 8.4 Hz, 2H), 8.03-7.88 (m, 3H), 7.71-7.63 (m, 3H), 7.47-7.36 (m, 3H), 6.97-6.88 (m, 3H), 6.59 (s, 1H), 6.52 (s, 1H), 6.32-6.26 (m, 1H), 6.16-6.13 (m, 1H), 4.28-4.20 (m, 2H), 4.18-4.16 (m, 2H), 3.87-3.80 (m, 14H), 3.77-3.20 (m, 12H), 2.08 (s, 3H), 2.02 (s, 3H), 1.80-1.25 (m, 64H). ESI-MS (*m/z*). Calcd for $C_{97}H_{148}N_{23}O_{12}F_2^{191}$ Ir $[M + 2H]^{2+}$: 1029.0637; found: 1029.0638.

 78: Phosphorus oxychloride (0.25 mL) was added dropwise to dimethylformamide (2 mL) solution and the resulting reaction mixture was stirred at room temperature for 30 min, after which *fac***-21** (55 mg, 74 μmol) was added to yield a yellow color solution. After stirring the reaction mixture at 80 $^{\circ}$ C for 14 h, the deep-red colored reaction mixture was allowed to cool to 0° C, and 2° M aqueous NaOH (6.0 mL) was then added whereupon the product was precipitated as a yellow precipitate. The yellow colored Ir complex was isolated by filtration and washed with water to afford **78** as a yellow powder (54 mg, 94%). IR (ATR): $v = 1679$, 1595, 1566, 1556, 1472, 1396, 1283, 1239, 1208, 1161, 1098, 1067, 984, 857, 823, 784, 747, 716, 567, 523, 500, 432 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/TMS): δ = 10.18 (s, 1H), 8.31 (d, *J* = 8.4 Hz, 2H), 8.07 (s, 1H), 8.02 (d, *J* = 8.1 Hz, 1H), 7.74-7.65 (m, 3H), 7.48-7.43 (m, 3H), 6.99-6.88 (m, 3H), 6.73 (s, 1H), 6.42-6.28 (m, 3H), 6.18 (dd, *J* = 8.4, 2.4 Hz, 1H), 2.47 (s, 3H) ppm. ESI-MS (*m/z*). Calcd for C₃₅H₂₂N₃OF₄¹⁹¹Ir [M]⁺: 769.1328; found: 769.1322.

79: A mixture of NaClO₂ (190 mg, 2.1 mmol) and NaH₂PO₄·2H₂O (328 mg, 2.1 mmol) in water (10 mL) was added dropwise to a solution of **78** (65 mg, 85 μmol) and 2-methyl-2 butene (230 μ L, 2.3 mmol) in DMSO (8 mL) at room temperature. After stirring for 5 h at room temperature, the mixture was concentrated under reduced pressure followed by the addtion of 1 M aqueous HCl to pH 1 to precipitate the yellow solid product. The solid was isolated by filtration and washed with water to obtain **79** as a yellow powder (49 mg, 74%). IR $(ATR): v = 1718, 1678, 1596, 1557, 1472, 1421, 1398, 1283, 1238, 1160, 1098, 983, 860, 822,$ 777, 750, 718, 567, 524, 427 cm⁻¹. ¹H NMR (400 MHz, DMSO- d_6 /TMS): δ = 12.34 (brs, 1H), 8.28-8.24 (m, 3H), 8.20 (d, *J* = 9.6 Hz, 1H), 7.96-7.86 (m, 3H), 7.54 (d, *J* = 5.1 Hz, 2H), 7.45 (d, *J* = 5.7 Hz, 1H), 7.27-7.21 (m, 3H), 6.72-6.66 (m, 2H), 6.49 (s, 1H), 6.11-6.05 (m, 2H), 2.27 (s, 3H). ESI-MS (m/z) . Calcd for C₃₅H₂₂N₃O₂F₄¹⁹¹Ir [M]⁺: 785.1277; found: 785.1272.

 80: DIEA (30 mg, 0.23 mmol), PyBOP (0.10 g, 0.19 mmol), and mono-Boc-protected octamethylenediamine (22 mg, 81 μmol) were added to a solution of **79** (32 mg, 41 μmol) in freshly distilled DMF (1 mL). The reaction mixture was stirred overnight at room temperature, and then concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (CHCl₃) and gel permeation chromatography (CHCl₃) to afford the Boc protected 80 (36 mg, 86%) as a yellow powder. R_f value of TLC (CHCl₃): 0.91. IR (ATR): = 3279, 2927, 2856, 1704, 1637, 1598, 1567, 1557, 1473, 1424, 1398, 1365, 1284, 1266, 1240, 1163, 1099, 984, 860, 824, 787, 752, 718, 567, 525 cm⁻¹. ¹H NMR (400 MHz, CDCl₃/TMS): δ = 8.27 (t, *J* = 8.1 Hz, 2H), 7.85 (d, *J* = 5.4 Hz, 1H), 7.68-7.61 (m, 4H), 7.47 (d, *J* = 5.1 Hz, 1H), 7.43 (d, *J* = 5.7 Hz, 1H), 7.40 (d, *J* = 5.7 Hz, 1H), 6.93-6.85 (m, 3H), 6.59 (s, 1H), 6.46-6.32 (m, 2H), 6.30 (dt, *J* = 9.3, 3.0 Hz, 2H), 4.56 (brs, 1H), 3.50-3.40 (m, 2H), 3.10- 3.07 (m, 2H), 2.23 (s, 3H), 1.43 (s, 9H), 1.34-1.20 (m, 12H). ¹³C NMR (100 MHz, CDCl3/TMS): *δ* = 171.2, 165.7, 163.5, 163.2, 156.0, 146.9, 141.5, 139.2, 136.7, 136.4, 129.3, 128.9, 127.4, 122.7, 122.0, 118.9, 118.2, 96.0, 79.0, 40.5, 39.8, 30.0, 29.2, 29.0, 28.4, 26.9,

26.7, 20.5, 20.3 ppm. ESI-MS (m/z). Calcd for C₄₈H₄₉N₅O₃F₄¹⁹¹Ir [M + H]⁺: 1012.3323; found: 1012.3391.

 81: 80 (19 mg, 19 μmol) was dissolved in 2.0 mL of ethyl acetate and treated with 1.0 mL of 2 M aqueous HCl. After stirring at 60 ºC for 4 h, the mixture was concentrated under reduced pressure and the residue was purified by silica gel column chromatography $CHCl₃/MeOH$ = $10/1$) to afford 81 as a yellow powder (16 mg, 93% yield as HCl salt). R_f value of TLC $(CHCl₃/MeOH (10/1))$: 0.87. IR (ATR): $v = 2926$, 2855, 1596, 1566, 1556, 1471, 1422, 1397, 1283, 1264, 1238, 1161, 1098, 1067, 982, 860, 823, 785, 750, 717, 566, 524 cm⁻¹. ¹H NMR $(400 \text{ MHz}, \text{CD}_3 \text{OD/TMS})$: $\delta = 8.32 - 8.26 \text{ (m, 2H)}$, $8.07 \text{ (d, } J = 7.8 \text{ Hz}, 1H)$, $7.81 - 7.77 \text{ (m, 3H)}$, 7.73 (s, 1H), 7.61 (d, *J* = 4.5 Hz, 1H), 7.55 (d, *J* = 6.9 Hz, 1H), 7.51 (d, *J* = 4.5 Hz, 1H), 7.10- 7.00 (m, 3H), 6.59 (s, 1H), 6.50-6.41 (m, 2H), 6.21 (dd, *J* = 9.3, 2.4 Hz, 1H), 6.15 (dd, *J* = 9.3, 2.4 Hz, 1H), 2.90 (t, *J* = 7.5 Hz, 2H), 2.17 (s, 3H), 1.66-1.58 (m, 4H), 1.41-1.26 (m, 11H). ¹³C NMR (100 MHz, CDCl3/TMS): *δ* = 173.9, 166.2, 163.4, 148.1, 147.9, 147.8, 142.6, 139.1, 138.1, 137.9, 137.2, 130.3, 128.3, 123.1, 122.9, 119.6, 118.0, 96.1, 39.6, 39.5, 29.1, 28.8, 27.3, 26.6, 26.1, 18.8 ppm. ESI-MS (m/z) . Calcd for C₄₃H₄₁N₅OF₄¹⁹¹Ir [M + H]⁺: 912.2876; found: 912.2855.

 82: DIEA (11 mg, 86 μmol), PyBop (31 mg, 55 μmol), and protected KKKGG peptide (31 mg, 33 μmol) were added to a solution of **81** (11 mg, 11 μmol) in dist. DMF (1 mL). The reaction mixture was stirred at room temperature for 23 h. The reaction mixture was concentrated under reduced pressure and resulting residue was purified by silica gel column chromatography (CHCl₃/MeOH = $1/0$ to $10/1$) and gel permeation chromatography (CHCl₃) to afford **82** as a yellow powder (18 mg, 89%). R_f value of TLC (CHCl₃/MeOH (10/1)): 0.62. IR (ATR): $v = 3289, 3072, 2931, 2861, 1692, 1628, 1599, 1555, 1513, 1473, 1399, 1365, 1283,$ 1245, 1164, 1099, 1067, 984, 862, 834, 824, 782, 752, 718, 567, 524 cm⁻¹. ¹H NMR (400 MHz,

CDCl3/TMS): *δ* = 8.29 (t, *J* = 8.1 Hz, 2H), 7.90 (d, *J* = 7.2 Hz, 1H), 7.68-7.65 (m, 3H), 7.47 (d, *J* = 6.0 Hz, 1H), 7.44 (d, *J* = 5.4 Hz, 1H), 7.40 (d, *J* = 6.9 Hz, 1H), 6.95-6.89 (m, 3H), 6.60 (s, 1H), 6.38-6.30 (m, 2H), 6.29 (d, *J* = 9.0 Hz, 3H), 4.90-4.74 (m, 3H), 4.08-3.99 (m, 15H), 3.16- 3.03 (m, 12H), 2.24 (s, 3H), 1.66-1.27 (m, 62H). ESI-MS (m/z). Calcd for C₈₅H₁₁₅N₁₃F₄O₁₄¹⁹¹Ir $[M + H]^+$: 1832.8252; found: 1810.8264. Anal. Calcd for $C_{85}H_{114}F_{4}IrN_{13}O_{14}$ and 2.8CHCl₃: C, 49.18; H, 5.49; N, 8.49%; found: C, 49.01; H, 5.43; N, 8.66%.

73: A mixture of TMSCl (0.34 g, 3.2 mmol) and NaI (0.40 g, 2.7 mmol) in CH₂CN (2.0 mL) was added to a mixture of 82 (18 mg, 10 μ mol) in CH₃CN (3.0 mL). The mixture was stirred at room temperature for 10 min and sonicated for 2 min. The insoluble compound was centrifuged and washed with CH_2CN to give 73 as the HI salt. The product was purified by preparative RP-HPLC [H₂O (0.1% TFA)/CH₃CN (0.1% TFA) = 80/20 to 30/70 (30 min), t_r = 26 min, 8.0 mL/min], and lyophilized to give **73** as a yellow powder (10 mg, 56% as 4TFA salt). IR (ATR): $v = 1599, 1580, 1564, 1551, 1470, 1397, 1285, 1263, 1158, 1097, 981, 816,$ 772, 675, 666, 578, 567, 524, 419 cm⁻¹. ¹H NMR (400 MHz, CD₃OD/TMS): δ = 8.30 (t, *J* = 8.7 Hz, 2H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.84-7.77 (m, 3H), 7.73 (s, 1H), 7.61 (d, *J* = 4.2 Hz, 1H), 7.55 (d, *J* = 4.8 Hz, 1H), 7.51 (d, *J* = 4.8 Hz, 1H), 7.08-7.00 (m, 3H), 6.59 (s, 1H), 6.48-6.37 (m, 2H), 6.21 (dd, *J* = 9.0, 2.7 Hz, 1H), 6.16 (dd, *J* = 9.0, 2.4 Hz, 1H), 4.38-4.32 (m, 1H), 4.29- 4.25 (m, 1H), 3.96-3.83 (m, 5H), 3.21-3.15 (m, 2H), 2.96-2.91 (m, 6H), 2.17 (s, 3H), 1.87-1.82 (m, 4H), 1.71-1.64 (m, 8H), 1.59-1.51 (m, 10H), 1.45-1.35 (m, 10H). ESI-MS (*m/z*). Calcd for $C_{65}H_{83}N_{13}F_4O_6^{191}$ Ir [M + H]⁺: 1410.6155; found: 1410.6160.

X-ray Data Collection and Refinement

All crystalline samples of the Ir complexes were recrystallized from hexanes/CH₂Cl₂ (for **15b**, *fac*-20, *mer*-20, *mer*-23, *mer*-24, 29b, and 42a), hexanes/CHCl₃ (for *mer*-50), methanol/CH2Cl² (for **66c**) or CHCl3/DMSO (for **74** and **79**) to give crystals suitable for singlecrystal X-ray analysis. Single-crystal X-ray studies were performed on a Bruker APEX CCD diffractometer equipped with a Bruker Instruments low-temperature attachment. Data were collected at 103 K (for **34b**, **42a**, and **66c**), 123 K (for **74**), 173 K (for *fac***-20**, *mer***-20**, *mer***-23**, *mer***-24**, and *mer***-50**), or 296K (for **15b** and **74**) using graphite-monochromated Mo−K α radiation ($\lambda = 0.71073$ Å). The frames were indexed, integrated, and scaled using the SMART and SAINT software packages.⁸⁹ An empirical absorption correction was applied to the collected reflections with SADABS⁹⁰ using XPREP.⁹¹ All of the structures were solved by the direct method using the program SHELXS-97 and were refined on F^2 by the full-matrix leastsquares technique using the SHELXL-97 program package.⁹² All non-hydrogen atoms were refined anisotropically in the structure. Hydrogen atoms of **34b** were placed in calculated positions by means of the 'riding' model. Disorder problems were found during the refinement of **34b** and solved using the split model. All crystal data in this manuscript can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.summary.ccdc.cam.ac.uk/structure-summary-form.

Crystal Data for 15b. C₄₈H₄₀Br₂Ir₂N₄, $Mr = 1217.06$, monoclinic, $P2_1/n$, $a = 11.6206(6)$ $b =$ 14.8110(7), $c = 25.0132(13)$ Å, $\beta = 96.241(2)$, $V = 4279.6(4)$ Å³, $Z = 4$, $\rho_{calc} = 1.889$ g.cm⁻³, R $= 0.0650$ (for 5375 reflection with $I > 2\sigma(I)$), $Rw = 0.1570$ (for 7525 reflections), GOF = 1.139. CCDC 1501488 contains the supplementary crystallographic data for the paper.

Crystal Data for *fac***-20**. C₃₅H₂₆F₂IrN₃, *Mr* = 718.79, monoclinic, *C*2/*c*, *a* = 34.105(16) *b* = 9.227(4), $c = 17.928(8)$ Å, $\beta = 99.7100(2)$, $V = 5561(5)$ Å³, $Z = 8$, $\rho_{calc} = 1.717$ g.cm⁻³, $R =$ 0.0484 (for 5437 reflection with $I > 2\sigma(I)$), $R_W = 0.1094$ (for 6249 reflections), GOF = 1.137. CCDC 1501443 contains the supplementary crystallographic data for the paper.

Crystal Data for *mer***-20**. C₃₅H₂₆F₂IrN₃⋅CH₂Cl₂, $Mr = 803.71$, monoclinic, $P2/c$, $a =$ $19.6105(16)$ $b = 8.6627(7)$, $c = 17.9826(15)$ Å, $\beta = 96.5980(1)$, $V = 3024.6(4)$ Å³, $Z = 4$, $\rho_{calc} =$ 1.759 g.cm⁻³, $R = 0.0366$ (for 5970 reflection with $I > 2\sigma(I)$), $Rw = 0.0972$ (for 6870 reflections), $GOF = 1.042$. CCDC 1502134 contains the supplementary crystallographic data for the paper.

Crystal Data for *mer***-23**. C₄₃H₃₀F₂IrN₃⋅CH₂Cl₂, *Mr* = 903.83, triclinic, *P*1, *a* = 10.878(3) *b* = 12.483(4), $c = 13.740(4)$ Å, $\alpha = 76.509(5)$, $\beta = 73.212(5)$, $\gamma = 89.340(6)$ \degree , $V = 1733.9(9)$ Å \degree , Z $= 2, \rho_{calc} = 1.731$ g.cm⁻³, $R = 0.0582$ (for 5600 reflection with $I > 2\sigma(I)$), $Rw = 0.1103$ (for 6426) reflections), GOF = 1.140 . CCDC 1501444 contains the supplementary crystallographic data for the paper.

Crystal Data for *mer***-24**. C₃₈H₂₄F₄IrN₃, *Mr* = 790.08, monoclinic, *P*2₁/*c*, *a* = 13.336(2) *b* = $10.9986(18)$, $c = 20.398(3)$ Å, $\beta = 101.168(3)$, $V = 2934.0(8)$ Å³, $Z = 4$, $\rho_{calc} = 1.790$ g.cm⁻³, R $= 0.0399$ (for 4794 reflections with $I > 2\sigma(I)$), $Rw = 0.0770$ (for 5331 reflections), GOF = 1.099. CCDC 1817976 contains the supplementary crystallographic data for the paper.

Crystal Data for 34b. C₄₇H₃₂Br₂Cl_{1.79}F₄Ir₂N₄O₂, *Mr* =1370.48, tetragonal, *I*⁴₁*cd*, *a* = 16.8132(19) $b = 16.8132(19)$, $c = 31.087(3)$ Å, $V = 8788(2)$ Å³, $Z = 8$, $\rho_{calc} = 2.072$ g.cm⁻³, $R =$ 0.0360 (for 4623 reflections with $I > 2\sigma(I)$), $Rw = 0.0799$ (for 5158 reflections), GOF = 1.094. CCDC 1817974 contains the supplementary crystallographic data for the paper.

Crystal Data for 42a. $C_{38}H_{27}F_{2}IrN_4O_2S$, $Mr = 833.90$, triclinic, $P\bar{1}$, $a = 10.215(5) b = 10.959(5)$, $c = 16.413(5)$ Å, $\alpha = 80.353(5)$, $\beta = 86.921(5)$, $\gamma = 71.791(5)$ \degree , $V = 1720.7(13)$ Å³, $Z = 2$, ρ_{calc} $= 1.610$ g.cm⁻³, $R = 0.0623$ (for 5131 reflection with $I > 2\sigma(I)$), $Rw = 0.1380$ (for 6240)

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reflections), GOF = 1.050 . CCDC 1501455 contains the supplementary crystallographic data for the paper.

Crystal Data for *mer***-50**. C₃₅H₂₅BrF₂IrN₃⋅1.5H₂O, *Mr* = 821.69, triclinic, *P*¹, *a* = 9.5171(7) *b* $= 12.9204(9)$, $c = 13.0846(10)$ Å, $\alpha = 70.745(1)$, $\beta = 86.152(1)$, $\gamma = 84.710(1)$ \degree , $V = 1511.32(19)$ \mathring{A}^3 , $Z = 2$, $\rho_{calc} = 1.806$ g.cm⁻³, $R = 0.0311$ (for 5767 reflection with $I > 2\sigma(I)$), $Rw = 0.0893$ (for 5992 reflections), GOF = 1.066. CCDC 1504245 contains the supplementary crystallographic data for the paper.

Crystal Data for 66c. C₃₉H₂₈F₂IrN₃, $Mr = 786.84$, monoclinic, $P2_1/c$, $a = 13.0751(19)$ $b =$ 14.971(2), $c = 15.705(2)$ Å, $\beta = 95.642(2)$, $V = 3059.3(7)$ Å³, $Z = 4$, $\rho_{calc} = 1.669$ g.cm⁻³, $R =$ 0.0424 (for 4298 reflections with $I > 2\sigma(I)$), $R_W = 0.1120$ (for 5390 reflections), GOF = 1.059. CCDC 1817970 contains the supplementary crystallographic data for the paper.

Crystal Data for 74. $C_{37}H_{26}F_{2}IrN_{3}O_{2}$; FW = 774.81, triclinic, space group *P*-1, $a = 11.6449(2)$ Å, $b = 12.0042(1)$ Å, $c = 15.4338(2)$ Å, $\alpha = 81.793(1)$ °, $\beta = 84.931(1)$ °, $\gamma = 74.780(1)$ °, $V =$ 2057.54(5) \mathring{A}^3 , $Z = 2$, $D_{\text{caled}} = 1.251$ g cm⁻³; $R_1 = 0.0327$ (for 11234 reflections with $I > 2$ s (*I*)), wR_2 = 0.1313 (for 50126 reflections), GOF = 0.585. CCDC 2004202 contains the supplementary crystallographic data for the paper.

Crystal Data for 79. C₃₅H₂₂F₄IrN₃O₂; FW = 784.75, monoclinic, space group *C*2/*c*, $a =$ 44.975(7) Å, $b = 9.0309(13)$ Å, $c = 18.118(3)$ Å, $\alpha = \gamma = 90^{\circ}$, $\beta = 92.504(2)^{\circ}$, $V = 7352(2)$ Å³, $Z = 4$, $D_{\text{caled}} = 0.709$ g cm⁻³; $R_1 = 0.0383$ (for 8397 reflections with $I > 2$ s (*I*)), $wR_2 = 0.1162$ (for 27902 reflections), GOF = 0.791 . CCDC 2055929 contains the supplementary crystallographic data for the paper.

Measurements of UV/vis Absorption and Luminescence Spectra.

UV/vis spectra were recorded on a JASCO V-550 UV/vis spectrophotometer and emission spectra were recorded at 25 °C on a JASCO FP-6200 and FP-6500 spectrofluorometer, respectively. All samples for spectroscopic measurements were carefully purified by recrystalization twice prior to use in order to minimize contamination and the same spectra were observed after recrystallization. Sample solutions in quartz cuvettes equipped with teflon septum screw caps were degassed by bubbling Ar through the solution for 10 min prior to conducting the luminescence measurements. Phosphorescence quantum yields (*Φ*) were determined using quinine sulfate (Φ = 0.55 in 0.1 M H_2SO_4 ⁹³ or *fac*-Ir(mpiq)₃ (*fac*-17) $(\Phi = 0.26$ in toluene).⁵⁹ Equation (1) was used to calculate the emission quantum yields, in which Φ_s and Φ_t denote the quantum yields of the sample and the reference compound, respectively. The η_s and η_r terms are the refractive indexes of the solvents used for the measurements of the sample and the reference $(\eta: 1.477$ for DMSO, 1.333 for H₂O, 1.422 for CH_2Cl_2 , and 1.497 for toluene). The A_s and A_r terms are the absorbance of the sample and the reference, and the I^s and I^r terms stand for the integrated areas under the emission spectra of the sample and reference, respectively (all of the Ir compounds were excited at 366 nm for luminescence measurements in this study).

$$
\varPhi_{\rm s} = \varPhi_{\rm f}(\eta_{\rm s}^2 A_{\rm r} I_{\rm s})/(\eta_{\rm r}^2 A_{\rm s} I_{\rm r})\tag{1}
$$

The luminescence lifetimes of sample solutions of Ir(III) complexes in degassed DMSO at 298 K were measured on a TSP1000-M-PL (Unisoku, Osaka, Japan) instrument by using THG (355 nm) of Nd: YAG laser, Minilite I (Continuum, CA, USA) as excitation source. The signals were monitored with an R2949 photomultiplier. Data were analyzed using a single exponential decay equation (for *mer***-20-24**, *fac***-20**, **6**, **25**, **29b**, **35**-**38, 66a,c**, **66a**, and **67**) or a biexponential decay equation (for **41**, **42a**, and **43**). Sample solutions in quartz cuvettes equipped with teflon septum screw caps were degassed by bubbling Ar through the solution for 20 minutes prior to measuring the lifetime.

Cell Culture

 Cancer (Jurkat, HeLa S3, and A549) and normal IMR90 cells were cultured in RPMI 1640 (Jurkat and IMR-90), MEM (HeLa S3), or DMEM (A549) medium supplemented with 10% fetal bovine serum (FBS), L-glutamine, HEPES (2-[4-(2-hydroxyethyl)-1 piperazinyl]ethanesulfonic acid, pH = 7.5), penicillin/streptomycin, and monothioglycerol (MTG) in a humidified 5% $CO₂$ incubator at 37 °C.

MTT assay

Jurkat (2.0 \times 10⁵ cells/mL), HeLa S3, A549 cancer cells, and IMR90 normal cells (1.5 \times 10⁵ cells/mL) cells were incubated in 10% FBS RPMI 1640 medium, MEM, or DMEM medium (100 μ L) containing the Ir complexes (**71–73**) under 5% CO₂ at 37 °C for 24 h in 96 well plates (BD Falcon), and then 0.5% MTT reagent in PBS (10 μL) was then added to the cells. After incubation at 37 ºC for 4 h, a formazan lysis solution (10% SDS in 0.01N HCl, 100 μL) was added and the resulting solution was incubated overnight under the same conditions, followed by measurement of the absorbance at 570 nm using a microplate reader (BIO-RAD).

Fluorescent Microscopic Studies of Jurkat Cells with Ir Complexes.

Jurkat cells $(1.0 \times 10^5 \text{ cells})$ were incubated without or with Ir complexes **71–73** (25 μM) in RPMI 1640 medium containing 10% FBS for 30 min (100 μ L) and 1 h under 5% CO₂ at 37 °C. After the incubation, the cells were washed twice with ice-cold PBS containing 0.1 % NaN3

and 0.5% FBS, and observed on fluorescent microscopy (Fluoview, FV-1000, Olympus) by using Greiner CELLviewTM petri dish (35 \times 10 mm). Excitation was 405 nm for Ir complexes.

Microscopic Observations of Jurkat Cells Stained with Rhod-4 or Rhod-2

Jurkat cells $(2.0\times10^5 \text{ cells})$ were incubated with Rhod-2/AM (5 μ M) or Rhod-4/AM (5 μ M) in RPMI 1640 medium containing 10% FBS (100 μ L) at 37 °C for 30 min, respectively. The cells were collected by centrifuge (2000rpm at 4° C, 3 min), wash with PBS, and then incubated in RPMI 1640 medium containing 10% FBS (100 μ L) in the presence of **71–73** (25 μ M) at 37 °C for 5, 30, or 60 min. The cells were plated on Greiner CELLviewTM petri dish (35 \times 10 mm) and the fluorescent intensity of Rhod-2 or Rhod-4 was observed by confocal fluorescent microscopy (Fluoview, FV-1000, Olympus). Excitation was 559 nm for Rhod-2 and Rhod-4.

Flow Cytometric Analysis of Jurkat Cells Stained with Rhod-2 or Rhod-4

Jurkat cells $(2.0\times10^5 \text{ cells})$ were preincubaed with Rhod-2/AM (5 μ M) or Rhod-4/AM (5 μ M) in RPMI 1640 medium containing 10% FBS (100 μ L) under 5% CO₂ at 37 °C for 30 min, centrifuged (2000rpm at 4° C for 3 min) and wash with PBS. The collected cells were incubated in RPMI 1640 (10% FBS) in the presence of $71-73$ (25 μ M) for a given incubation time (0, 5, 10, 20, 30, 40, 50, and 60 min), instantly after which the cells were suspended in 300 μ L RPMI 1640 medium and then analyzed on flow cytometer (FACSCalibur cytometer, Becton), and the data were analyzed on FlowJo software (FlowJo, LCC).

Fluorescent Confocal Microscopic Studies of Jurkat Cells Treated with LysoTracker Red, MitoTracker Red, and ER-Tracker Red in the Presence of Ir Complexes 71–73

Jurkat cells $(2.0 \times 10^5 \text{ cells})$ were incubated with **71–73** (25 µM) in RPMI 1640 medium containing 10% FBS (100 μ L) at 37 °C under 5% CO₂ for 30 min or 1 h, and then stained with LysoTracker Red (500 nM, 100 μ L), MitoTracker Red (100 nM, 100 μ L), or ER-Tracker Red (500 nM, 100 μ L) in PBS for 1 h. After the incubation, the cells were washed with ice-cold PBS containing 0.1 % NaN₃ and 0.5% FBS and observed by confocal fluorescent microscopy (Fluoview, FV-1000, Olympus) using a Greiner CELLviewTM petri dish (35 \times 10 mm). Excitation was 405 nm for Ir complexes, 559 nm for LysoTracker Red, MitoTracker Red and ER-Tracker Red. Emission from 470–530 nm for **71–73**, 580–660 nm for LysoTracker Red, MitoTracker Red, and ER-Tracker Red was used.

Methylene Blue Staining and Transmission Electron Microscopy (TEM) of Jurkat cells Treated with 71 and 72

Jurkat cells $(3 \times 10^6 \text{ cells})$ were treated with **71** or **72** (25 µM) in RPMI 1640 medium with 10% FBS (100 μL) and incubated at 37 °C under 5% CO₂ for 1 h. The cells were fixed with 2.5% glutaraldehyde at 4 °C for 40 min and washed twice with ice-cold PBS. Post-fixation was conducted with 1% osmium tetroxide and the cells were then included in an agarose gel, which was dehydrated by treatment with 50-100% anhydrous EtOH. The cells were embedded in epoxy resin (Poly 812, Nisshin EM Co. Ltd.) at 60 \degree C for 3 days. The resin was sliced with a glass knife (150 nm thickness) and stained with methylene blue for microscopic observation on a microscope (BX51, Olympus). For TEM observation, the sliced samples (100 nm thickness) were stained with EM stainer (Nisshin EM Co. Ltd.) and observed by TEM instrument (H-7650, HITACHI) with electron irradiation at 100 kV.

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- (43) The ¹H NMR signals of **34a** and **34b** in the lowest magnetic field were assigned to be hydrogen atoms on carbon next to the nitrogen (Figure 2-4). The left peaks of **34b** (at δ $= ca. 9.5$) exhibits downfield shift than **34a** (at $\delta = ca. 9.2$), inidicating that the shielding effect of bromine atom is weaker than that of chlorine atom.
- (44) It should be mentioned that the decomposition of *fac***-20** requires longer reaction time than that of *mer***-20** (Entry 1 vs. Entry 2 in Table 2-6), possibly because *fac***-20** is more stable by *ca*. 26 kJ/mol than *mer***-20**, as calculated by DFT calculation (Difference in total energies of *fac***-20** and *mer***-20** is 0.01 a.u., which corresponds to *ca*. 26 kJ/mol, as indicated in Table 2-7).
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- (69) Jurkat cells were fixed using glutaraldehyde and osmium tetroxide (OsO4) after the treatment with 68 and 69 ($25 \mu M$ for 1 h) and observed on microscopy. Vacuolization of cytoplasm and morphological alternations in the cell membrane were observed, which show good agreement with Figure 4-6 in the text.
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